P65 inactivation in adipocytes and macrophages attenuates adipose inflammatory response in lean but not in obese mice

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P65 inactivation in adipocytes and macrophages attenuates adipose inflammatory response in lean but not in obese mice. Am J Physiol Endocrinol Metab 308: E496–E505, 2015. First published January 6, 2015; doi:10.1152/ajpendo.00532.2014.—NF-κB induces transcriptional expression of proinflammatory genes and antiapoptotic genes. The two activities of NF-κB remain to be characterized in the mechanism of chronic inflammation in obesity. To address this issue, we inactivated NF-κB in adipose tissue by knocking out p65 (RelA) in mice (F-p65-KO) and examined the inflammation in lean and obese conditions. In the lean condition, KO mice exhibited a reduced inflammation in adipose tissue with a decrease in macrophage infiltration, M1 polarization, and proinflammatory cytokine expression. In the obese condition, KO mice had elevated inflammation with more macrophage infiltration, M1 polarization, and cytokine expression. In the mechanism of enhanced inflammation, adipocytes and macrophages exhibited an increase in cellular apoptosis, which was observed with more formation of crown-like structures (CLS) in fat tissue of KO mice. Body weight, glucose metabolism, and insulin sensitivity were not significantly altered in KO mice under the lean and obese conditions. A modest but significant reduction in body fat mass was observed in KO mice on HFD with an elevation in energy expenditure. The data suggest that in the control of adipose inflammation, NF-κB exhibits different activities in the lean vs. obese condition. NF-κB is required for expression of proinflammatory genes in the lean but not in the obese condition. NF-κB is required for inhibition of apoptosis in the obese condition, in which proinflammation is enhanced by NF-κB inactivation.

inflammation; crown-like structure; macrophage polarization; apoptosis; nuclear factor-κB

ADIPOCYTES AND MACROPHAGES are the major sources of proinflammatory cytokines in adipose tissue (49, 54). They are activated by hypoxia (20, 56), oxidative stress (11, 34), endoplasmic reticulum (ER) stress (35), Toll-like receptor 4 activation (25), cell death (7), etc., in the inflammatory response in obesity. It is generally believed that activation of the IKKB/NF-κB signaling pathway in those cells occurs in obesity to promote the inflammatory response (42). In agreement, gain-of-function of NF-κB in adipose tissue promotes chronic inflammation in mice with overexpression of p65 subunit (47) or IKKB (21). Inhibition of the pathway is proposed as an approach in the control of obesity-associated inflammation for improvement of insulin resistance (8). However, the view remains to be tested by loss-of-function of NF-κB in adipose tissue. Of the two subunits (p65 and p50) of NF-κB, the p65 subunit contains the activation domain and determines the transcriptional activity of NF-κB (4). Knockout of p65 gene is an approach to inactivate NF-κB activity in vivo. To address this issue, we made fat-selective p65-knockout (F-p65-KO) mice and examined the inflammatory status of adipose tissue in this study.

NF-κB regulates gene transcription through direct binding to the target gene promoter DNA. NF-κB induces expression of proinflammatory cytokines [such as TNFα, IL-1, IL-6, monocyte chemotactic protein-1 (MCP-1), etc.] in response to stress signals (4). However, NF-κB also controls cell survival by induction of antiapoptotic genes (22). NF-κB induces expression of multiple genes to block the apoptotic programs. The genes include those that inhibit caspase function, such as the cellular inhibitor of apoptosis proteins, X-chromosome-linked IAP (XIAP), and caspase-8-c-FLIP (FLICE inhibitory protein) (46); 2 inhibit the apoptotic signaling downstream of TNF receptor 1 (TNF-R1), such as TNF-R-associated factors 1 and 2 (22); 3 preserve mitochondrial function, such as Bel-1xl and A1/Bfl1 (22); and 4 inhibit proapoptotic signaling by the c-Jun kinase (JNK) pathway, such as XIAP and Gadd45β (37, 46, 60). These genes eventually block activation of the caspase pathways to control apoptosis. Although inhibition of NF-κB may reduce expression of proinflammatory cytokines, it may also increase the risk of apoptosis. Adipocyte apoptosis is a risk factor of chronic inflammation in adipose tissue (1, 7, 52). The relative significance of two possibilities remains to be determined in obesity after NF-κB inhibition.

To test the NF-κB activities in adipose tissue, we inactivated NF-κB activity by knocking out the p65 gene in adipocytes and macrophages using the FABP4-Cre (fatty acid-binding protein 4 Cre recombinase, aP2-Cre) system. The knockout was a consequence of exon 2–4 deletion in p65 gene. The results suggest that NF-κB inactivation promotes adipose tissue inflammation in obesity through induction of cellular apoptosis.

EXPERIMENTAL PROCEDURES

Generation of floxed-p65 mice. The loxP p65 mice were made by inserting loxP sites in introns between exons 1 and 5 of the p65 gene, which was designed to remove exons 2–4 after Cre-mediated recombination to inactivate the p65 gene (Fig. 1). FABP4-Cre mice on the C57BL/6 genetic background (aP2-Cre mice, stock no. 005069) were...
Food intake was determined manually in individually housed mice on chow diet at 16 wk of age. The average daily food intake was determined over 3 days by the net reduction in diet weight with exclusion of spilled diet. Food intake in grams was converted to kilocalories and adjusted by lean body mass. Food intake on HFD was measured every 2 wk. Body composition was measured using quantitative nuclear magnetic resonance (NMR; Minispec Mn10 NMR scanner; Brucker Canada, Milton, ON, Canada), as described previously (47). Antibodies to NF-κB p65, Cre, and IκBα were determined by Western blot. mRNAs of the NF-κB target genes were determined by quantitative (q)RT-PCR. In the bars, values are means ± SE (n = 8). *P < 0.05 by Student’s t-test. WT, wild type; KO, knockout; MCP-1, monocyte chemotactic protein-1.

Energy expenditure. Indirect calorimetry was performed in mice at 6 wk on HFD within the Comprehensive Laboratory Animal Monitoring System (Columbus Instruments, Columbus, OH). Mice were kept in the metabolic chamber for 6 days. Oxygen consumption (V\textsubscript{O2}), carbon dioxide production (V\textsubscript{CO2}), spontaneous physical activity, and food intake were recorded daily. Energy expenditure (EE; kcal·kg\textsuperscript{-1}·day\textsuperscript{-1}) was calculated with data on day 5 using the formula EE = [3.815 + 1.232 × V\textsubscript{CO2}/V\textsubscript{O2}] × V\textsubscript{O2} × 0.001 (19). EE data were normalized with body lean mass, and the average daily food intake was derived over 3 days (days 4 – 6).

Insulin tolerance test and glucose tolerance test. Insulin tolerance test (ITT) was conducted with peritoneal injection of insulin (0.75 U/kg) after 4-h fasting in mice at 9 wk on HFD. Glucose tolerance test (GTT) was performed with peritoneal injection of glucose (2 g/kg) after overnight fasting in mice at 10 wk on HFD. Blood glucose was monitored in the tail vein blood using the FreeStyle OneTouch blood glucose-monitoring system (TheraSense, Phoenix, AZ).

Western blot. Whole cell lysates were prepared from epididymal white adipose tissue and used in Western blotting according to the protocols described elsewhere (47). Antibodies to NF-κB p65 (sc-8008), IκBα (sc-371), phospho-c-JUN (sc-822), and c-JUN (sc-1694) were purchased from Santa Cruz Biotechnology (Santa Cruz, CA). Antibodies to β-actin (ab6276), Cre (ab24608), and tubulin (ab7291) were obtained from Abcam (Cambridge, UK). Antibodies to cleaved caspase 7 (no. 8438) and caspase 7 (no. 12827) were obtained from Cell Signaling Technologies (Danvers, MA). Tubulin and β-actin were used as internal controls.

Quantitative real time RT-PCR. Total mRNA was extracted using TRIzol reagent, following the manufacturer’s protocol (Invitrogen, Carlsbad, CA). The ABI 7900 platform with TaqMan Universal PCR Master Mix (4304437; Applied Biosystems, Carlsbad, CA) was used to quantify mRNA in total RNA extracts. Target gene expression was normalized to ribosomal 18S RNA as the endogenous control. Primers and probes were purchased from Applied Biosystems (Carlsbad, CA).
These included F4/80 (Mm00802530-m1), integrin aM (CD11b; Mm00434455-m1), integrin aX (CD11c; Mm00498698-m1), arginase 1 (Mm00475988-m1), sterol regulatory element-binding protein (SREBP; Mm0550338-m1), PPARγ (Mm00440945_m1), fatty acid synthase (FAS; Mm00662319-m1), hormone-sensitive lipase (HSL; Mm00499539-m1), lipoprotein lipase (LPL; Mm00484770-m1), p65 (Mm00501346_m1), adiponectin (ACDC; Mm00456425_m1), aP2 (Mm00445880_m1), leptin (Mm00434759_m1), IL-1β (Mm0043228_m1), IL-6 (Mm00446190_m1), TNFα (Mm00443258_m1), MCP-1 (Mm00441242_m1), and IkBα (Mm00477798_m1). For adipokine expression, macrophage markers, lipogenic genes, inflammatory cytokines, and angiogenic genes, RNA was extracted from epididymal white adipose tissue (WAT) at 8 wk of age from mice on chow diet and 26 wk of age from mice on HFD.

Apoptosis in adipose tissue. After 18 wk of HFD feeding, epididymal WAT was isolated from mice. In the assay, formalin-fixed epididymal WAT was processed, embedded in paraffin, and sectioned at 5 μm. Viable cells were blotted with a periplerin antibody (ab3526; Abcam, Cambridge, MA) at 1:200 and a FITC-conjugated secondary antibody (F2765; Invitrogen) at 1:400. Apoptotic cells were detected using the In situ Cell Death Detection Kit (TUNEL assay) according to the manufacturer’s protocol (12156792910; Roche Applied Science, Indianapolis, IN). Cleaved caspase 3 was determined in the immunohistostaining using a primary antibody (no. 9661; Cell Signaling Technology, Danvers, MA). Cleaved caspase 7 was determined in the tissue in Western blot to facilitate the apoptosis assay. Fluorescent images were captured with an upright microscope (Zeiss Axioplan 2, Intelligent Imaging Innovations, Denver, CO) and a confocal microscope (Leica TCS SP5; Leica Microsystems, Wetzlar, Germany) equipped with an AOBS system and tandem scanner (resonant scanner). Slidebook Software version 2.0 (Intelligent Imaging Innovations) and Leica TCS SP5 LASAF software (Leica Microsystems) were used in image analysis.

Apoptosis in vitro. Primary adipocytes and macrophages were treated with TNFα (10 ng/ml) for different times, as indicated. Apoptosis was determined in the cells by measuring cleaved caspase 7 in Western blot. Adipocytes were differentiated from preadipocytes of inguinal fat, as described elsewhere (47). Macrophages were prepared as described below.

Crown-like structure. Epididymal WAT was isolated from mice at 16 wk on HFD. Hematoxylin and eosin (H & E) staining of tissue sections and fluorescent staining of living tissue were performed as described elsewhere (53). In the fluorescent staining, griffonia simplificofolia IB4 isoclitin Alexa Fluor 488 (40 μg/ml in PBS) was used to stain macrophages and endothelial cells. BODIPY 558/568 (5 μM in PBS) was used in the staining of adipocytes. Images were captured using the confocal microscope (Leica Microsystems).

Triglyceride analysis. Triglyceride content in the feces was measured in mice after 8 wk on chow diet or 16 wk of HFD feeding.

Macrophage function. Primary peritoneal macrophages were isolated from mice 48–72 h after intra peritoneal injection of a sterilized starch solution (2% wt/wt, 2 ml; Sigma 85643), as described elsewhere (47). LPS (2 ng/ml; Sigma) was used to induce cytokine expression in serum-free RPMI 1640 medium. Total RNA was extracted from cells, and mRNA expression was determined for IL-1β, IL-6, and TNFα.

Serum cytokine. Serum was collected through retro-ocular bleeding at 18 wk on HFD. The collection was made after overnight fasting. Cytokines IL-1β, IL-6, and TNFα were tested using a multiplex kit (MCYTOMAG-70K; Millipore).

Statistical analysis. Statistical analysis was performed using two-tailed, unpaired Student’s t-test for the WT littermates vs. KO mice. P < 0.05 was considered significant. Results are presented as means ± SE.

RESULTS

Inflammation in F-p65-KO mice on chow diet. Our earlier study suggested that activation of NF-κB in adipose tissue leads to increased chronic inflammation in aP2-p65 mice (47), which suggests that inhibition of NF-κB may attenuate the adipose inflammation. To test this possibility, we inactivated the p65 gene to suppress NF-κB activity in adipocytes using the Cre-LoxP strategy. Floxed-p65 mice were generated by flanking the exons 2–4 with a pair of LoxP sites, and the flox result was confirmed by PCR genotyping (Fig. 1, A and B). The selection marker Neo (neomycin resistance gene) was removed from the p65 gene in floxed-p65 mice. Fat-selective inactivation was made by crossing the floxed-p65 mice with aP2-Cre mice. In the homozygous F-p65-KO mice (p65−/−, aP2-Cre+/−), P65 protein was reduced by 90% in the fat tissue relative to the wild-type (WT) littermates of floxed p65 (p65+/−), expression of NF-κB target genes was also decreased in the fat tissue, as indicated by IkBα (protein and mRNA), TNFα, IL-6, IL-1β, and MCP-1 (Fig. 1D). Expression of Cre protein was detected in the adipose tissue of KO mice (Fig. 1C). The results suggest that NF-κB is inactivated in the adipose tissue of F-p65-KO mice. The inactivation reduced expression of pro-inflammatory cytokines.

Metabolic phenotype was examined in F-p65-KO mice on chow diet. The data were collected on body weight, body fat mass, lean body mass, food intake, nutrient absorption, energy expenditure, physical activity, substrate utilization, insulin tolerance, and glucose tolerance (data not shown). These data demonstrate that KO mice exhibit no difference from WT mice in energy balance and insulin sensitivity, suggesting that NF-κB inactivation has no impact on metabolism in the lean mice.

Macrophage in KO mice on chow diet. Macrophage infiltration is a marker of chronic inflammation in the white adipose tissue during obesity (49, 54). The aP2 promoter exhibits an activity in macrophages (47, 50). Macrophages were examined in F-p65-KO mice on the chow diet. P65 protein was decreased, and expression of NF-κB target genes (IL-1, IL-6, and TNFα) was reduced in macrophages of KO mice (Fig. 2, A–D). The reduction was especially significant in response to LPS (Fig. 2, B–D). Macrophage infiltration and polarization were examined in epididymal fat. Compared with WT mice (floxed-p65 mice), KO mice exhibited a significant reduction in macrophage markers (F4/80, CD11b, and CD11c) (Fig. 2E). The reduction in CD11c (not detectable) suggests a decrease in type 1 macrophages (30) in KO mice. Expression of arginase 1 (a marker of type 2 macrophages) was enhanced (Fig. 2E), suggesting more type 2 macrophages in KO mice. The data suggest that macrophage infiltration, type 1 polarization, and expression of pro-inflammatory mediators are decreased in adipose tissue of F-p65-KO mice.

Reduced fat content in F-p65-KO mice on HFD. The data in lean mice suggest that p65 inactivation may reduce adipose inflammation in the obese condition. To test this possibility, KO mice were fed HFD (58% calories in fat) for 18 wk to induce obesity. Compared with WT mice, KO mice gained a little less weight on HFD. However, the difference was not significant in the 18-wk study (14-wk data are presented in Fig. 3A). Despite the lack of change in body weight, body fat mass was significantly reduced in KO mice, and the lean body mass
was not altered (Fig. 3, B and C), suggesting that the modest body weight reduction is from fat decrease. Food intake was not different between the two groups (Fig. 3D). However, EE was slightly elevated in KO mice compared with WT (Fig. 3E). The increase was observed in both day and night times. The physical activity, insulin, and glucose tolerance were not significantly different between KO and WT mice (Fig. 3, F–H). ITT and GTT were tested at 9–10 and 16–18 wk in KO mice on HFD. The data are consistent at the two time points, and the data for 9–10 wk are presented. The data suggest that NF-κB inactivation may modestly reduce fat mass through increasing EE without a significant impact on lean body mass and insulin sensitivity.

**Enhanced inflammation in fat of F-p65-KO mice on HFD.** Inflammatory status was determined in the epididymal fat of mice at 8 wk of age on chow diet (n = 8). *P < 0.05, **P < 0.001 by Student’s t-test. CNT, control.

**Cell apoptosis in fat tissue of KO mice.** It was surprising that the inflammation was enhanced in KO mice on HFD. To understand the mechanism, we examined cell apoptosis in the fat tissue of F-p65-KO mice on chow and HFD. NF-κB controls cellular apoptosis, but the activity remains unknown in adipose tissue. Adipocyte death contributes to the adipose inflammation in obesity and is characterized by “crown-like structure” (CLS) in adipose tissue (7). Viable cells and apoptotic cells were determined with perilipin (viable) and TUNEL (apoptotic) assays in the adipose tissue. DAPI was used to determine cell density for nuclear DNA staining. On chow diet, KO mice exhibited no difference from WT mice in viable and apoptotic cells (chow; Fig. 5A). On HFD, KO mice had less viable cells and more apoptotic cells (HFD; Fig. 5A). Reduced viable cells were observed by decreased perilipin signal, and increased apoptotic cells were observed with enhanced TUNEL signal in KO mice (Fig. 5A). The apoptosis was confirmed with caspase assays. Cleavage of caspase 7 was increased in KO mice (Fig. 5B). In vitro, adipocytes and macrophages of KO mice exhibited more cleaved caspase 7 in response to TNFα stimulation (Fig. 5, C and D). The data suggest that NF-κB inactivation increased apoptosis in adipocytes and macrophages in adipose tissue of F-p65-KO mice under obesity.

**Increased CLS in KO mice.** CLS are formed by accumulated macrophages around a dead adipocyte (7). An increase in CLS was observed in KO mice with hematoxylin and eosin staining, in which macrophages were in heavy purple color in the extracellular matrix of adipose tissue (Fig. 6A, arrows). CLS was further examined by analysis of macrophages and adipocytes using fluorescent staining, in which macrophages were stained by isolectin (green) and adipocytes were stained with perilipin (violet) (Fig. 6A, arrows). CLS are formed by accumulated macrophages around a dead adipocyte (7).

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**Regulation of adipose inflammation by NF-κB.** A: p65 protein in macrophages of KO mice. Western blot was conducted with cell lysate of peritoneal macrophages. B: IL-1. mRNA of NF-κB target gene was expressed in fold change in peritoneal macrophages. LPS (2 ng/ml) was used to induce the target gene expression in macrophages. C: IL-6. D: TNFα. E: macrophages in adipose tissue. mRNA was determined in epididymal fat of mice at 8 wk of age on chow diet (n = 8). *P < 0.05, **P < 0.001 by Student’s t-test. CNT, control.
Bodipy (red) (Fig. 6B). Macrophages were located in groups as indicated by green dots in the extracellular matrix (Fig. 6B, white arrows). Adipocytes were in red color from Bodipy staining. Apoptotic cells in CLS were observed by double staining with TUNEL signal (red) and cleaved caspase 3 (green) (Fig. 6, C and D). The apoptotic adipocytes are in orange color due to colocalization of the red and green signals (Fig. 6, C and D). Apoptotic macrophages are small cells double stained by red and green colors in the extracellular metrics (Fig. 6D). The data suggest that, in obesity, NF-κB...
inactivation leads to an increase in CLS in adipose tissue of F-p65-KO mice through adipocyte and macrophage apoptosis.

Enhanced JNK activity. NF-κB inhibits JNK (c-Jun NH2-terminal kinase) activity through induction of gene expression. The increased apoptosis may be a result of enhanced JNK activity, which is known to induce cellular apoptosis in NF-κB-deficient cells. To test this possibility, JNK activity was determined with the phosphorylation status of c-Jun in the adipose tissue of KO mice on HFD. The phosphorylation was significantly enhanced in Western blots of KO mice (Fig. 7), suggesting enhanced JNK activity in KO mice.

DISCUSSION

This study provides evidence that NF-κB may control adipose tissue inflammation through apoptosis. It is generally believed that NF-κB promotes inflammation through induction of transcriptional expression of proinflammatory cytokines. This activity is supported by the reduced adipose inflammation of F-p65-KO mice on chow diet in this study and increased adipose inflammation in mice with p65 or IKK overexpression in other studies (21, 47). However, p65 inactivation did not inhibit the inflammation in F-p65-KO mice on HFD, suggesting that NF-κB is not necessary for chronic inflammation in the obese condition. Apoptosis was enhanced in adipocytes and macrophages in adipose tissue of F-p65-KO mice on HFD but not on chow diet, suggesting that NF-κB is required for the inhibition of apoptosis in the obese condition. The association of increased inflammation and apoptosis suggests that the antiapoptotic activity of NF-κB is more important in the control of inflammation in the obese condition. The apoptosis may block expression of cytokine proteins from the elevated mRNA in macrophages. As such, plasma cytokines were not upregulated, and adiponectin expression was not downregulated in KO mice relative to WT mice.

Apoptosis is a mechanism of removing aging or damaged adipocytes in adipose tissue, which is elevated in obese mice and patients (5, 7). Average lifespan of adipocytes is ~10 yr in humans (3). The adipocyte lifespan is reduced in obesity with acceleration in adipocyte turnover rate (43, 44). Adipocytes have less activity in apoptosis than preadipocytes, as indicated by expression of apoptotic genes. Adipocyte differentiation reduces expression of proapoptotic molecule (TRAIL-R) (23) and increases expression of antiapoptotic molecule (bcl-2) (33). Although adipocytes are more tolerant of apoptosis, adipocyte apoptosis is elevated by obesity, with increased expression of proapoptotic molecule FAS (CD95) (5, 52). The expression may mediate signals of inflammatory cytokines (38) or hypoxia (58) in the induction of apoptosis. Strong apoptosis reduces adipose tissue mass through decreasing adipocyte number, which is reported in transgenic mice with caspase 8 overexpression (36). Adipocyte apoptosis contributes to adipose inflammation in obesity (5, 7). Reduction of adipocyte apoptosis leads to a decrease in adipose inflammation in obese mice with Bid or FAS inactivation (1, 52). The antiapoptosis
activity of NF-κB has never been tested in the context of adipose inflammation. The current study provides evidence that apoptosis from NF-κB inactivation promotes adipose inflammation in F-p65-KO mice on HFD. The apoptosis was not altered in the KO mice on chow diet, which is likely a result of less apoptotic stimuli in the lean condition. The data suggest that NF-κB has an anti-inflammatory effect on the adipose tissue in obesity through inhibition of cell apoptosis.

The reduced adipose tissue mass is related to the adipocyte apoptosis and energy expenditure without an alteration in food intake in F-p65-KO mice. The apoptosis modestly reduced fat mass, which was not strong enough to make a significant reduction in the body weight. An increase in adipogenesis may compensate for the apoptosis to avoid a huge fat reduction in F-p65-KO mice. The increased CLS suggests more adipogenic activities. CLS serves as an adipogenic niche in the fat tissue (27). NF-κB inhibition by IκBα overexpression in adipocytes does not significantly change body weight in mice (50). The energy expenditure was elevated in F-p65-KO mice, which may contribute to the reduced fat mass. The food intake was not significantly altered in KO mice. The energy expenditure is likely a result of elevated inflammation in the fat tissues, which promotes lipolysis and fat mobilization.

Our data suggest that NF-κB may regulate macrophage polarization in adipose tissue. Differentiation of M2 to M1 macrophages (macrophage polarization) is increased in adipose tissue in obesity (30). In F-p65-KO mice, the decreased M1 in the lean and increased M1 in the obese conditions suggest that the NF-κB activity is dependent on fat expansion. The increased M1 is likely a result of macrophage uptake of adipocyte debris from apoptosis in the obese condition. The engulf activity may promote macrophage apoptosis in F-p65-KO mice. Our result is different from myeloid IKKβ-knockout mice, in which proinflammation is reduced in obese mice (2). The difference in IKKβ vs. NF-κB and aP2-Cre vs. Lys-Cre may contribute to the discrepancy as well.

Activation of JNK may contribute to the enhanced inflammatory response in F-p65-KO mice in the obese condition. JNK induces expression of inflammatory genes through activation of transcription factor AP-1. JNK is activated in adipose tissue to promote chronic inflammation in obesity (18). The JNK activity is supported by fat-selective knockout using the aP2-Cre system (40) and myeloid cell-selective knockout using...
the Lyz-Cre system (15). Alternatively, JNK induces cellular apoptosis to promote inflammation. JNK activity was enhanced in the fat tissue of KO mice on HFD, suggesting a role for JNK in promotion of inflammation and cellular apoptosis.

Chronic inflammation of adipose tissue is disassociated with systemic insulin resistance in F-p65-KO mice on HFD. Although adipose inflammation was elevated in KO mice, insulin sensitivity was not altered by the inflammation. ITT and GTT were not impaired in KO mice on HFD, as suggested by data at two time points (9–10 and 16–18 wk). Energy expenditure was elevated in F-p65-KO mice. The increased energy expenditure may be a mechanism for disassociation of inflammation and insulin sensitivity. Energy expenditure improves insulin sensitivity in many mouse models, including the inflammation model (21, 41, 47, 55). Energy expenditure is a positive impact of inflammation on metabolism (57). The positive effect provides a potential answer to the unaltered insulin sensitivity in our KO mice. It also explains the disappointing results of anti-inflammatory therapies in clinical trials for insulin sensitization. Various anti-inflammatory medicines have been tested to improve insulin sensitivity in human studies, in which the medicines yielded negative results in most small clinical trials (8, 57). In the best case, salsalate yielded positive results in the small clinical trials (10, 14) but gave a negative result in a large clinical trial (12, 13). The anti-inflammatory therapies induce body weight in patients in the large clinical trial with 1-kg gain on average (12, 13). The mechanism of weight gain is unknown, but a reduction in energy expenditure may play a role. In animals, salsalate does not improve insulin sensitivity in lipodystrophic mice (17). In current study, the increased energy expenditure may protect insulin sensitivity from the detrimental effects of elevated inflammation in our KO mice.

In the present study, the conditional p65 gene knockout was made with aP2-Cre mice from The Jackson laboratory (stock no. 005069) (16). This aP2-Cre system exhibited recombination efficiency in adipocytes and macrophages in gene deletion. The observation is consistent with aP2 promoter activity in both adipocytes and macrophages in many studies (6, 24, 31, 47, 50, 52). The macrophage activity provides an answer to the aP2-promoter activity in multiple tissues such as brain, liver, and muscle in embryo or adult mice (26, 32, 59). However, in some studies, the macrophage activity was not observed in transgenic mice (28, 29). The aP2-promote does not exhibit leakage in myeloid cells (52). The reason for this discrepancy remains unknown and may be related to floxed genes. Although the aP2-Cre system is limited in specificity to adipocytes, the system remains a powerful tool in the study of adipocytes and macrophages together, according to recent reports (28, 50).

Caution is recommended in the data interpretation for systemic insulin resistance, aP2-Cre system is limited in specificity to adipocytes, the unknown and may be related to floxed genes. Although the reason for this discrepancy remains unknown, and may be related to floxed genes. Although the aP2-Cre system is limited in specificity to adipocytes, the system remains a powerful tool in the study of adipocytes and macrophages together, according to recent reports (28, 50).

In summary, this study suggests that NF-κB regulates proinflammation in adipose tissue through different mechanisms according to the adiposity conditions. In the lean condition, NF-κB contributes to inflammation through induction of proinflammatory cytokines. NF-κB inactivation leads to reduced inflammation, as suggested by expression of proinflammatory genes, macrophage infiltration, and M1 polarization in F-p65-KO mice. In the obese condition, NF-κB exhibits an antiinflammation effect through inhibition of apoptosis. NF-κB inactivation leads to elevated inflammation, as suggested by increased expression of proinflammatory genes, macrophages, and CLS in the white adipose tissue. This study suggests that NF-κB may not be an ideal drug target in the control of chronic inflammation in obesity.

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DISCLOSURES

The authors have no conflicts of interest, financial or otherwise, in the publication of this study.

AUTHOR CONTRIBUTIONS


REFERENCES


