Cholesterol-enriched diet disrupts the blood-testis barrier in rabbits

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Submitted 5 September 2014; accepted in final form 17 October 2014

Cholesterol-enriched diet disrupts the blood-testis barrier in rabbits. Am J Physiol Endocrinol Metab 307: E1125–E1130, 2014. First published October 21, 2014; doi:10.1152/ajpendo.00416.2014.—About 15% of heterosexual couples in the USA suffer from infertility issues; male infertility accounts for ~50% of all infertility cases and roughly 50% of male infertility is idiopathic. Increased levels of plasma cholesterol affect spermatogenesis and male fertility negatively, but by unclear mechanisms. Clearly, spermatogenesis occurs in immune-privileged seminiferous tubules that are protected by the blood-testis barrier (BTB), and BTB disruption results in sperm damage and male infertility. Accordingly, using rabbits fed a 2% cholesterol-enriched diet for 2, 4, and 6 wk to raise levels of plasma cholesterol, we tested the hypothesis that elevated levels of plasma cholesterol disrupt the BTB functionally and biochemically. The cholesterol-enriched diet increased lipid deposition dramatically and time-dependently in the seminiferous tubules and disrupted the BTB as evidenced by increased IgG staining within the seminiferous tubules. Total protein levels of the tight-junction proteins ZO-1 and occludin were increased in the seminiferous tubules of rabbits fed the cholesterol-enriched diet, and the distribution patterns of tight-junction proteins were markedly affected, including an increased accumulation of tight-junction proteins in endosomes. Disruption of the integrity of the BTB due to increased plasma levels of cholesterol might play a role in male infertility.

MATERIALS AND METHODS

Rabbits. New Zealand white male rabbits (1.5 to 2 yr old) weighing 3–4 kg were fed either normal chow or normal chow supplemented with 2% cholesterol for 2, 4, or 6 wk (n = 4). At necropsy, animals were perfused with Dulbecco’s phosphate-buffered saline, and testes were dissected, frozen on a liquid nitrogen cooled surface, and stored at −80°C until taken for experimentation. The animal protocol was approved by the University of North Dakota Animal Care and Use Committee, and adhered to the Guide for the Care and Use of Laboratory Animals (NIH publication no. 80-23).

Cholesterol measurement. Total serum levels of total cholesterol were measured in blood collected from rabbit ear veins. Lipid levels were measured by standard techniques with an Olympus AU640 clinical analyzer.

Oil red O staining. Testes were sectioned (thickness 14 μm) using a cryostat (Micron) and were fixed with 10% formalin for 10 min, washed with H2O, and incubated with 60% isopropanol for 5 min. Once dried, sections were stained with Oil red O (Sigma) for 10 min and washed with H2O. Images were acquired using a Leica microscope.

Immunostaining. Cryostat sections (as described above) were stained for target proteins using antibodies against EEA1 (Santa Cruz Biotechnology), rabbit IgG (Invitrogen), ZO-1 (Invitrogen), and occludin (Invitrogen). Double fluorescence staining was used to determine subcellular codistribution of tight-junction proteins with endo-

Man infertility contributes to roughly 50% of all infertility cases, and ~15% of heterosexual couples in the USA experience fertility issues (7, 16). Although varicoceles, obstructions, ejaculatory dysfunction, infections, and hormonal deficiencies are known causes of male infertility, a significant proportion (40–50%) of male infertility is idiopathic (7). Elevated levels of plasma cholesterol in humans can decrease semen quality and contribute to male infertility (10, 14, 17), while high levels in animals can decrease sperm concentration, impair sperm motility, reduce length of sperm midpiece, and lower rates of in vitro fertilization (1, 20, 22, 31). In addition, increased cholesterol accumulation in Sertoli cells reduces testicular function and compromises fertility (15, 18, 21). Thus, it appears clear from human and animal studies that increased plasma cholesterol impairs spermatogenesis and affects male fertility.

Spermatogenesis occurs in seminiferous tubules, an immune-privileged environment that is protected by the blood-testis barrier (BTB). The BTB is a barrier that consists of tight-junction complexes between adjacent Sertoli cells and that divides the seminiferous epithelium into basal and adu-
with horseradish peroxidase for 1 h at room temperature, reacted with a control. Blots were probed with secondary antibodies conjugated against rabbit IgG, ZO-1, and occludin. PVDF membranes, and subjected to immunoblotting with antibodies resolved by SDS-PAGE under reducing conditions, transferred to a nitrocellulose membrane, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occludin, and ZO-2. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were transferred to nitrocellulose membranes, and immunoblotted with primary antibodies against ZO-1, occlude...
In addition, the cholesterol-enriched diet markedly increased the numbers of EEA-1-positive endosomes. The effects of the cholesterol diet on ZO-1 and endosomes were increasingly apparent as the rabbits were maintained on the diet for 4 and 6 wk. Importantly, we demonstrated that the cholesterol-enriched diet time-dependently increased the codistribution of ZO-1 with endosomes (Fig. 3A). Consistent with our findings from immunostaining studies, we demonstrated that the cholesterol-enriched diet not only time-dependently increased total protein levels of ZO-1 but also significantly increased protein levels of ZO-1 in crude endolysosome fractions (Fig. 3B).

To further confirm our findings, we determined the effects of cholesterol-enriched diet on internalization of occludin, an integral membrane tight-junction protein. Similarly to ZO-1, we demonstrated that cholesterol-enriched diet dramatically altered protein levels and distribution patterns of occludin in a time-dependent manner. In control testes, occludin was distributed only at the base of seminiferous tubules, and very little positive staining for endosomes (EEA1) was present. However,
the cholesterol-enriched diet dramatically altered the distribution of occludin immunostaining, with positive punctate staining becoming apparent in the adluminal side of the seminiferous tubules and this punctate staining colocalized with the endosome marker EEA1 (Fig. 4A). For rabbits fed the cholesterol-enriched diet for 2 or 4 wk, the distribution pattern of occludin immunostaining changed dramatically from a linear and barrier-like pattern in controls to a nonlinear and punctate pattern; as in controls, the punctate staining colocalized with EEA1-positive endosomes. For rabbits fed the cholesterol-enriched diet for 6 wk, the punctate staining pattern of occludin persisted, but the linear and barrier-like pattern of occludin reappeared. Consistent with our immunostaining findings, we demonstrated that a cholesterol-enriched diet not only time-dependently increased total protein levels of occludin but also significantly increased protein levels of occludin in crude endolysosome fractions (Fig. 4B).

**DISCUSSION**

The present study tested our hypothesis that elevated levels of plasma cholesterol, as induced by feeding rabbits a diet enriched in cholesterol, would disrupt BTB integrity and disturb expression levels of tight-junction proteins. Rabbits were used for these studies because they are an excellent model for reproductive system research (12) and they exhibit a functional BTB. Rabbits have the additional advantage of being a well-used model for hypercholesterolemia and its pathological consequences including decreased sperm concentration, impaired sperm motility, reduced length of sperm midpiece, and lowers rate of in vitro fertilization (20, 31).

The BTB consists of tight-junction complexes between adjacent Sertoli cells near the base of the seminiferous epithelium (2) in seminiferous tubules where spermatogenesis occurs. Seminiferous tubules are immune privileged, in part because the BTB limits the entry of toxins, large hydrophilic molecules, and immune cells, thus creating a unique nurturing environment for developing germ cells (5, 27). The importance of the BTB in reproductive health is highlighted clearly by findings that BTB dysfunction leads to sperm damage and male infertility (3, 5, 6, 8, 23). At the molecular level, the BTB forms a complex network of tight-junction proteins that are segregated into three major classes: integral membrane proteins, peripheral adaptors and their associated signaling molecules, and cytoskeletal proteins. The cytoplasmic domains of the integral membrane proteins are linked to the actin cytoskeletal network via adaptor proteins. Three types of transmembrane tight-junction proteins exist: junctional adhesion molecules, occludin, and claudins. The major adaptor proteins that connect transmembrane tight-junction proteins to actin cytoskeleton are zona occludens (ZO-1) (2, 27).

The BTB is a very dynamic structure (2) and undergoes cycles of “opening” and “closing” to accommodate migration of spermatocytes from basal to adluminal compartments. BTB integrity must be maintained for developing meiotic and maturing postmeiotic germ cells (4, 19), and disruption of the BTB leads to sperm damage and male infertility (5). Mechanisms underlying this dynamic reconstruction of tight junctions during spermatogenesis include the relatively new concept of endocytic trafficking of tight-junction proteins (30, 32), and disturbances in endocytosis and/or recycling of tight-junction proteins could play critical roles in disruption of BTB integrity under pathological conditions.

Sertoli cells, which form the BTB, are capable of taking up extracellular cholesterol through receptor-mediated endocyto-
sis (9). Therefore, receptor-mediated endocytosis of cholesterol in Sertoli cells would be enhanced under conditions when plasma cholesterol is high, and this leads to cholesterol accumulation in Sertoli cells. Indeed, in rabbits fed a cholesterol-enriched diet we observed a dramatic increase in cholesterol and triglyceride deposition in seminiferous tubules. Here, we examined BTB integrity using a double fluorescence staining method for endogenous IgG as a marker of BTB leakage and ZO-1 as a marker of tight-junction proteins integral to the BTB. We found that the cholesterol-enriched diet dramatically changed the staining pattern of IgG and ZO-1. In testes from control rabbits, ZO-1 was localized to the basal regions of seminiferous tubules, and IgG staining was excluded from the seminiferous tubules; both indicate that the BTB was functionally intact. In contrast, in testes from rabbits fed a cholesterol-enriched diet, more ZO-1 was present in the adluminal region of seminiferous tubules, and increasingly larger amounts IgG staining were present within the seminiferous tubule, indicating that the integrity of BTB was disrupted.

Enhanced cholesterol uptake may affect endocytic trafficking of tight-junction proteins. Thus, we determined the extent to which the cholesterol-enriched diet affected tight-junction protein internalization. We demonstrated that cholesterol-enriched diet time-dependently altered the protein expression levels and distribution patterns of the tight-junction proteins ZO-1 and occludin. In control testes, these tight-junction proteins were observed only at the base of seminiferous tubules, and very little positive staining for endosomes was present. However, in rabbits fed the cholesterol-enriched diet, positive staining for tight-junction proteins became apparent in the adluminal side of seminiferous tubules, and the numbers and sizes of endosomes were markedly increased. Furthermore, in rabbits fed the cholesterol-enriched diet there was a temporally dependent increase in the codistribution of tight-junction proteins with endosomes. Consistent with our findings from immunostaining studies, we also demonstrated that cholesterol-enriched diet significantly increased protein levels of ZO-1 in endolysosome fractions. Thus, our findings suggest that elevated plasma cholesterol promotes endocytic accumulation of tight-junction proteins and thereby leads to a disrupted integrity of the BTB. Somewhat unexpectedly, we demonstrated that cholesterol-enriched diet also dramatically changed the distribution and increased the total protein levels of both ZO-1 and occludin. Such changes might be an adaptive change, because Sertoli cells increasingly synthesize tight-junction proteins and insert more tight-junction proteins in the adluminal side of seminiferous tubules when one attempts to stop leakage at the basal side of the seminiferous tubules. Such a notion is supported by our observation that in animals fed the cholesterol-enriched diet for 6 wk the amount of the IgG accumulated in testes reverted back to levels similar to those of animals fed the diet for 2 wk.

In summary, we demonstrated, in rabbits, that cholesterol-enriched diet promoted cholesterol accumulation within Sertoli cells, increased accumulation of tight-junction proteins in endocytic compartments, and disrupted the integrity BTB. Our findings suggest that elevated plasma levels of cholesterol disrupt the BTB integrity and contribute to male infertility by affecting endocytic trafficking of tight-junction proteins.

Therapeutic interventions to keep the BTB intact might help prevent and/or reverse some male infertility issues.

REFERENCES


