Cholesterol-enriched diet disrupts the blood-testis barrier in rabbits

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Cholesterol-enriched diet disrupts the blood-testis barrier in rabbits. Am J Physiol Endocrinol Metab 307: E1125–E1130, 2014. First published October 21, 2014; doi:10.1152/ajpendo.00416.2014.—About 15% of heterosexual couples in the USA suffer from infertility issues; male infertility accounts for ∼50% of all infertility cases and roughly 50% of male infertility is idiopathic. Increased levels of plasma cholesterol affect spermatogenesis and male fertility negatively, but by unclear mechanisms. Clearly, spermatogenesis occurs in immune-privileged seminiferous tubules that are protected by the blood-testis barrier (BTB), and BTB disruption results in sperm damage and male infertility. Accordingly, using rabbits fed a 2% cholesterol-enriched diet for 2, 4, and 6 wk to raise levels of plasma cholesterol, we tested the hypothesis that elevated levels of plasma cholesterol disrupt the BTB functionally and biochemically. The cholesterol-enriched diet increased lipid deposition dramatically and time-dependently in the seminiferous tubules and disrupted the BTB as evidenced by increased IgG staining within the seminiferous tubules. Total protein levels of the tight-junction proteins ZO-1 and occludin were increased in the seminiferous tubules of rabbits fed the cholesterol-enriched diet, and the distribution patterns of tight-junction proteins were markedly affected, including an increased accumulation of tight-junction proteins in endosomes. Disruption of the integrity of the BTB due to increased plasma levels of cholesterol might play a role in male infertility.

Materials and Methods

MATERIALS AND METHODS

Rabbits. New Zealand white male rabbits (1.5 to 2 yr old) weighing 3–4 kg were fed either normal chow or normal chow supplemented with 2% cholesterol for 2, 4, or 6 wk (n = 4). At necropsy, animals were perfused with Dulbecco’s phosphate-buffered saline, and testes were dissected, frozen on a liquid nitrogen cooled surface, and stored at −80°C until taken for experimentation. The animal protocol was approved by the University of North Dakota Animal Care and Use Committee, and adhered to the Guide for the Care and Use of Laboratory Animals (NIH publication no. 80-23).

Cholesterol measurement. Total serum levels of total cholesterol were measured in blood collected from rabbit ear veins. Lipid levels were measured by standard techniques with an Olympus AU640 clinical analyzer.

Oil red O staining. Testes were sectioned (thickness 14 μm) using a cryostat (Micron) and were fixed with 10% formalin for 10 min, washed with H2O, and incubated with 60% isopropanol for 5 min. Once dried, sections were stained with Oil red O (Sigma) for 10 min and washed with H2O. Images were acquired using a Leica microscope.

Immunostaining. Cryostat sections (as described above) were stained for target proteins using antibodies against EEA1 (Santa Cruz Biotechnology), rabbit IgG (Invitrogen), ZO-1 (Invitrogen), and occludin (Invitrogen). Double fluorescence staining was used to determine subcellular codistribution of tight-junction proteins with endo-

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Male infertility contributes to roughly 50% of all infertility cases, and ∼15% of heterosexual couples in the USA experience fertility issues (7, 16). Although varicoceles, obstructions, ejaculatory dysfunction, infections, and hormonal deficiencies are known causes of male infertility, a significant proportion (40–50%) of male infertility is idiopathic (7). Elevated levels of plasma cholesterol in humans can decrease semen quality (40–50%) of male infertility is idiopathic. Increased levels of plasma cholesterol might play a role in male infertility. Male infertility; blood-testis barrier; tight junctions

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some and with endogenous IgG. Controls for specificity included staining with an isotype-matched irrelevant antibody as a negative control, staining with primary antibodies without fluorescence-conjugated secondary antibodies (background controls), and staining with only secondary antibodies; these controls eliminated autofluorescence in each channel and bleed-through (crossover) between channels.

Immunoblotting. Total cell lysates and crude endolysosome fractions were prepared using an endolysosome isolation kit (Sigma). Testes (0.5 g wet wt) were ground into powder and homogenized in 4 volumes of 1× extraction buffer. Homogenates were centrifuged at 1,000 g for 10 min at 4°C. Supernatants (total cell lysate) were collected and then centrifuged at 20,000 g for 20 min at 4°C. Pellets were resuspended in 0.5 ml of 1× extraction buffer, and this fraction contained a mixture of light mitochondria, endosomes, lysosomes, peroxisomes, and endoplasmic reticulum. Protein concentration was determined by Bradford assay. Equal amounts of protein (10 μg) were resolved by SDS-PAGE under reducing conditions, transferred to PVDF membranes, and subjected to immunoblotting with antibodies against rabbit IgG, ZO-1, and occludin. β-Actin (Abcam) was used as a control. Blots were probed with secondary antibodies conjugated with horseradish peroxidase for 1 h at room temperature, reacted with luminal reagent, exposed, visualized, and analyzed by LabWorks 4.5 software on a UVP Bioimaging System (Upland). Quantification was performed by densitometry, and the results were analyzed and normalized.

Statistical analysis. All data were expressed as means ± SE. Statistical significance was analyzed with one-way ANOVA plus a Tukey post hoc test. *P < 0.05 was considered to be statistically significant.

RESULTS

Cholesterol-enriched diet promoted cholesterol accumulation in seminiferous tubules. Total plasma levels of cholesterol were markedly increased in rabbits ingesting for 2, 4, and 6 wk a diet supplemented with 2% cholesterol (Fig. 1A). Because Sertoli cells take up extracellular cholesterol through receptormediated endocytosis (9), we first determined the effects of cholesterol-enriched diet on cholesterol accumulation in seminiferous tubules. Using an Oil red O staining method, which stains cholesterol and triglycerides, we found that the cholesterol-enriched diet increased lipid droplet accumulation significantly and time-dependently in seminiferous tubules (Fig. 1B).

Cholesterol-enriched diet disrupted the BTB. We determined next whether rabbits fed the cholesterol-enriched diet exhibited functional and morphological evidence of BTB dysfunction. BTB integrity was examined using a double fluorescence staining method for endogenous IgG as a marker of BTB leakage and ZO-1 as a marker of tight-junction proteins integral to the BTB. In testes from control rabbits, ZO-1 was localized at the basal regions of the seminiferous tubule, and IgG staining was excluded from the seminiferous tubule (Fig. 2A), observations indicating that the BTB was functionally intact. However, the staining pattern of ZO-1 and IgG changed dramatically in testes from rabbits fed the cholesterol-enriched diet for 2, 4, and 6 wk; more ZO-1 was present in the adluminal region of the seminiferous tubule, and increasingly larger amounts of IgG staining were present within the seminiferous tubule (Fig. 2A). Consistent with leakage of IgG into seminiferous tubules as shown by immunostaining, we demonstrated that the cholesterol-enriched diet dramatically increased protein levels of IgG in testes (Fig. 2B).

Cholesterol-enriched diet increased accumulation of tight-junction proteins in endocytic compartments. Endocytic trafficking of tight-junction proteins participates in the dynamic regulation of the BTB (24, 28–30, 32). Because elevated levels of plasma cholesterol promote cholesterol accumulation in seminiferous tubules, it might thereby affect endocytic trafficking of tight-junction proteins. Accordingly, we next determined the effects of cholesterol-enriched diet on tight-junction protein internalization using immunostaining and immunoblotting methods. We demonstrated that the cholesterol-enriched diet time-dependently and statistically significantly altered protein levels and distribution patterns of ZO-1 (Fig. 3). In control testes, ZO-1 was distributed only at the base of seminiferous tubules, and very little positive staining for endosomes (EEA1) was present. However, after 2 wk on the cholesterol-enriched diet, ZO-1 staining became apparent in the adluminal side of

Fig. 1. Cholesterol-enriched diet promoted lipid accumulation in seminiferous tubules. A: total plasma levels of cholesterol were markedly increased following feeding rabbits a diet supplemented with 2% cholesterol for 2, 4, and 6 wk (n = 4, *P < 0.05). B: Oil red O staining demonstrated little to no lipid accumulation along or within the seminiferous tubules in testes from control rabbits, but increasing amounts of lipid accumulation were observed in testes from rabbits fed the cholesterol-enriched diet for 2, 4 and 6 wk. Scale bar, 50 μm.
seminiferous tubules. In addition, the cholesterol-enriched diet markedly increased the numbers of EEA-1-positive endosomes. The effects of the cholesterol diet on ZO-1 and endosomes were increasingly apparent as the rabbits were maintained on the diet for 4 and 6 wk. Importantly, we demonstrated that the cholesterol-enriched diet time-dependently increased the codistribution of ZO-1 with endosomes (Fig. 3A). Consistent with our findings from immunostaining studies, we demonstrated that the cholesterol-enriched diet not only time-dependently increased total protein levels of ZO-1 but also significantly increased protein levels of ZO-1 in crude endolysosome fractions (Fig. 3B).

To further confirm our findings, we determined the effects of cholesterol-enriched diet on internalization of occludin, an integral membrane tight-junction protein. Similarly to ZO-1, we demonstrated that cholesterol-enriched diet dramatically altered protein levels and distribution patterns of occludin in a time-dependent manner. In control testes, occludin was distributed only at the base of seminiferous tubules, and very little positive staining for endosomes (EEA1) was present. However,
the cholesterol-enriched diet dramatically altered the distribution of occludin immunostaining, with positive punctate staining becoming apparent in the adluminal side of the seminiferous tubules and this punctate staining colocalized with the endosome marker EEA1 (Fig. 4A). For rabbits fed the cholesterol-enriched diet for 2 or 4 wk, the distribution pattern of occludin immunostaining changed dramatically from a linear and barrier-like pattern in controls to a nonlinear and punctate pattern; as in controls, the punctate staining colocalized with EEA1-positive endosomes. For rabbits fed the cholesterol-enriched diet for 6 wk, the punctate staining pattern of occludin persisted, but the linear and barrier-like pattern of occludin reappeared. Consistent with our immunostaining findings, we demonstrated that a cholesterol-enriched diet not only time-dependently increased total protein levels of occludin but also significantly increased protein levels of occludin in crude endolysosome fractions (Fig. 4B).

DISCUSSION

The present study tested our hypothesis that elevated levels of plasma cholesterol, as induced by feeding rabbits a diet enriched in cholesterol, would disrupt BTB integrity and disturb expression levels of tight-junction proteins. Rabbits were used for these studies because they are an excellent model for reproductive system research (12) and they exhibit a functional BTB. Rabbits have the additional advantage of being a well-used model for hypercholesterolemia and its pathological consequences including decreased sperm concentration, impaired sperm motility, reduced length of sperm midpiece, and lowers rate of in vitro fertilization (20, 31).

The BTB consists of tight-junction complexes between adjacent Sertoli cells near the base of the seminiferous epithelium (2) in seminiferous tubules where spermatogenesis occurs. Seminiferous tubules are immune privileged, in part because the BTB limits the entry of toxins, large hydrophilic molecules, and immune cells, thus creating a unique nurturing environment for developing germ cells (5, 27). The importance of the BTB in reproductive health is highlighted clearly by findings that BTB dysfunction leads to sperm damage and male infertility (3, 5, 6, 8, 23). At the molecular level, the BTB forms a complex network of tight-junction proteins that are segregated into three major classes: integral membrane proteins, peripheral adaptors and their associated signaling molecules, and cytoskeletal proteins. The cytoplasmic domains of the integral membrane proteins are linked to the actin cytoskeletal network via adaptor proteins. Three types of transmembrane tight-junction proteins exist: junctional adhesion molecules, occludin, and claudins. The major adaptor proteins that connect transmembrane tight-junction proteins to actin cytoskeleton are zonula occludens (ZO-1) (2, 27).

The BTB is a very dynamic structure (2) and undergoes cycles of “opening” and “closing” to accommodate migration of spermatocytes from basal to adluminal compartments. BTB integrity must be maintained for developing meiotic and maturing postmeiotic germ cells (4, 19), and disruption of the BTB leads to sperm damage and male infertility (5). Mechanisms underlying this dynamic reconstruction of tight junctions during spermatogenesis include the relatively new concept of endocytic trafficking of tight-junction proteins (30, 32), and disturbances in endocytosis and/or recycling of tight-junction proteins could play critical roles in disruption of BTB integrity under pathological conditions.

Sertoli cells, which form the BTB, are capable of taking up extracellular cholesterol through receptor-mediated endocyto-
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Therapeutic interventions to keep the BTB intact might help prevent and/or reverse some male infertility issues.

REFERENCES


