A short-term diet and exercise intervention ameliorates inflammation and markers of metabolic health in overweight/obese children


1Department of Integrative Biology and Physiology, University of California Los Angeles (UCLA), Los Angeles, California; 2Exercise and Metabolic Disease Laboratory, Translational Sciences Section, School of Nursing, UCLA, Los Angeles, California; 3Department of Medicine, UCLA, Los Angeles, California; 4Department of Internal Medicine, University of Michigan, Ann Arbor, Michigan; and 5Department of Computational Medicine and Bioinformatics, University of Michigan, Ann Arbor, Michigan

A short-term diet and exercise intervention ameliorates inflammation and markers of metabolic health in overweight/obese children. Am J Physiol Endocrinol Metab 303: E542–E550, 2012. First published June 19, 2012; doi:10.1152/ajpendo.00190.2012.—The present study was designed to examine the effects of short-term diet and exercise on markers of metabolic health, serum-stimulated production of inflammatory biomarkers from cultured monocytes and adipocytes, and serum lipomics. Twenty-one overweight/obese children (9 boys and 12 girls, age 13.0 ± 0.5 yr, BMI 33.0 ± 1.8 kg/m²) were placed on a 2-wk ad libitum, high-fiber, low-fat diet and daily exercise regimen. Fasting serum samples were taken pre- and postintervention for determination of cytokines, metabolic risk markers, and lipomics. Monocytes and adipocytes were incubated with pre- and postintervention serum to investigate changes in cytokine secretion. Correlative associations were calculated, followed by hierarchical clustering to determine relationships between fatty acid (FA) species and clinical biomarkers. Despite remaining overweight/obese, interleukin (IL)-6, IL-8, TNFα, PAI-1, resistin, amylin, leptin, insulin, and IL-1ra decreased and adiponectin increased. Culture studies indicated decreases in cytokine secretion of IL-6, TNFα, and IL-1β and adipocyte secretion of IL-6. Lipomic analysis revealed a decrease in total lipids and decreases in saturated FAs and an increase in 18:1/18:0. In general, Pearson’s correlations revealed that inflammatory markers are negatively associated with a cluster of polyunsaturated FAs and positively correlated with several saturated FAs. These results indicate significant modification of multiple indices of metabolic health with short-term rigorous lifestyle modification in overweight/obese children prior to obesity reversal.

THE OBESITY EPIDEMIC IN CHILDREN AND ADOLESCENTS is well recognized and poses a major threat to the health and longevity of American children. Just as obesity in adults is associated with chronic diseases such as cardiovascular disease, type 2 diabetes, metabolic syndrome, and certain forms of cancer (27, 39), some have suggested that the children of today may be the first generation not to outlive their parents due to the premature development of these obesity-related diseases (26). The role that childhood obesity plays in the development of chronic diseases has been well documented. For example, the Pathobiological Determinants of Atherosclerosis in Youth study demonstrated that the extent of coronary artery disease in young males is positively correlated to body mass index (BMI) (21). Weiss et al. (45) reported that, in a large multiethnic group of children, as severity of obesity increased so did metabolic syndrome incidence, reaching 50% in the most obese group. In addition, studies have indicated that obese children tend to remain obese as adults (20). The causes of the obesity epidemic in youth today are multifactorial, and a Westernized high-fat, refined-carbohydrate diet and decreased physical activity play significant roles. One of the links between obesity and the aforementioned comorbidities is chronic low-grade systemic inflammation.

In a pair of seminal studies, we reported that a short-term diet and exercise intervention resulted in significant reductions in serum lipids, several markers of atherosclerosis, and the chronic low-grade inflammatory marker high-sensitivity C-reactive protein (CRP). These differences occur despite small changes in weight, and the subjects remained overweight or obese after the 2-wk intervention (6, 35). Based on the previous observation of a 40% reduction in high-sensitivity CRP, indicating a reduction in chronic low-grade inflammation, the present study was designed to further investigate the effects of daily exercise and an ad libitum plant-based diet on inflammatory cytokines, serum lipomics, and other serum markers of metabolic health. We hypothesized that changes in serum factors, especially a decrease in saturated fatty acids (SFAs), could decrease the production of inflammatory cytokines in cultured adipocytes and monocytes. To test these hypotheses, we first performed serum analyses pre- and postintervention. In addition, adipocytes and monocytes were cultured separately and stimulated with serum obtained before and after the intervention, and levels of cytokines were measured postculture. Finally, given that numerous FA species may influence inflammatory status, we investigated the effects of this intervention on lipomic profiling.

METHODS

Subjects. Twenty-one children, classified as overweight/obese by the Centers for Disease Control (CDC) sex-specific BMI-for-age percentiles, ages 8–17 (mean 13.0 ± 0.5 yr), participated voluntarily in a 2-wk residential lifestyle modification program at the Pritikin Longevity Center in Florida. Other than being overweight/obese, and eight of 21 having the metabolic syndrome, the children did not have any other known medical problems. Pre- and postintervention data were obtained from nine males and 12 females participating in the...
2-wk program. None of the subjects were using drugs or therapies for obesity, and none had prior histories of disease or injury that would prevent daily exercise. Consent to participate in a research program was obtained from the parents, all subjects agreed to provide data for the study, and the project was approved by the University of California Los Angeles (UCLA) Human Subjects Protection Committee.

**Diet and exercise intervention.** Participants in the program received a complete physical examination and underwent a 14-day diet and exercise intervention, as described previously (6, 35). Briefly, prepared meals, which were well tolerated by the subjects, contained 12–15% of calories from fat (polysaturated/saturated FA ratio = 2.4:1), 15–20% of calories from protein, and 65–70% of calories from primarily refined carbohydrate high in dietary fiber (>40 g/day). All foods except for animal-derived protein sources were served ad libitum. The exercise intervention consisted of 2–2.5 h/day of supervised activity. Blood samples were drawn after a 12-h overnight fast on days 1 and 12 of the intervention. The blood was separated by centrifugation, and serum was shipped on dry ice to UCLA, where it was stored at −80°C until analysis. Frozen serum was shipped on dry ice to the University of Michigan for lipomics analysis. Anthropometric data were collected as described previously (40).

**Determination of serum lipids, glucose, insulin, homeostatic model assessment for insulin resistance, and quantitative insulin sensitivity check index.** Total cholesterol, triglyceride (TG), HDL, and glucose levels were measured at a national commercial laboratory (Quest Diagnostics, Miami, FL) using standardized techniques, as described previously (44). LDL was calculated as described by the Friedewald formula (9). Insulin was quantified in duplicate using Luminex xMAP Multiplex (Millipore, Billerica, MA). The degree of insulin resistance was estimated with the use of the homeostatic model assessment for insulin resistance (HOMA-IR) and calculated as the product of the fasting plasma insulin (μU/ml) and the fasting plasma glucose (mmol/l) divided by 22.5. Insulin sensitivity was also estimated by the quantitative insulin sensitivity check index (QUICKI), as defined by 1/[log(fasting insulin (μU/ml) + log(fasting glucose (mg/dl))].

**Determination of serum interleukins, TNFα, adiponectin, plasminogen activator inhibitor-1, resistin, amylin, and leptin.** Serum IL-8, IL-10, IL-1 receptor antagonist (IL-1ra), IL-6, TNFα, plasminogen activator inhibitor-1 (PAI-1), resistin, amylin, and leptin were measured in duplicate using specific Luminex xMAP Multiplex kits (Millipore) according to the manufacturer’s instructions. IL-1β was measured using an enzyme-linked immunosorbent assay (ELISA) kit (minimum detectable dose is 1 pg/ml) but was not detectable in the serum samples (R & D Systems, Minneapolis, MN). Serum adiponectin was also measured using an ELISA kit (R & D Systems).

**Adipocyte cell culture in vitro studies.** Human preadipocytes, isolated from the subcutaneous thigh regions of an obese female, were plated at a density of ~40,000 cells/cm² on a 24-well plate by a commercial adipocyte culture supplier (Zen-Bio, Research Triangle Park, NC) (7). The cells differentiated into spindle-shaped primary adipocytes in ~2 wk, and the plate was vacuum-sealed and shipped to UCLA in FBS- and insulin-free DMEM-Ham’s F-12 culture medium supplemented with HEPES, biotin, pantothenate, dexamethasone, penicillin, streptomycin, and amphotericin B. Upon arrival, excess medium was removed, and adipocytes were incubated at 37°C in a humidified 5% CO2-95% air incubator.

After 1 wk of incubation for stabilization, the cultured adipocytes were washed three times in basal medium, which was comprised of DMEM-Ham’s F-12 medium, HEPES, biotin, and pantothenate (Zen-Bio). Addition of pre- and postintervention serum (10%) in the culture medium was used to investigate adipocyte secretion of IL-6 and monocyte chemoattractant protein-1 (MCP-1) as a result of lifestyle modification. Serum from five subjects (n = 5: 3 males and 2 females) was used. Two-hundred microliters of subject serum was added to each culture well to achieve a total concentration of 10% serum in basal medium. Pre- and postintervention serum samples from each subject were added to wells in duplicate, and four wells remained serum free. After serum addition, cells were incubated for 72 h at 37°C with 5% CO2-95% air.

Cell culture supernatants were collected from the wells by gentle suction and stored at −20°C until analysis of IL-6 and MCP-1 by ELISA (R & D Systems). The immunoassays were performed according to the manufacturer’s instructions for analyzing cell culture supernatants. To calculate adipocyte secretion of IL-6 and MCP-1, the serum concentrations measured previously were subtracted from the measured supernatant levels and adjusted for the 10% dilution.

**Monocyte cell culture in vitro studies.** Peripheral blood was obtained from healthy human subjects, and monocyes were isolated by Ficoll/Hypaque separation and adherence to culture dishes in the presence of 5% (vol/vol) human type AB serum and 15% heat-inactivated fetal calf serum. After the adhered monolayers were washed successively with PBS, a test well was analyzed for monocyte purity (CD14+) via flow cytometry and found to be on average >97% pure. Adhered monocytes were removed from the petri dishes with 4 ml of Versene and resuspended in Iscove’s modified Dulbecco’s medium (DMEM; Irvine Scientific) supplemented with 15% fetal calf serum, 5% human AB serum, 1% glutamine, and antibiotics (penicillin-streptomycin, each at 100 U/ml final volume). The monocytes were plated in a flat-bottomed 96-well plate at 2 × 10⁴ cells/well to provide an even monolayer of monocytes in each well. The 96-well plate was then incubated at 37°C with 5% CO2-95% air for 24 h to allow the cells to attach to the wells.

After 24 h, the medium was carefully removed from each well and replaced with 200 μl of Iscove’s DMEM with 20% subject serum, 1% glutamine, and antibiotics (penicillin-streptomycin, each at 100 U/ml final volume). The pre- and postintervention serum were used to determine the effect of the intervention on monocyte secretion of TNFα, IL-6, and IL-1β and the effect on the proinflammatory JNK pathway in the monocytes. The same subset of subjects from the adipocyte cultures (n = 5: 3 males and 2 females) was used. Pre- and postintervention serum samples from each subject were added to wells in duplicate. After serum addition, cells were incubated for 96 h at 37°C with 5% CO2-95% air.

After incubation, cell culture supernatants were collected from the wells by gentle suction and stored at −80°C until analysis of TNFα, IL-6, and IL-1β by ELISA (R & D Systems). The immunoassays were performed according to the manufacturer’s instructions for analyzing cell culture supernatants. To calculate monocyte secretion of TNFα, IL-6, and IL-1β, the serum concentrations measured previously were subtracted from the measured supernatant levels. Immediately after the supernatants were removed from the wells, the Cellular Activation of Signaling ELISA kit for JNK T183/Y185 (SuperArray, Frederick, MD) was used on the monocytes in the 96-well plate to measure activity of JNK (phosphorylated JNK/total JNK).

**Serum lipids.** Total lipids were extracted from the serum of 16 of the 21 subjects according to the method of Bligh and Dyer (2). Heptadecanoic acid (C17) was added as an internal standard for the quantification of FAs. Lipids were methylated using BF₃-methanol (14% solution from Sigma) and analyzed by gas chromatography on an Omega Wax 250 capillary column (Supelco). Relative abundance of 22 different FA species was done by comparison of retention times with known standards.

**Statistical analysis.** Statistical analyses were performed with GraphPad Prism (GraphPad, San Diego, CA). Preintervention and postintervention values were compared using paired t-tests. Cell culture pre- and postintervention values (n = 5) were compared using matched paired Wilcoxon signed-rank tests for nonparametric data. Associations between lipomics and other measurements were calculated using Pearson’s correlations. Heat maps of correlation matrices, including hierarchical clustering, were generated using the package “gplots” in R (version 2.14.0). All data are expressed as means ± SE unless otherwise noted. A P value of ≤0.05 was considered statistically significant.
RESULTS

Physical characteristics, blood pressure, serum lipids, glucose, and insulin. Anthropometric and metabolic data are summarized in Table 1. The mean BMI before the intervention was 33.0 ± 1.8 kg/m². All subjects had a BMI greater than the 75th percentile, and 18 of the 21 subjects were overweight (>85th percentile) or obese (>95th percentile) according to CDC BMI standards.

Following the intervention, all serum lipids improved significantly, with the exception of HDL, which did not change significantly (Table 1). In addition, both the total cholesterol/HDL and LDL/HDL ratios decreased. Fasting insulin decreased following the program (Table 1). Although blood glucose increased postintervention, both HOMA-IR and QUICKI improved postintervention, driven by the decrease in insulin (Table 1). Prior to the intervention, 8 of the 21 children were classified as having the metabolic syndrome, and at departure, none were classified as having the metabolic syndrome (Table 1).

Effect of the intervention on serum cytokines, adipokines, and endocrine markers. After the diet and exercise intervention, we noted significant decreases in IL-6 (51% decrease), IL-8 (43%), TNFα (40%), and IL-1ra (25%), and the anti-inflammatory cytokine IL-10 did not change (Fig. 1A). Serum PAI-1 (26% decrease), resistin (38%), amylin (36%), and leptin (45%) all decreased, whereas adiponectin increased (39%) (Fig. 1B).

In vitro adipocyte cytokine secretion. The mean level of IL-6 in the supernate was ~4,000 times greater than that in added serum, whereas the supernate MCP-1 was ~100 times the level in added serum. When the calculated amounts of these cytokines were compared between cells treated with preintervention vs. cells exposed to postintervention serum, it was found that secreted IL-6 decreased 32% (Fig. 2). Secreted MCP-1 also decreased nonsignificantly (P = 0.09; Fig. 2).

In vitro monocyte cytokine secretion and JNK activity. The calculation for the secretion of cytokines was performed in the same manner as described for the adipocytes. Secretions of IL-6 (19% decrease), TNFα (58%), and IL-1β (6%) were all reduced in monocytes incubated with postintervention serum compared with monocytes incubated with preintervention serum (Fig. 3). Activation of the JNK pathway, calculated by the ratio of phosphorylated JNK to total JNK, also trended toward reduction in the monocytes with postintervention serum (data not shown), but it was not significant (P = 0.16).

Serum FA profiles. Lipomic analysis of the serum distinguished the types and amounts of FAs in the serum. Generally, the saturated FAs in serum decreased postintervention, and the unsaturated FAs tended to increase from their baseline (BL) levels preintervention. Significant reductions were noted for 14:0 (76% of BL), 18:0 (83% of BL), and 20:0 (62% of BL) (Fig. 4), whereas the 18:1/18:0 ratio (124% of BL), an index of the ratio of unsaturated to saturated FAs, increased significantly (Fig. 4), consistent with an increase in consumption of a lower-fat diet. Other unsaturated FAs, including 20:1 (121% of BL, P = 0.10) and 22:6 (126% of BL, P = 0.09), also increased, but not significantly (Fig. 4).

To assess relationships between changes in clinical and serum parameters, we performed hierarchical clustering of the Pearson’s correlation coefficients among these values at BL and examined these same relationships following the intervention (Fig. 5). At BL, we found a cluster of longer-chain saturated and unsaturated FAs (20:4, 18:2, 24:0, 22:5, 22:4, and total polyunsaturated FAs) with positive correlation with each other and with HDL cholesterol and resting heart rate. This same cluster was negatively correlated with inflammatory markers (IL-6, TNFα, PAI-1, resistin, and amylin), total FA and TG levels, and shorter-chain FAs. In general, these relationships persisted following the intervention, suggesting that these polyunsaturated FAs may suppress inflammation. This cluster of FAs was also negatively correlated with systolic and diastolic blood pressure. Again, this negative correlation was present both before and after intervention. Likely reflecting the change in diet, following the intervention, additional longer-chain polyunsaturated FAs, including α-linolenic acid [18:3 (n-3)], showed a positive correlation with the aforementioned FA cluster.

A series of monounsaturated FAs and the 18:1/18:0 ratio were also negatively correlated with the cluster of saturated FAs and inflammatory markers both before and after the intervention. When we examined the correlation of changes with the various parameters following the intervention (Fig. 6), we noted a significant correlation between the reduction in insulin and HOMA-IR and the increase in 18:1 and 18:1/18:0 ratio, suggesting a relationship between the change in diet and the improvement in a surrogate of insulin sensitivity.

Furthermore, total saturated FAs were positively correlated with proinflammatory markers (IL-6, IL-8, IL-1ra, and TNFα), and total polyunsaturated FAs had negative associations with inflammation (IL-6 and IL-1ra) (all P < 0.05; Fig. 5). In terms of other metabolic markers, the change in amylin was positively correlated with the changes seen in total FAs, IL-6, IL-8, 14:0, 20:0, insulin, HOMA-IR, and TG (all P < 0.05; Fig. 6). Additionally, other than PAI-1, TNFα, and leptin being correlated with BMI, we did not detect any significant correlations between body weight or BMI and serum cytokines, adipokines, or endocrine markers.

Table 1. Anthropometric and lipid measurements in 21 children undergoing a 14-day diet and exercise intervention

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Preintervention</th>
<th>Postintervention</th>
<th>%Change</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight, kg</td>
<td>91.5 ± 6.8</td>
<td>87.9 ± 6.6</td>
<td>-3.9%</td>
</tr>
<tr>
<td>BMI, kg/m²</td>
<td>33.0 ± 1.8</td>
<td>31.7 ± 1.7</td>
<td>-3.8%</td>
</tr>
<tr>
<td>BMI percentile</td>
<td>93.8 ± 1.5</td>
<td>90.6 ± 2.5</td>
<td>-3.4%</td>
</tr>
<tr>
<td>Systolic blood pressure, mmHg</td>
<td>125 ± 4</td>
<td>115 ± 2</td>
<td>-7.8%</td>
</tr>
<tr>
<td>Diastolic blood pressure, mmHg</td>
<td>72 ± 3</td>
<td>68 ± 1</td>
<td>-6.0%</td>
</tr>
<tr>
<td>Blood glucose, mg/dl</td>
<td>82.2 ± 1.9</td>
<td>81.7 ± 1.5</td>
<td>0.9%</td>
</tr>
<tr>
<td>Insulin, μU/ml</td>
<td>21.4 ± 3.3</td>
<td>15.3 ± 3.6</td>
<td>-28.8%</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>4.9 ± 0.8</td>
<td>3.5 ± 0.7</td>
<td>-28.7%</td>
</tr>
<tr>
<td>QUICKI</td>
<td>2.71 ± 0.03</td>
<td>2.85 ± 0.04</td>
<td>5.1%</td>
</tr>
<tr>
<td>Triglycerides, mg/dl</td>
<td>146.5 ± 15.1</td>
<td>99.9 ± 7.7</td>
<td>-38.7%</td>
</tr>
<tr>
<td>Total cholesterol, mg/dl</td>
<td>167.7 ± 5.7</td>
<td>131.8 ± 5.3</td>
<td>-20.9%</td>
</tr>
<tr>
<td>LDL cholesterol, mg/dl</td>
<td>94.3 ± 5.8</td>
<td>71.5 ± 4.6</td>
<td>-24.1%</td>
</tr>
<tr>
<td>HDL cholesterol, mg/dl</td>
<td>43.2 ± 2.1</td>
<td>42.3 ± 2.5</td>
<td>-2.0%</td>
</tr>
<tr>
<td>Total cholesterol/HDL cholesterol</td>
<td>4.09 ± 0.28</td>
<td>3.33 ± 0.38</td>
<td>-18.5%</td>
</tr>
<tr>
<td>LDL cholesterol/HDL cholesterol</td>
<td>2.32 ± 0.21</td>
<td>1.85 ± 0.18</td>
<td>-20.4%</td>
</tr>
<tr>
<td>Subjects with metabolic syndrome</td>
<td>8</td>
<td>0</td>
<td></td>
</tr>
</tbody>
</table>

All data are expressed as means ± SE. BMI, body mass index; HOMA-IR, homeostatic model assessment for insulin resistance; QUICKI, quantitative insulin sensitivity check index. *P < 0.05; ‡P < 0.01.
DISCUSSION

Previous studies in adults (1, 36, 44) and in children (6, 35), using similar diet and exercise interventions as presented here, resulted in significant reductions in serum lipids, insulin, inflammatory markers, oxidative stress, adhesion molecule expression, and small but significant reductions in body weight and BMI. In the present study, we sought to determine the effects of this short-term intensive diet and exercise program in youth on serum inflammatory cytokines and how changes in serum following the intervention affected the ex vivo production of cytokines by cultured adipocytes and monocytes. We also assessed changes in the FA composition of serum lipids and the correlations with changes in anthropometric and serum markers using hierarchal clustering.

The primary findings demonstrate that 1) even a short-term lifestyle modification program with minimal weight loss may ameliorate serum inflammatory cytokines and other serum metabolic risk factors, 2) incubation of adipocytes and monocytes with postintervention serum led to reductions in levels of secreted cytokines, and 3) the lifestyle intervention modified serum levels of several lipomic species, including saturated fatty acids thought to be related to inflammation.

The present study involved overweight and obese insulin-resistant youth with elevated levels of proinflammatory cytokines. Previous studies have documented associations between these risk factors (12, 15, 37). Short-term lifestyle intervention resulted in reductions in proinflammatory cytokines and metabolic risk factors, including IL-6, IL-8, TNFα, PAI-1, resistin,
amylin, and leptin, and an increase in adiponectin. The reductions in proinflammatory cytokines are possibly the result of the low saturated fat content in the diet, since saturated fatty acids are potent inducers of inflammation by activating the NF-κB pathway (23, 40). Indeed, before and after the intervention, we noted strong correlations between saturated FAs and IL-6, IL-8, and TNFα (Fig. 5). Additionally, an increased intake of ω-3 FA from fish and plant sources, which blocks the NF-κB pathway and stimulates anti-inflammatory pathways (17, 25), may also have contributed to the reduced inflammation.

In vitro, coculture of adipocytes and macrophages, showed that proinflammatory cytokines are positively correlated with inflammatory markers (4, 10). In the current study, MCP-1 from the cultured cells (3). In the current study, MCP-1 from the cultured cells was lower following treatment with postintervention serum than with preintervention serum, although this finding was not statistically significant. Previously, no significant changes in MCP-1 serum levels in the study subjects were found, although addition of postintervention serum to human aortic endothelial cells resulted in a decrease in MCP-1 production (35).

Obesity is also associated with an increase in macrophage infiltration of adipose tissue (10, 22). Itoh et al. (14), using an in vitro coculture of adipocytes and macrophages, showed that ω-3 fatty acids reversed the coculture-induced decrease in adiponectin secretion at least in part through downregulation of TNFα in macrophages. Recent work by Oh et al. (25) revealed a mechanistic link between ω-3 fatty acids and inhibition of inflammatory pathways in the macrophage as well as reduction of inflammatory macrophages in adipose tissue. This suggests an additional mechanism by which this intervention reduced inflammation. Indeed, lipomic analysis of FA composition in the serum reflects the reduction in saturated FAs and an increase in docosahexaenoic acid in the diet, and the polyunsaturated FAs were inversely correlated with proinflammatory cytokines IL-6 and IL-1ra.

One of the key findings of the lipomic analyses showed an increase in the 18:1/18:0 ratio, which is commonly used as an index of hepatic stearoyl-CoA desaturase 1 activity (42). Increased stearoyl-CoA desaturase 1 activity has been associated with a decrease in liver fat and improvement in insulin sensitivity (42). Also, the change in the 18:1/18:0 ratio was found to be negatively correlated with insulin and HOMA-IR. In addition, we noted a 24% drop in 14:0 FA levels, and shorter-chain saturated FA levels, especially 14:0, in TG have been associated with an increased risk of type 2 diabetes (34). The decrease in amylin further illustrates the improvement in metabolic health, since hyperamylinemia occurs in type 2 diabetes,
Fig. 5. Pearson’s correlation matrices for associativity between measurements pre- and postintervention (16 subjects). Measurements with strong positive correlations are colored in green, and items with negative correlations are colored in red. Measurements from the preintervention set were clustered along the y-axis using hierarchical clustering, and that ordering was induced for the postintervention set. Correlative relationships are considered significant at $\alpha < 0.05$ and are indicated with an asterisk within the corresponding cell. The white boxes highlight correlative associations of interest in the Discussion.
and amylin has been shown to cause insulin resistance in animal models (18, 46). The decrease in amylin was positively correlated with decreases seen in total FAs, IL-6, IL-8, insulin, HOMA-IR, and TG, which is in agreement with previous studies (13, 32). Furthermore, the change in amylin was positively correlated to the reductions in 14:0 and 20:0 FAs, which to our knowledge has not been shown previously.

In addition to changes in diet, the exercise intervention is also believed to be a mediator of anti-inflammatory effects, since increases in physical activity correlate with reductions in inflammation. Platat et al. (31) found negative correlations between exercise and inflammation in children independent of adiposity and fat localization. Although our subjects did lose a statistically significant amount of weight, body weight was reduced by 4%, whereas inflammatory cytokines were reduced 40–50%, suggesting that there may be a direct effect of physical activity independent of adiposity, mediating the reduction in inflammation. Pedersen and Fischer (29) and Pedersen and Pedersen (30) demonstrated that IL-6 released by skeletal muscle acts in an anti-inflammatory manner to inhibit proinflammatory pathways and stimulate anti-inflammatory pathways. Although this theory conflicts with the traditional proinflammatory role of IL-6 and a better understanding of myokines is needed, the difference in IL-6 action may be related to chronic vs. acute production and tissue specificity. Nevertheless, IL-6 acting as a myokine has been associated with an upregulation in IL-1ra and IL-10 and a decrease in production of TNFα and IL-1β (28–30). The overall effect also decreases insulin resistance induced by these inflammatory pathways (28). This mechanism may in part explain the significant changes in inflammation and the metabolic profile that we observed while the subjects remained overweight/obese.

In summary, a major strength of the present study is that the subjects had all of their food provided and physical activity monitored to ensure adherence to the intervention. Additionally, the diet was ad libitum, a major advantage in cases where overeating is an issue, and thus it is a more realistic program to implement into the daily lives of children rather than induced caloric restriction. Overall, the results provide evidence that a
short-term diet and exercise intervention can ameliorate the increase in serum metabolic risk factors and inflammation seen in obesity associated with an unhealthy diet and physical inactivity.

**REFERENCES**


