Treatment with constitutive androstane receptor ligand during pregnancy prevents insulin resistance in offspring from high-fat diet-induced obese pregnant mice

Hisashi Masuyama and Yuji Hiramatsu

Department of Obstetrics and Gynecology, Okayama University Graduate School of Medicine, Dentistry and Pharmaceutical Sciences, Okayama, Japan

Submitted 4 April 2012; accepted in final form 25 May 2012

Masuyama H, Hiramatsu Y. Treatment with constitutive androstane receptor ligand during pregnancy prevents insulin resistance in offspring from high-fat diet-induced obese pregnant mice. Am J Physiol Endocrinol Metab 303: E293–E300, 2012. First published May 29, 2012; doi:10.1152/ajpendo.00167.2012.—The constitutive androstane receptor (CAR) has been reported to decrease insulin resistance even during pregnancy, while exposure to a high-fat diet (HFD) in utero in mice can induce a type 2 diabetes phenotype that can be transmitted to the progeny. Therefore, we examined whether treatment with a CAR ligand during pregnancy could prevent hypertension, insulin resistance, and hyperlipidemia in the offspring from HFD-induced obese pregnant mice (OH mice). We employed four groups of offspring from HFD-fed and control diet-fed pregnant mice with or without treatment with a CAR ligand. Treatment with a CAR ligand during pregnancy improved glucose tolerance and the levels of triglyceride and adipocytokine and restored the changes induced by HFD with amelioration of hypertension in the adult OH mice. This treatment also increased adiponectin mRNA expression, suppressed leptin expression in adipose tissues of OH mice, and abolished the effect of HFD on the epigenetic modifications of the genes encoding adiponectin and leptin in the offspring during immaturity and adulthood. Our data suggest that CAR might be a potential therapeutic target to prevent metabolic syndrome in adulthood of offspring exposed to an HFD in utero.

adipocytokine; constitutive androstane receptor; epigenetic change; insulin resistance

MATERIALS AND METHODS

Materials and animal procedures. TCPOBOP, d-glucose, and human insulin were purchased from Sigma-Aldrich (St. Louis, MO). Female, 8-wk-old ICR strain mice were obtained from Charles River (Tokyo, Japan), and six female pregnant mice and 24 offspring were examined per group for all in vivo experiments. After 4 wk of feeding with the HFD (energy content 62% fat, 18% protein, and 20% carbohydrate) or a control diet (CD; 12% fat, 28% protein, and 60% carbohydrate) purchased from Oriental Yeast (Tokyo, Japan), the mice were weighed and mated. Females were checked daily for postcoital plugs, and the presence of a plug was taken to indicate day 0.5 of pregnancy. Eight-week-old male mice for mating were fed with the CD for 4 wk before experiments or mating. The pregnant
mice had free access to food and water, and their food consumption was estimated by weighing the remaining food every day. Pregnant HFD- and CD-fed mice received once-weekly intraperitoneal injections of TCPOBOP (0.5 mg/kg) dissolved in corn oil, or with corn oil alone (control) during pregnancy. Injections were started at day 0.5 of pregnancy or at the indicated day for specific experiments and finished at day 0.5 after delivery. The maternal weight on day 20 of gestation and neonatal weight on day 0.5 after birth were measured, and the offspring weights were measured every 2 wk. All offspring were weaned on the CD at 3 wk of age. After weaning, the offspring had free access to food and water, and their food consumption was estimated by weighing the remaining food every day. Body composition was analyzed in live mice using EchoMRI-100 (Echo Medical Systems, Houston, TX). The systolic blood pressure of offspring was measured at 12 and 24 wk after birth by the tail-cuff method using a Softron BP98A tail-cuff hemodynamometer (Softron, Tokyo, Japan) after the behavior and heart rate of the mice had stabilized. Blood pressure is reported as the mean of at least three measurements recorded during the same session, which had to vary by <5%. Most of the blood pressure values were within the required range once the mice had stabilized. At 2, 12, and 24 wk of age, the mice were anesthetized with ether, and the white mesenteric adipose tissue were removed, frozen immediately, and stored at −70°C until analysis (n = 6, three male and three female mice for each group). Total RNA was extracted using TRIzol reagent (Life Technologies, Carlsbad, CA), according to the manufacturer’s instructions. The mice were kept in a temperature- and light-controlled room with free access to food and water except during tolerance tests for glucose (GTT) and insulin (ITT). All animal procedures were approved by the Institutional Animal Care and Use Committee of Okayama University.

### Table 1. Maternal characteristics and neonatal weights

<table>
<thead>
<tr>
<th>Group</th>
<th>Maternal Wt, g</th>
<th>HOMA-IR</th>
<th>Triglycerides, mg/dl</th>
<th>Leptin, ng/dl</th>
<th>Adiponectin, µg/dl</th>
<th>Neonatal Wt, g</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD</td>
<td>38.2 ± 3.4</td>
<td>3.51 ± 0.44</td>
<td>244 ± 24</td>
<td>3.8 ± 0.5</td>
<td>10.4 ± 1.2</td>
<td>1.24 ± 0.15</td>
</tr>
<tr>
<td>HFD</td>
<td>48.1 ± 6.8*</td>
<td>4.95 ± 0.61*</td>
<td>335 ± 48*</td>
<td>6.6 ± 0.8*</td>
<td>5.2 ± 0.5*</td>
<td>1.61 ± 0.18*</td>
</tr>
<tr>
<td>CD + TCPOBOP</td>
<td>37.8 ± 3.5</td>
<td>3.33 ± 0.49</td>
<td>221 ± 33</td>
<td>3.5 ± 0.4</td>
<td>11.0 ± 0.8</td>
<td>1.22 ± 0.11</td>
</tr>
<tr>
<td>HFD + TCPOBOP</td>
<td>39.3 ± 4.0**</td>
<td>3.63 ± 0.55**</td>
<td>237 ± 28**</td>
<td>3.9 ± 0.5**</td>
<td>10.4 ± 1.0**</td>
<td>1.28 ± 0.14**</td>
</tr>
</tbody>
</table>

Data are means ± SE. CD, control diet; HFD, high-fat diet; HOMA-IR, homeostasis model assessment-insulin resistance; TCPOBOP, 1,4-bis[2-(3,5-dichloropyridyloxy)]benzene. P < 0.01 vs. CD-fed pregnant mice (*) and vs. HFD-fed pregnant mice with TCPOBOP treatment (**).
GTT, ITT, and measurements of insulin, total triglyceride, adiponectin, and leptin levels. Offspring at 12 or 24 wk of age were fasted for 16 h before receiving an intraperitoneal injection of D-glucose (2 g/kg body wt) for the GTT or for 4 h before receiving an intraperitoneal injection of human insulin (1.0 U/kg body wt) for the ITT (n = 6, three male and three female mice for each group). Blood samples from tails were taken before and at 30, 60, 90, and 120 min after the injection. Blood glucose levels were measured by the glucose oxidase method using a Medisafe automated analyzer (Termo, Tokyo, Japan). Fasting insulin, total triglyceride, adiponectin, and leptin levels were determined using enzyme-linked immune sorbent assay (ELISA) kits [insulin and triglycerides (Morinaga Institute of Biological Sciences, Yokohama, Japan); adiponectin and leptin (R&D Systems, Minneapolis, MN)]. Blood sample volumes for each measurement were 10–20 μl, and the total sample volume collected from each mouse was <200 μl, which was <5% of total blood volume. Homeostasis model assessment-insulin resistance (HOMA-IR) was calculated as the fasting insulin concentration (μU/ml) × fasting glucose concentration (mg/dl)/405 (12).

Real-time quantitative PCR. Real-time quantitative PCR was performed to measure the mRNA levels of the leptin and adiponectin genes using a StepOne Real-time PCR System and a TaqMan RNA-to-CT Gene Kit (Applied Biosystems, Carlsbad, CA). Specific primers for the mouse leptin, adiponectin, and β-actin sequences were purchased from Applied Biosystems. Sequences of specific primers and accession numbers were as described (8, 17, 33). RNA samples (25 ng) were assayed in triplicate using 15 pmol of gene-specific primers and 5 pmol of gene-specific probes. Because we observed that there were no significant differences of β-actin expression under different diet conditions or TCPOBOP treatment using another housekeeping gene, GAPDH as a control (data not shown), mouse β-actin mRNA levels were measured as an internal control using a predeveloped TaqMan primer and a probe mixture (Applied Biosystems). The mRNA levels of the target genes were normalized by the β-actin mRNA levels.

Chromatin immunoprecipitation assays. Chromatin immunoprecipitation (ChIP) assays were performed using a ChIP assay kit (Upstate Biotechnology, Lake Placid, NY) according to the manufacturer’s protocol. Adipose tissues from the HFD-induced obese (OH) and OC groups (n = 6, three male and three female mice for each group) at 2 and 24 wk of age were taken for sampling. Briefly, 20-mg aliquots of frozen samples were ground in liquid nitrogen using a mortar and pestle and then washed with PBS at room temperature. The samples were then resuspended in PBS and cross-linked in 1% formaldehyde for 10 min. After centrifugation, the pellet was resuspended in nucleus-swelling buffer containing protease and phosphorylation inhibitors. The nuclei were lysed in SDS lysis buffer containing protease and phosphorylation inhibitors. The chromatin was sonicated to reduce DNA fragment lengths to 0.3–1.0 kb. Chromatin was precleared in the presence of 20 μl of normal serum, 2 μg

Fig. 2. Effects of treatment with a CAR ligand during pregnancy on glucose intolerance and insulin resistance in offspring showing glucose tolerance test (GTT), insulin tolerance test (ITT), and homeostasis model assessment-insulin resistance (HOMA-IR) results at 12 (A–C) and 24 (D–F) wk of age. Pregnant HFD- and CD-fed mice received once-weekly intraperitoneal injections of TCPOBOP (0.5 mg/kg) dissolved in corn oil or corn oil alone (control). Offspring at 12 and 24 wk of age were fasted for 16 h before receiving an ip injection of D-glucose (2 g/kg body wt) for the GTT or for 4 h before receiving an ip injection of human insulin (1.0 U/kg body wt) for the ITT. Blood samples were taken before and at 30, 60, 90, and 120 min after the injections of glucose or insulin. Blood glucose levels (mg/dl) were measured by the glucose oxidase method, and fasting serum insulin levels (μU/ml) were determined using enzyme-linked immune sorbent assay (ELISA) kits. The results are shown as means ± SD (n = 6/group). *P < 0.01 vs. OHT, OCT, and OC mice (see legend to Fig. 1).
of salmon sperm DNA, and 80 μl of 25% protein A-agarose slurry. Precleared chromatin samples were subjected to immunoprecipitation at 4°C overnight in the presence of 2 μg of rabbit polyclonal antibodies against acetyl-histone H3 at lysine 9 (acetyl H3K9; Millipore, Bedford, MA), dimethyl histone H3 at lysine 9 (dimethyl H3K9; Millipore), and monomethyl histone H4 at lysine 20 (monomethyl H4K20; Abcam, Cambridge, MA), or nonimmune rabbit IgG (Millipore). After the complex was collected by incubation with 60 μl of a 25% protein A-Sepharose slurry and centrifugation, the beads were washed five times, and the chromatin-immune complex was eluted. After the cross-linking was reversed, DNA was purified and used as a template for PCR. PCR was performed using primer sets specific for the promoter region of the mouse adiponectin gene (positions −549 to −481) (25) and the promoter region of the mouse leptin gene (−181 to +20) (35).

Statistical analysis. Statistical analyses were performed by two-way ANOVA for comparison among four offspring groups, HFD-fed or CD-fed with or without TCPOBOP treatment during pregnancy, and by repeated-measures ANOVA for GTT and ITT followed by Dunnett’s test. All statistical analyses were performed using StatView software, version 5.0 (Abacus Concepts, Berkeley, CA). Data are presented as means ± SD, and P < 0.05 was taken to indicate statistical significance.

RESULTS

The weight, caloric intake, fat mass-to-body weight ratio, and blood pressure of offspring. The offspring from CD-fed dams with TCPOBOP treatment are designated OCT mice, and the offspring from CD-fed dams without treatment are designated OC mice; the offspring of HFD-fed dams with TCPOBOP treatment are designated OHT mice, and the offspring of HFD-fed dams without treatment are designated OH mice. TCPOBOP treatment decreased the maternal weight of HFD-fed pregnant mice on day 20 of gestation and restored the increased maternal weight induced by HFD (Table 1). Neonatal weights from the HFD-fed pregnant mice on day 0.5 after birth were significantly greater than that from CD-fed pregnant mice (P < 0.01), whereas TCPOBOP treatment decreased the mean neonatal weight of the offspring from HFD-fed pregnant mice and restored the increased fetal weight induced by maternal HFD (Table 1). There was no significant difference in mean litter size among the four groups (overall 9.1 ± 0.6). The mean weight of the OHT mice was significantly lower than the OH mice from 14 wk of age, and there were no significant differences of weight between the OHT and OC mice (Fig. 1A). The caloric intake of OHT mice was lower than that of the OH mice and was not significantly different from the OC and OCT mice from 8 wk of age, before a significant increase in body weight (Fig. 1B). Also, the gain of fat mass in the OHT mice was decreased significantly compared with the OH mice and was not significantly different from the OC and OCT mice from 12 wk of age using MRI analysis (Fig. 1C). Systolic blood pressure in the OH mice was elevated significantly compared with the OC mice at 24 wk of age (P < 0.01, Fig. 1E), but not at 12 wk (Fig. 1D). TCPOBOP treatment during pregnancy prevented the elevation of blood pressure in OH mice at 24 wk of age (Fig. 1E).

Fig. 3. The effects of treatment with a CAR ligand during pregnancy on serum triglyceride, adiponectin, and leptin levels in offspring. Pregnant HFD- and CD-fed mice received once-weekly ip injections of TCPOBOP (0.5 mg/kg) dissolved in corn oil or corn oil alone (controls). Offspring at 12 and 24 wk of age were fasted for 16 h, and blood samples were taken. Serum total triglyceride (A and D), adiponectin (B and E), and leptin (C and F) levels were determined using ELISA kits. The results are shown as means ± SD (n = 6/group). *P < 0.01 vs. OH, OCT, and OC mice (see legend to Fig. 1).
CAR activation during pregnancy prevented glucose intolerance and insulin resistance in OH mice. We performed GTTs and ITTs, measured serum insulin level, and calculated the HOMA-IR in offspring from HFD-fed or CD-fed pregnant mice with or without treatment with TCPOBOP at 12 and 24 wk of age, to examine the effects of TCPOBOP treatment during pregnancy on glucose metabolism in offspring. There were no significant differences of GTT, ITT, and HOMA-IR between the OH and OC mice at 12 wk of age (Fig. 2, A–C). However, the OH mice exhibited significantly worse glucose tolerance and insulin sensitivity and significantly increased HOMA-IR values compared with the OC mice at 24 wk of age ($P < 0.01$ for all significant differences; Fig. 2, D–F). Next, we examined whether TCPOBOP treatment during pregnancy could improve the glucose tolerance in the offspring. OHT mice exhibited significantly improved glucose tolerance compared with the OH mice at 24 wk of age ($P < 0.01$, Fig. 2, D and E). HOMA-IR was also decreased in OHT mice compared with the control vehicle-treated OH mice at 24 wk of age, and the HFD-induced changes were restored ($P < 0.01$, Fig. 2F). OCT mice showed no significant changes compared with control OC mice at 12 and 24 wk of age.

Treatment with a CAR ligand during pregnancy improved serum triglyceride and adipocytokine levels in OH mice. To clarify whether TCPOBOP treatment during pregnancy would affect lipid metabolism and adipocytokine levels in offspring, serum triglyceride, leptin, and adiponectin levels were examined. The total triglyceride and leptin levels were significantly higher, whereas the adiponectin level was significantly lower in OH mice compared with OC mice at 12 (Fig. 3, A–C) and 24 (Fig. 3, D–F) wk of age ($P < 0.01$ for all significant differences). The triglyceride and leptin levels were decreased and the adiponectin level was increased in OHT mice, and the changes in triglyceride, leptin, and adiponectin levels were restored by TCPOBOP treatment during pregnancy (Fig. 3, A–F). OCT mice showed no significant changes compared with vehicle-treated control OC mice at both 12 and 24 wk of age.

Effects of treatment with a CAR ligand during pregnancy on leptin and adiponectin gene expression levels in the adipose tissue of offspring. The leptin gene was upregulated significantly, whereas the adiponectin gene was downregulated in the white adipose tissue of OH mice compared with the OC mice at both 12 (Fig. 4, A and B) and 24 wk (Fig. 4, C and D) of age ($P < 0.01$ for all significant differences). TCPOBOP treatment significantly decreased the leptin mRNA expression and increased the adiponectin mRNA expression in the white adipose tissue of the OHT mice compared with the OH control mice ($P < 0.01$ for all significant differences), and the changes in both expression levels induced by the HFD during pregnancy were restored (Fig. 4, A–D). There were no significant differences in these gene expression levels between the OC and OCT mice.

Effects of treatment with a CAR ligand during pregnancy on modifications to H3K9 and H4K20 in the promoter regions of the adiponectin and leptin genes in the adipose tissue of offspring. To investigate whether treatment with a CAR ligand during pregnancy would affect histone modifications in the white adipose tissue, we performed ChIP assays using antibodies for acetyl and dimethyl H3K9 and monomethyl H4K20 at 2 and 24 wk of age. The acetyl H3K9 level in the adipose tissue of offspring, we performed ChIP assays using antibodies for acetyl and dimethyl H3K9 and monomethyl H4K20 at 2 and 24 wk of age. The acetyl H3K9 level in the promoter region of OCT mice was remarkably higher than in OH mice, whereas the dimethyl H3K9 was decreased in adipose tissues of OC mice compared with the OH mice. Moreover, treatment with the CAR ligand during pregnancy abolished the effect of HFD on the modification of H3K9 at 2 and 24 wk of age. However, there were only weak detection levels and no significant differences in monomethyl H4K20 levels at 2 and 24 wk of age (Fig. 5A). However, the

![Fig. 4. Effects of treatment with a CAR ligand during pregnancy on leptin and adiponectin gene expression in white adipose tissue of offspring.](http://ajpendo.physiology.org/doi/10.1152/ajpendo.00167.2012)
monomethyl H4K20 level was increased significantly in the *leptin* promoter region of OH mice compared with OC mice, but treatment with the CAR ligand during pregnancy abolished the effect of HFD on the modification of H4K20 at 2 and 24 wk of age. There were only weak detections and no significant differences in acetyl and dimethyl H3K9 levels in both groups at 2 and 24 wk of age (Fig. 5B). There were no effects of maternal diet on the association of IgG binding with the promoter regions of *leptin* or *adiponectin* in adipose tissues and no significant gender differences in epigenetic changes by maternal HFD and the effect of TCPOBOP (data not shown).

Effect of different treatment timing with a CAR ligand on modifications to H3K9 and H4K20 in the promoter regions of the adiponectin and leptin genes in the adipose tissue of offspring. We used offspring from HFD-fed dams with different timings of TCPOBOP treatment to examine the effects on epigenetic changes in the *adiponectin* and *leptin* genes (Fig. 6A). Treatments started at days −7.5, 0.5, and 7.5 of gestation abolished the effect of HFD on the modification of H3K9 at 24 wk of age, but the treatments started at day 14.5 of gestation and 0.5 days after birth had no effect on this modification (Fig. 6C).

DISCUSSION

Adipose tissue functions as a highly specialized endocrine and paracrine tissue, producing an array of adipocytokines such as leptin, tumor necrosis factor-α, and adiponectin, as well as eliciting cell-mediated effects via proinflammatory and anti-inflammatory cells, producing various cytokines and chemokines. Such factors have local and systemic biological effects and influence insulin sensitivity and the development of metabolic diseases (9). Leptin plays important roles not only in modulating satiety and energy homeostasis (16, 23) and adiponectin might maintain endothelial function and its deficiency can lead to endothelial dysfunction/hypertension (14, 27, 30). Previously, we demonstrated that HFD-fed pregnant mice exhibited signs of preeclampsia, including hypertension, pro-

---

**Fig. 5.** Effects of treatment with a CAR ligand during pregnancy on modifications to histone H3 at lysine (H3K9) and histone H4 at lysine 20 (H4K20) in the promoter regions of the *leptin* and *adiponectin* genes in the adipose tissue of offspring. Chromatin immunoprecipitation (ChIP) assays were performed on the chromatin extracts from the white mesenteric adipose tissue of OH and OC mice to examine the effect of treatment with a CAR ligand during pregnancy on H3K9 and H4K20 modifications in the promoter regions of the *adiponectin* (A) and *leptin* (B) genes of offspring at 2 and 24 wk of age using anti-acetyl and anti-dimethyl H3K9 and anti-monomethyl H4K20 antibodies. The results are shown as means ± SD (n = 6/group). *P < 0.01 vs. OH, OCT, and OC mice (see legend to Fig. 1).
...activation during pregnancy. Exposure to an HFD in utero might be restored by CAR ligand TCPOBOP in immature and mature mice, suggesting that the effects of an HFD on the epigenetic modifications of H3K9 in the promoter regions of the adiponectin and leptin genes in the adipose tissue of offspring are ameliorated by CAR ligand TCPOBOP in the promoter regions of the adiponectin (B) and leptin (C) genes at 24 wk of age using anti-acetyl and anti-dimethyl H3K9 and anti-monomethyl H4K20 antibodies. The results are shown as means ± SD (n = 6/group). *P < 0.01 vs. OH, OCT, and OC mice (see legend to Fig. 1).

A previous report demonstrated that exposure to HFD in utero had the potential to program the glucose production and altered glucose sensitivity in adulthood (28). In addition, H3K9 methylation status is important in regulating the expression of metabolic genes in adipose tissue (13, 31), while methylation of H3K9 had a positive effect, but that of H4K20 had negative effect on adipogenesis through the peroxisome proliferator-activated receptor γ-mediated pathway (34). Moreover, there have been several reports that modifications of H3K4 and H3K9 in the promoter region of adiponectin might play important roles in adipogenesis (22, 25) and that the majority of target genes, including leptin for peroxisome proliferator-activated receptor-γ modified by H4K20 monomethylation, were expressed at high levels during adipocyte differentiation (13). We have previously demonstrated that OH mice exhibited a modification of H3K9 from methylation to acetylation in the adiponectin promoter region and methylation of H4K20 in the leptin promoter region of adipose tissue, suggesting that these histone modifications might suppress the expression of these genes in adipose tissues of OH mice (18). Therefore, we examined whether treatment with a CAR ligand during pregnancy would affect these changes in histones in OH mice. Our data showed that the effect of HFD on the epigenetic modifications of H3K9 in the adiponectin promoter and of H4K20 in the leptin promoter in the offspring were abolished in both immature and mature mice, suggesting that the effects of an exposure to an HFD in utero might be restored by CAR activation during pregnancy.

We also observed that the timing of TCPOBOP administration affected the epigenetic changes of adiponectin and leptin genes differently in adipose tissues of OH mice. Thus, administration of TCPOBOP in midpregnancy as well as in the preconception stage and early pregnancy abolished the effect of HFD on the modification of adiponectin and leptin genes, but administration at late pregnancy and after birth had no effect on this. Findings from the Dutch famine of 1944 showed that maternal undernutrition during gestation has important effects on health in later life, but the effects on health depend on its timing during gestation, and early gestation seems to be a particularly vulnerable period (24). Moreover, differences in DNA methylation patterns after exposure to prenatal undernutrition have been demonstrated to be timing-specific in the mouse (32). Although there has been no report examining the effects of different HFD exposure times in utero on metabolic syndrome-like phenomena, treatment with a CAR ligand at a preconception period or during early pregnancy might prevent the onset of metabolic syndrome in adulthood of offspring exposed to an HFD in utero. However, we observed that the adipocyte function in OH mice seemed to be improved after TCPOBOP treatment during pregnancy although CAR has not been detected in adipose tissues (20). It is possible that treatment with a CAR ligand in utero may indirectly affect the epigenetic modification of adipocytokines in adipose tissue of OH mice through combined changes to gluconeogenesis and lipogenesis. Further analysis will be required to resolve this issue.

Measures to alleviate fetal overgrowth in obese and diabetic women represent an early intervention strategy that could help decrease the prevalence of obesity and diabetes in future generations. Previously, we demonstrated that TCPOBOP treatment ameliorated fetal growth in addition to improving maternal glucose metabolism and adipocyte function in HFD-fed pregnant mice (20). In the present study, we observed that TCPOBOP treatment during pregnancy abolished metabolic-like phenomenon in OH mice, suggesting that CAR activation will decrease the risk of offspring developing metabolic syndrome and diabetes in adulthood.

Taken together, our data suggest that treatment with the CAR ligand TCPOBOP during pregnancy can improve insulin sensitivity and adipocytokine levels and that these improvements were associated with the amelioration of adult hypertension in OH mice. CAR might be a potential therapeutic...
target to prevent metabolic syndrome in adulthood of offspring exposed to an HFD in utero.

**GRANTS**

This work was supported in part by a research grant (22591856) from the Ministry of Education, Culture, Sports, Science and Technology of Japan.

**DISCLOSURES**

No conflicts of interest, financial or otherwise, are declared by the authors.

**AUTHOR CONTRIBUTIONS**

Author contributions: H.M. conception and design of research; H.M. performed experiments; H.M. analyzed data; H.M. interpreted results of experiments; H.M. prepared figures; H.M. drafted manuscript; H.M. and Y.H. performed experiments; H.M. and Y.H. edited and revised manuscript; H.M. and Y.H. approved final version of manuscript.

**REFERENCES**