Multiparity leads to obesity and inflammation in mothers and obesity in male offspring

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Multiparity leads to obesity and inflammation in mothers and obesity in male offspring. Am J Physiol Endocrinol Metab 302: E449–E457, 2012. First published November 29, 2011; doi:10.1152/ajpendo.00487.2011.—Multiparity is an independent risk factor for obesity in parous females. In addition to being a health issue for the mother, offspring of multiparous females may also be at risk for obesity later in life. The aim of the current study was to establish a mouse model that mimics the human pathology of multiparity and determine the effects of multiparity-induced obesity (MIO) on offspring in adulthood. C57BL/6 mice were mated and studied when primiparous (1st pregnancy) or multiparous (4th pregnancy). Dams became obese with multiparity, an effect that was independent of the age of the dam. Multiparous dams also had increased markers of inflammation (JNK activation, cytokine expression) in adipose tissue and liver that was greater than inflammation in nulliparous females made obese with a high-fat diet. Placental inflammation was prevalent in multiparous vs. primiparous dams as well. Male offspring of the multiparous dams developed increased adiposity by 24 wk of age relative to the progeny of primiparous dams, although food consumption was similar in both groups. Lipid metabolism was altered in liver and fat in that mRNA levels of regulatory genes (PGC-1α) as well as metabolic genes (CPT I) and Akt phosphorylation were decreased in offspring of multiparous dams. Thus, in mice, as in humans, multiparity increases adiposity and is associated with hepatic and placental inflammation and abnormal glucose tolerance. Importantly, MIO leads to increased body fat and metabolic dysfunction in the offspring, suggesting a role in the propagation of obesity.

Developmental programming; diabetes; peroxisome proliferator-activated receptor-γ coactivator-1α

Obesity has become an epidemic in developed countries and is on the rise in underdeveloped countries. Consequently, a significant number of women in their reproductive years, including those that are pregnant, are obese or overweight (30, 31). Obesity during gestation poses a significant clinical problem since it is linked to a spectrum of maternal complications, including gestational diabetes, exaggerated inflammation, hypertension, thromboembolism, preeclampsia, and delivery complications (26, 31, 55, 62, 69). Multiparity is a cause of weight gain in women during their reproductive years and is in fact an independent predictor of obesity (12, 20). The increase in multiparity-induced obesity (MIO) has followed overall trends in obesity, since gestational weight gain is additive to prepregnancy weight and the inability to lose weight gained postpartum (51, 53, 54). Moreover, overweight women are more likely than leaner women to retain the weight gained during pregnancy (19). Thus, as the population becomes heavier, the effect of multiparity on body weight is amplified. Similar to obesity, pregnancy has been described as an inflammatory state, with the placenta contributing increased levels of cytokines from both the trophoblasts and resident placental macrophages (reviewed in Ref. 22) and with an increase in the numbers of activated monocytes (39). Thus multiparous women may be subjected to repeated episodes of this placental inflammation with adverse consequences. In fact, it is known that women who have had multiple pregnancies are at an increased risk to develop diabetes, independent of visceral adiposity (1), and diabetes has been associated with chronic inflammation (5, 48, 66). Since obese pregnant women have increased inflammation compared with lean pregnant women (10, 59), obese multiparous women could have an even greater inflammatory state.

Gestational obesity is known to cause acute and chronic consequences in the offspring as well as the female itself. Acute consequences range from shoulder dystocia, low Apgar score, macrosomia, congenital defects, and death (42, 46). Chronic consequences include developmental programming, leading to an increased risk of obesity and glucose intolerance in adulthood (reviewed in Refs. 2, 9, 23, and 44). The fact that obesity during pregnancy increases the incidence of obesity in the offspring raises the worrisome prospect of transgenerational perpetuation of obesity (8, 51). Although the mediators of developmental programming of metabolism are not clear, gestational diabetes, maternal overnutrition, circulating factors associated with maternal obesity, or factors associated with inflammation in the placenta or other tissues of obese pregnant females are all possible routes of regulation (2, 9, 23, 44). Although multiparity is a risk factor for obesity, there is little information about the risks of adverse metabolic outcomes in the offspring of these women.

Adiposity is the result of positive energy balance, and caloric intake greater than caloric expenditure. Hunger and satiety are controlled by a number of factors in specific regions in the brain, and changes in activity of any number of these pathways could affect food consumption and thereby lead to altered adiposity. A reduction in energy expenditure could also lead to accelerated fat deposition. As in the brain, a variety of proteins and pathways are involved in peripheral lipid metabolism and energy expenditure. However, one protein has been suggested to be key in the regulation of energy expenditure, and that protein is peroxisome proliferator-activated receptor (PPAR)γ coactivator-1α (PGC-1α; reviewed in Refs. 15, 21, and 25). PGC-1α is upstream of mitochondrial biogenesis and fatty acid oxidation, as well as oxidative phosphorylation and other mitochondrial functions, and as such plays a pivotal role in energy metabolism. PGC-1α coactivates...
mitochondrial biogenesis and fatty acid oxidation with other proteins, i.e., the estrogen-related receptor. Although processes are regulated by protein complexes, levels of PGC-1α are thought to be the driving force for gene transcription of the downstream targets (64).

Thus, the present study was designed in two parts. In the first part, we established a mouse model that mimics MIO in humans. Female parous mice became obese by the 4th pregnancy with exaggerated inflammation. In the second part, we demonstrated that male offspring of these multiparous females have altered metabolism, resulting in increased adiposity. The change in body composition was not the result of a change in input of energy intake but was due to a change in energy output (lipid catabolism). Specifically, we found marked decreases in the mRNA levels of carnitine palmitoyltransferase I (CPT I), the rate-limiting step for fatty acid oxidation. The decrease was likely mediated by decreased levels of direct upstream regulators of CPT I, those being PGC-1α and/or PPARα. Thus, multiparous females with marked obesity and inflammation induce developmental programming, leading to increased adiposity in adult male offspring.

RESEARCH DESIGN AND METHODS

Animals

Pregnant dams. All male and nonpregnant female C57BL/6 mice were purchased from The Jackson Laboratories and fed a pelleted chow (Harlan Laboratories); females were 10 wk old when they arrived. Animals were housed in a temperature- and humidity-controlled room and subjected to 12 h of light and 12 h of darkness. After ~1 wk acclimation, females were fed a breeder chow that contained 9% fat (wt/wt; PMI Nutrition International) for 1–2 wk prior to mating. To mate females, one male was placed in a cage with three females. The presence of a postcopulatory plug was checked when the lights came on and denoted 0.5 days postconception (dpc). When a plug was detected, female mice were separated into their own cage. After 5 days of mating, males were separated from remaining non-pregnant females. After 2 days, the process was repeated. Females underwent mating until a plug was detected or until three mating cycles were completed. For multiparous females, pups were born and allowed to suckle. Pups were weaned at 21 dpc and tissues were collected after a 6-h fast. All animal protocols were approved by the Institutional Animal Care and Use Committee at the University of Cincinnati.

GTT

GTT were performed similarly in fasted pregnant females and male offspring (6-h fast). Animals were injected intraperitoneally (ip) with glucose (2 mg/g). Blood was collected from the tail, and glucose was measured in triplicate with a glucometer using glucose strips (Abbott Laboratories) at 0, 15, 30, 60, 120, and 180 min postinjection.

Real-Time PCR

Livers and adipose tissues were collected rapidly in pregnant females (18.5 dpc) or adult offspring (~5 mos of age) after exsanguination, snap-frozen in liquid nitrogen, and stored at −80°C until use. Tissue RNA was isolated using TRIzol and stored in TRIzolol and stored in FORMAzol at −80°C. RNA was treated with RNase-free DNase I and reverse transcribed to cDNA by SuperScript II reverse transcriptase using random hexamers. PCR assays were performed on a Bio-Rad iCycler iQ real-time PCR Detection System, using SYBR green as our fluorophore; primers will be given upon request. A serial dilution of a randomly picked sample was used to generate a standard curve for each gene examined. This standard curve was used to calculate the relative levels of mRNA for the gene of interest compared with the reference/housekeeping gene (cyclophilin).

Hormone Levels and Adiposity

Insulin concentrations were measured by LINCOplex by the Mouse Metabolic Phenotype Center (MMPC) of the University of Cincinnati. Adiposity was measured by MRI (Echo MRI; Echo Medical Systems, Houston, TX).

Plasma and Liver Lipid Concentrations

Plasma lipids were measured enzymatically. Plasma was pooled and separated into lipoproteins by fast-protein liquid chromatography using two Sephadex columns in tandem as described (41), and cholesterol was measured in each fraction enzymatically. Lipids were extracted from livers with Folch, and triglyceride content was measured chemically by the MMPC of the University of Cincinnati (6). Cholesterol was extracted from saponified liver and measured by gas chromatography, using stigmastanol as an internal standard (63).

Western Blots

Expression levels of proteins or phosphorylation of proteins were determined by Western blot. Briefly, tissues were homogenized in a lysis buffer. Proteins were then separated by gel electrophoresis, and the relative amounts of phosphorylated (Cell Signaling Technology) and total JNK (Pharmingen), phosphorylated (Cell Signaling Technology) and total AKT (Cell Signaling Technology), and PGC-1α (Novus Biologicals) were determined by Western blotting using appropriate secondary antibodies and enhanced chemiluminesence plus, as described (68); similar gel loading was verified as needed after stripping membranes and reprobing with antibodies to tubulin or β-actin (33).

Statistics

Data are presented as means ± SE. Differences between dams of the 1st vs. 4th or offspring of 1st vs. 4th dams were determined by two-tailed Student’s t-tests (P < 0.05). When three groups of mice were compared, i.e., 1st, 4th, and 1st old pregnancy, animals were compared by one-way ANOVA, followed by Student-Newman-Keuls test to determine differences between groups (P < 0.05).
RESULTS

Effects of Multiparity on Maternal Adiposity and Inflammation

Previous studies have demonstrated that multiparous women become obese (12, 20) and likely have increased inflammation (1). To establish a mouse model that mimics the human pathology of multiparity, we mated C57BL/6 mice once (primiparous) or four times (multiparous). The body weights for multiparous and primiparous dams were 48.5 ± 2.2 and 33.7 ± 0.9 g, respectively. Both the retroperitoneal (RP) and ovarian (OV) fat depots weighed ≈6.5-fold more, and the inguinal (ING) depot weighed ≈3.5-fold more in the multiparous dams (Fig. 1A). In addition, multiparous mice had increased hepatic lipid content and plasma cholesterol concentrations (Table 1); the increase in plasma cholesterol was due to increased LDL as well as HDL cholesterol (data not shown).

To assess age as a contributing factor for increased adiposity, dams were mated their first time at a similar age as those of the multiparous dams (1st old pregnancy). There was significantly more fat in the ING (55% more) and OV (87% more) and a trend for more RP fat (45% more; \( P = 0.09 \)) of the multiparous dams vs. 1st old pregnant dams (Fig. 1A). These data demonstrate that in mice, as in humans, multiparity leads to increased adiposity independent of maternal age.

Since increased adiposity is often associated with glucose intolerance and insulin resistance, we examined glucose tolerance in a different set of dams at the same gestational age. Rising glucose levels were similar among primiparous and multiparous mice (Fig. 1B). However, glucose excursion was greater in dams in their fourth pregnancy compared with those in their first; specific time points post-glucose injection were significantly greater in the multiparous dams, and there was a trend for increased area under the curve of GTT in dams of the fourth compared with the first pregnancy (\( P = 0.09 \)). Glucose concentrations returned to preinjection levels by 120 min in the primiparous mice, whereas glucose levels were still elevated at 180 min in the multiparous cohort.

Another factor that is associated with obesity and can lead to abnormal glucose tolerance is inflammation (18, 66). Markers of inflammation were measured in the placentas, RP adipose tissues, and livers of the multiparous and primiparous dams. Proinflammatory cytokine (monocyte chemotactant protein-1, TNFα, IL-6) mRNA levels were increased in the RP adipose tissues of the multiparous females, whereas anti-inflammatory cytokine (arginase) mRNA levels were similar in all dams (Fig. 2A). In the placentas, levels of proinflammatory cytokines (monocyte chemotactant protein-1 and TNFα) as well as macropage markers (F4/80 and CD68) were increased, or there was a trend for an increase in the multiparous females (Fig. 2B). Consistent with greater cytokine production and more macrophages, JNK activation, as reflected in increased phosphorylation, was increased in the placentas (Fig. 2C) as well as the livers (Fig. 2D) of the multiparous females. Importantly, the increased inflammation was greater than that which occurs merely with adiposity since expression of proinflammatory cytokines was greater in RP adipose tissues of multiparous females than in nulliparous females fed a high-fat diet to age- and weight-match multiparous females (Fig. 2E). Together, these data indicate that multiparity causes exaggerated inflammation independently of its effect on adiposity.

Effects of Multiparity on Offspring Glucose Homeostasis and Adiposity

Knowing that diet-induced obese females have offspring with increased adiposity (2, 9, 23, 44), we next set out to determine whether offspring of multiparous obese dams would be at an increased risk of becoming obese. Offspring of

![Figure 1](AJP-Endocrinol Metab)
Offspring of multiparous dams (PGTT, plasma glucose concentrations had higher peaks in the offspring of multiparous females). Adipose tissues (A) are the same as those described in Fig. 1A, and livers and placenta (B–D) were collected from 7 (multiparous) or 4 (primiparous) of the dams described in Fig. 1A. A: relative mRNA levels of proteins associated with inflammation in retroperitoneal adipose tissues of young primiparous and multiparous dams compared with cyclophillin are presented. B: relative mRNA levels in the placenta are presented. C: phosphorylated and total JNK were determined in the liver by Western blot using tubulin for gel loading. D: phosphorylated and total JNK were determined in the placenta by Western blot using tubulin for gel loading. E: mRNA levels of cytokines expressed in nulliparous dams fed a high-fat diet for 4 mo (n = 5–6). Animals were age- and weight-matched to the multiparous controls. Dashed lines represent data from multiparous dams presented in B. All PCR results are presented as means ± SE. *Differences (P < 0.05) between tissues of offspring of primiparous and multiparous dams fed each diet; §P = 0.09. MCP-1, monocyte chemoattractant protein-1.

Metabolism of Adipose Tissue and Liver of Offspring

We used a targeted gene approach to determine the mechanism(s) responsible for the 40% increased adiposity independent of a change in food consumption. Initially, we studied genes involved in synthesis of fat [fatty acid synthase (FAS)] and oxidation of fat (CPT I) of adipose tissue. There was ~40% decrease in CPT I and ~45% decrease in FAS mRNA levels in male offspring of multiparous dams compared with male offspring of primiparous dams (Fig. 7, A and B). We next measured levels of genes responsible for the regulation of these genes (Fig. 7, A and B). PGC-1α mRNA levels were decreased in adipose tissues offspring of multiparous vs primiparous dams, whereas mRNA levels of PGC-1β, PPARγ, liver X receptor (LXR), and sterol regulatory element-binding protein-1 (SREBP-1) were not affected. Compared with expression in adipose tissues, CPT I mRNA levels were decreased almost 40% in liver extracts from offspring of multiparous dams, with only modest changes in FAS mRNA levels (Fig. 7B). PGC-1α mRNA levels were decreased in the livers of offspring of multiparous dams, as occurred in adipose tissue; hepatic PGC-1α protein levels paralleled mRNA levels (Fig. 7C) and were around twofold greater in the primiparous offspring, as noted by ImageJ software. Unlike adipose tissue, PGC-1β mRNA levels were also reduced significantly in the livers of offspring of multiparous dams, and PPARα mRNA levels tended to be less (P = 0.069). LXR and SREBP-1 mRNA levels did not differ from the primiparous group. The expression of phosphoenolpyruvate carboxykinase and glucose-6-phosphatase, key enzymes for hepatic glucose pro-

Fig. 2. Inflammation in adipose tissues, placenta, and livers of multi- or primiparous females. Adipose tissues (A) are the same as those described in Fig. 1A, and livers and placenta (B–D) were collected from 7 (multiparous) or 4 (primiparous) of the dams described in Fig. 1A. A: relative mRNA levels of proteins associated with inflammation in retroperitoneal adipose tissues of young primiparous and multiparous dams compared with cyclophillin are presented. B: relative mRNA levels in the placenta are presented. C: phosphorylated and total JNK were determined in the liver by Western blot using tubulin for gel loading. D: phosphorylated and total JNK were determined in the placenta by Western blot using tubulin for gel loading. E: mRNA levels of cytokines expressed in nulliparous dams fed a high-fat diet for 4 mo (n = 5–6). Animals were age- and weight-matched to the multiparous controls. Dashed lines represent data from multiparous dams presented in B. All PCR results are presented as means ± SE. *Differences (P < 0.05) between tissues of offspring of primiparous and multiparous dams fed each diet; §P = 0.09. MCP-1, monocyte chemoattractant protein-1.
DISCUSSION

In this set of studies, we have demonstrated that multiparity causes weight gain in mice, independent of age. In addition, multiparous females have increased inflammation in liver, adipose tissue, and placenta, processes that are not accounted for by increased adiposity. Multiparity also has metabolic consequences, since these animals were glucose intolerant compared with primiparous dams; this response could be explained by either the greater adiposity, increased inflammation, or some combination of these. In addition to the detrimental effects of obesity and its comorbidities, exaggerated inflammation is detrimental to the multiparous female and increases its risk for additional age-related diseases. For example, atherosclerosis is known to be a disease associated with chronic inflammation. Initial steps in atherosclerotic plaque formation involve recruitment of monocytes to the vessel walls that are then differentiated into macrophages, thereby initiating lesion formation (36, 50). Inflammation is linked with diabetes as well in that some proinflammatory cytokines can blunt insulin signaling, thereby leading to insulin resistance and diabetes (24). It is likely that obesity and all of its comorbidities and inflammation in the multiparous female could be difficult to overcome if inflammation is prominent in the brain because inflammation in the hypothalamus disrupts satiety signals originating in the brain (13, 73), making weight loss difficult.

The metabolic detriments of multiparity were not limited to the mothers but extended to their male offspring. Importantly, male mice born of multiparous mothers had increased levels of body fatness in adulthood. These findings support a model whereby multiple pregnancies induce fixed metabolic stresses on females that have heritable consequences. Since there are

Fig. 3. Body weights and composition of male offspring of primi- (1st) and multiparous (4th) dams. Male offspring were weaned at 3 wk of age and housed individually. A: weekly body weights were collected, and composition was measured at 14 and 24 wk of age. B: %fat of offspring at 14 and 24 wk of age. C: mass of fat in offspring at 14 and 24 wk of age. D: lean mass of offspring at 14 and 24 wk of age. Data are presented as means ± SE [n = 11 and 9 offspring (litters) from primi- and multiparous dams, respectively]. *Differences (P < 0.05) between tissues of offspring of primi- and multiparous dams at either age.

Fig. 4. Food consumption of offspring of multi- and primiparous dams. Consumption was measured weekly from 12 wk of age until the time of the study in the animals described in Fig. 3.

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parallels to some of these findings in humans, our results support yet another mechanism of cross-generational transmission of obesity risk. Interestingly, the differences became apparent when the mice were older, after 14 wk of age, suggesting that excess energy was stored as fat only after growth rate slowed down. Since food consumption was not different in offspring of dams with different parity, differences must have occurred in energy expended. Although we did not examine female offspring in these studies, some of the metabolic parameters measured may have been more exaggerated in the female mice based on previous reports by some (4, 34, 61), but not others (45).

We used a targeted gene approach to study metabolism of catabolic processes in two of the tissues where fatty acid oxidation occurs, the liver and adipose tissue (11, 67, 70). The lower CPT I mRNA levels in the offspring of multiparous dams in both tissues suggest that there is less fatty acid oxidation in the whole body, which has been shown to be related to obesity (38, 56). CPT I transcription was regulated differently in the multiparous offspring since expression of key metabolic regulators PGC-1α, PGC-1β, and PPARα was reduced as well. Since PGC-1α levels appear to be most important in mediating transcription compared with other proteins in the transcriptional complexes (64), reduction in its levels is likely enough to manipulate fatty acid oxidation. In addition to a direct effect on CPT I transcription, PGC-1α can manipulate fatty acid oxidation through effects on mitochondria (15, 21, 25). Although we did not measure mitochondrial activity or number in the current studies, we did find that mRNA levels for proteins expressed in the hepatic mitochondria of offspring of multiparous dams were reduced (cyp7a1: 2.72 ± 0.38 vs. 0.84 ± 0.08; citrate synthase: 4.47 ± 0.81 vs. 1.29 ± 0.11 for offspring of primiparous vs. multiparous females, respectively), suggesting a change in mitochondrial function.

PGC-1α is one of the key regulatory proteins in mediating cellular metabolism (reviewed in Refs. 15 and 25), including mitochondrial biogenesis and CPT I transcription. PGC-1α is a coregulator that interacts with various DNA-binding factors, including PPARα/γ, estrogen-related receptors, and nuclear receptor factors, to coordinate the regulation of a variety of genes and processes. PGC-1α will form complexes with these various transcription factors and recruit additional proteins that modify histones and remodel nucleosomes (49). PGC-1α levels and phosphorylation (activation)/acetylation (inactivation) appear to be regulated by the energy and/or nutritional status of the cells and are regulated by PPARs, myocyte enhancer factor 2, forkhead box O, cAMP response element-binding protein, estrogen-related receptors, the adiponectin receptors (adipoR1/R2), AMPK, and sirtuin (15, 25, 27, 71). Results from the current studies parallel studies in which pregnant females were overfed in that offspring of overfed mothers had reduced adipoR1/R2 expression and AMPK activity (7, 57), which can lead to less expression and activation of PGC-1α (27, 71).

PGC-1α levels can also be regulated epigenetically (3, 58). Previous studies have shown that a variety of conditions can methylate the promoter and lead to reduced mRNA levels. Epigenetic changes in genes have been proposed to be involved in the passing on of traits from one generation to another, and methylation of specific genes at birth can predict adiposity later in life (17). Although most common routes by which the epigenome is affected include DNA methylation and histone modification (72), it is likely that histone modification plays a major role in developmental programming via epigenetic regulation (47). The study of epigenetic regulation is complex, since some epigenetic changes can be silent and their action becomes apparent with different diets, stressors, or stages of life (16, 47, 65).

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Fig. 5. Plasma lipoprotein cholesterol levels in offspring of primiparous (1st) or multiparous (4th) dams; animals are the same as those described in Fig. 3. Plasma was collected from offspring at the termination of the study when they were 26 wk of age, and lipoproteins were separated by fast-protein liquid chromatography. Two pooled samples were used, and the results were averaged. Inset: total plasma triglyceride (TG) levels are presented.

Fig. 6. Glucose tolerance tests and insulin levels and signalling in offspring of multi- and primiparous dams; animals are the same as those described in Fig. 3. *Differences (P < 0.05) between plasma glucose levels at zero time; †linearly significant differences (P = 0.051) at 30 min postinjection. B: plasma insulin levels in offspring of dams of different parity. Data are presented as means ± SE. C: phosphorylated (Akt-p) and total Akt in pooled liver samples of offspring of dams with different parity.
One surprising observation of these studies was that FAS mRNA levels were decreased in adipose tissues and livers with greater adiposity, and the decreases were independent of changes in SREBP-1 and LXR mRNA levels, the typical route of regulation. Contrary to conventional thinking, recent studies show that PGC-1β and PGC-1α can be coactivators of SREBP-1/LXR and thereby affect their downstream targets independent of a change in SREBP-1/LXR levels (37, 60). Thus, the PGCs can affect the sum of lipid metabolism via both catabolic and anabolic pathways, with the greater impact upon oxidation under the current conditions.

An important outcome of these studies is their implication for mouse research. When efficiently maintaining mouse colonies such that parity of breeding females should be kept equal to minimize scatter and thereby increase significance. Thus, care should be taken when maintaining mice colonies such that parity of breeding females should be kept equal to minimize scatter and thereby increase significance.

Summary

A question that arises from these studies, as well as others, is, “What is mediating the programming?” The mediator of developmental programming is currently unknown but likely includes circulating maternal factors associated with obesity, inflammation, and/or diabetes and/or the placental inflammatory status (2, 9, 23, 44). Some maternal factors that are associated with these diseases have been proposed as potential mediators because they manipulate nutrient transport across the placenta or placental function, thereby affecting fetal metabolism (14, 23, 28, 29, 32, 33). Another potential mechanism is based on the fact that placentas of obese females contain more inflammation (10, 52, 59, 74), and the inflammatory in utero state that the fetus is exposed to may be mediating the effect.

In the current study, the placentas of the multiparous dams were in a more severe proinflammatory state than placentas of the primiparous dams, as depicted by increased JNK activation, and had more macrophages, as do placentas of obese humans, as depicted by increased CD68 and F4/80. Since the studies in mice parallel those in humans and developmental programming did indeed occur, we can use this model to begin to dissect out the role of various maternal factors in mediating programming.

To summarize, these studies support the propagation of the obesity cycle in multiparous females. Since more women are obese at the start of pregnancy, more retain weight during and between pregnancies, thereby developing multiparity-induced obesity. Thus, offspring of the obese multiparous dams with increased inflammation will themselves become obese in adulthood. Interventions targeted to break the obesity cycle could occur at the level of the pregnant female as well as the offspring. One could target inflammation of the pregnant female with various anti-inflammatory treatments, i.e., fish oil or exercise or antiobesity treatments, i.e., food restriction or exercise; it should be noted that the ω-3 fatty acids in fish oil could have a direct impact upon the fetus itself (43). In the offspring, one could target mitochondrial biogenesis or fatty acid oxidation with various factors, such as lipic acid or carnitine (35, 40). The current studies are important in supporting a healthier, less obese population in that we have defined specific metabolic pathways that are likely involved in the programming of obesity and can be targeted in either the mother or her offspring.

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.
AUTHOR CONTRIBUTIONS
S.L.R., T.J., K.T.B., A.J., and L.A.W. performed the experiments; S.L.R., T.J., K.T.B., A.J., P.T., and L.A.W. analyzed the data; S.L.R., T.J., A.J., and L.A.W. prepared the figures; S.L.R., T.J., A.J., P.T., D.A.D., and L.A.W. edited and revised the manuscript; A.J., P.T., D.A.D., and L.A.W. interpreted the results of the experiments; D.A.D. and L.A.W. did the conception and design of the research; L.A.W. drafted the manuscript; L.A.W. approved the final version of the manuscript.

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