IN MEN, HYPOGONADISM (i.e., reduced circulating testosterone) results in a variety of detrimental effects, including the loss of skeletal muscle mass and bone mineral density (BMD) and increased visceral adiposity (52). Testosterone replacement therapy has been proposed as a means of reducing these unfavorable changes (6). However, replacement doses of testosterone result in only minor improvements in muscle strength (39) and BMD (61) and only small reductions in adiposity (27). In contrast, supraphysiological testosterone effectively augments skeletal muscle mass (7, 8, 57), reduces adiposity (65), and increases BMD (2) in men and in orchiectomized animal models (10, 66) but also increases the risk for a variety of side effects, of which polycythemia, prostate enlargement, and increased incidence of prostate biopsy occur most frequently (13). Selective androgen receptor (AR) modulators (SARMs) are currently under development with the aim of producing anabolic effects in skeletal muscle and bone without prostate enlargement, polycythemia, or other androgenic side effects (33).

17β-Hydroxyestra-4,9,11-trien-3-one (trenbolone) is a potent synthetic testosterone analog (67) that does not undergo 5α-reduction to more potent metabolites (46). As such, trenbolone may induce less growth in prostate and other androgenic tissues that highly express 5α-reductase. This is in contrast to testosterone, which has approximately threefold greater potency in androgenic tissues that highly express 5α-reductase (63) due to its conversion to dihydrotestosterone (DHT).

The primary purpose of this study was to determine the effects of trenbolone-enantate (TREN; a slowly released trenbolone ester) on a variety of androgen-sensitive tissues, including skeletal muscle, bone, visceral adiposity, hemoglobin (HB), and the prostate of rodents. Because trenbolone is selectively metabolized to weaker androgens in vivo, we hypothesized that TREN will produce dose-dependent anabolic effects in skeletal muscle, bone, and fat that are at least equal to those of supraphysiological testosterone while producing a smaller increase in HB and less growth of the prostate.

METHODS

Animal Care

Barrier-raised and viral pathogen-free Fischer F344 male rats aged 3 mo were obtained from Charles River Laboratories (Wilmington, MA). Animals were housed individually in a temperature- and light-controlled room on a 12:12-h light-dark cycle. Rats were fed an ad libitum diet of Purina rodent chow containing 3.3 kcal/g distributed as 58.9% carbohydrate, 12.4% fat, and 28.7% protein (no. 5001; Purina Mills, St. Louis, MO) and tap water. All experimental procedures conformed to the Institute for Laboratory Animal Research Guide to the Care and Use of Experimental Animals and were approved by the Institutional Animal Care and Use Committee at the Gainesville Veterans Affairs Medical Center.

Address for reprint requests and other correspondence: J. F. Yarrow, VA Medical Center, Research - 151, 1601 SW Archer Rd., Gainesville, FL 32608-1197 (e-mail: jfyarrow@ufl.edu).
Pilot Study Experiment Design: Verify Hormone Delivery and Suppression of Endogenous Sex Hormones

A pilot study was performed to verify delivery of TREN (Steraloids, Newport, RI) following intramuscular administration and to evaluate the suppressive effects of TREN administration on circulating testosterone and DHT concentrations. For this experiment, intact 3-mo-old male Fisher F344 rats received intramuscular injections of either TREN (7.0 mg·wk⁻¹·animal⁻¹) or vehicle (sesame oil) at baseline and 7 days later (n = 5/group). Blood was sampled via tail tip amputation at baseline and every 2 days thereafter. Ten days following the initial injection, the rats were euthanized by an intraperitoneal injection of 120 mg/kg pentobarbital sodium, and blood was collected via cardiac puncture for the measurement of serum androgen concentrations.

Experimental Design Study 1: Muscle, Bone, Adipose, and Prostate Tissue Responses to Trenbolone Administration in Gonadectomized Male Rats

Male Fisher 344 rats, aged 4–5 mo, were divided into six groups (n = 10/group), including sham surgery plus vehicle (SHAM), orchiectomy (ORX) plus vehicle, ORX plus low-dose TREN (ORX + low TREN; 1.0 mg/wk), ORX plus moderate-dose TREN (ORX + mod TREN; 3.5 mg/wk), ORX plus high-dose TREN (ORX + high TREN; 7.0 mg/wk), or ORX plus supraphysiological testosterone-enanthate (ORX + TE; 7.0 mg/wk). We have reported previously that ORX induces catabolic effects in muscle, bone, and kidney and increases visceral fat accumulation (9), whereas supraphysiological TE administration prevents those effects but results in significant prostate enlargement (10, 11, 66). Blood was sampled via tail tip amputation prior to surgery and weekly thereafter. Food consumption was measured weekly. On day 29, rats were euthanized via intraperitoneal pentobarbital sodium (120 mg/kg) injection, and blood was collected via cardiac puncture for Hb analysis and hormone analyses. The levator ani/bulbocavernousus (LABC) muscle complex (an androgen-responsive skeletal muscle complex in rodents), soleus, plantaris, semimembranosus, retroperitoneal fat pads, left and right femurs and tibiae, kidney, and prostate were excised and weighed. With the semimembranosus, retroperitoneal fat pads, left and right femurs and LABC muscle complex and prostate were excised and weighed.

Hormone delivery. TREN and TE (Savient Pharmaceutical, East Windsor, NJ) were dissolved in sesame oil prior to intramuscular injection. Once weekly injections were alternated between the right and left quadriceps musculature and administered under brief isoflurane anesthesia. Sham surgery animals received vehicle sesame oil injections, where orchiectomized rats received either low (1.0 mg/wk), moderate (3.5 mg/wk), or high doses (7.0 mg/wk) of TREN, TE (7.0 mg/wk), or vehicle immediately following surgery and once every 7 days thereafter.

Hb and serum hormone analyses. Whole blood Hb was analyzed in duplicate immediately upon euthanization using the Hb Pro Professional Hemoglobin Testing System (ITC, Edison, NJ), which has a sensitivity of 4.0 g/dl and an intra-assay coefficient of variation (CV) of <3.3%. Serum was subsequently separated and stored at −80°C for further analysis. All hormones were assayed in duplicate using commercially available kits. Serum trenbolone was evaluated using qualitative enzyme-linked immunosorbent assay, which has a sensitivity of 0.1 ng/ml and an intra-assay CV of 3.76% (Neogen, Lexington, KY), and a standard curve was developed using trenbolone (Sigma-Aldrich, St. Louis, MO). Testosterone was assayed using a quantitative EIA kit (Diagnostic Systems Laboratories, Webster, TX) with a sensitivity of 0.04 ng/ml and an intra-assay covariance of 5.7%. DHT was assayed using a quantitative EIA kit (Alpco Diagnostics, Salem, NH) with a sensitivity of 6 pg/ml and an intra-assay covariance of 5.4%. Osteocalcin (a marker of bone formation) was determined with a Rat-Mid EIA, which has a sensitivity of 50 ng/ml and an intra-assay CV of <8% (Immunodiagnostic Systems, Fountain Hills, AZ). Bone resorption was measured by evaluating tartrate-resistant acid phosphatase form 5b (TRAP5b; an early marker of bone resorption) (1) with a RatTrap EIA (Immunodiagnostic systems), which has a sensitivity of 0.1 U/l and an intra-assay CV <5.8%, and by evaluating COOH-terminal telopeptide a1 chain of type I collagen (a late marker of bone resorption) with a RatLaps EIA (Immunodiagnostics Systems), which has a sensitivity of 2.0 ng/ml and an intra-assay CV <9.2%.

Bone morphometry and mechanical strength. Prior to peripheral quantitative computed tomography (pQCT), the femurs were thawed to room temperature and were kept in saline-soaked gauze except during measurement. The left femoral diaphysis and metaphysis were scanned by pQCT with a Stratec XCT Research M Instrument (Norland Medical Systems, Fort Atkinson, WI). Scans were performed at a distance of 5 (metaphysis) and 15 mm (diaphysis) proximal to the distal end of the femur for measurements of cancellous and cortical bone structure, respectively. The structural variables that were measured included total, trabecular, and cortical bone area (mm²), content (mg/mm), and density (mg/cm³).

Subsequent to pQCT, the midshaft of the left femora was subjected to a medial/lateral three-point bending test, using an MTS material testing machine (MTS Systems, Eden Prairie, MN) with methods we have reported previously (66). Before mechanical testing, a preload (10 N) was applied on the medial surface of the femur using a steel cross-bar fixture. The bending load was applied at 1.0 mm/s until failure. From the load-deformation curve, the following parameters were determined for the femoral shaft: breaking load, yield load, stiffness, and displacement. Bone mechanical strength is expressed as both force (measured in N) and stress (measured in Mpa).

Experimental Design Study 2: Prostate and LABC Responses to Trenbolone Administration in Intact Male Rats

Twenty male Fisher 344 rats, aged 3 mo, remained intact and were divided into the following four groups (n = 5/group): vehicle injection (SHAM), supraphysiological testosterone-enanthate (TE; 7.0 mg/wk), low-dose TREN (low TREN; 1.0 mg/wk), and one-half the low-dose TREN (half-Low TREN; 0.5 mg/wk). Animals remained intact to determine the effects of TREN on prostate and LABC masses and Hb concentrations when administered in the presence of endogenous sex hormones, a situation that simulates humans undergoing androgen replacement therapy. Once weekly intramuscular injections were delivered in a manner identical to that described in experiment 1. On day 29, rats were euthanized via intraperitoneal pentobarbital (120 mg/kg) injection, and blood was collected via cardiac puncture. The LABC muscle complex and prostate were excised and weighed. Whole blood Hb and serum testosterone, DHT, and trenbolone were analyzed using the methods and commercially available kits discussed previously (see experiment 1). For this experiment, the low TREN was a dose identical to that administered in experiment 1, a dose that produced robust muscle growth and preservation of BMD in ORX rats while maintaining prostate mass at the level of SHAMS. Additionally, TREN was administered in a dose lower than what was administered in experiment 1 (i.e., half-low TREN) to further evaluate the dose-response effects of TREN within skeletal muscle and prostate.

Statistical Analysis

Results are reported as means ± SE, and P < 0.05 was defined as the threshold of significance. One-Way ANOVAs (for normally dis-
RESULTS

Pilot Experiment: Verification of Hormone Delivery and Suppression of Endogenous Sex Hormones

Intramuscular TREN administration elevated serum trenbolone concentrations throughout the 7 days following injection (Fig. 1). On day 7, rats received an additional TREN injection, which resulted in peak trenbolone concentrations (46.0 ± 5.6 ng/ml) occurring on day 9. Thus, once weekly TREN injections resulted in a sustained elevation of serum trenbolone, with peak concentrations occurring within 48 h. Within 10 days of administration, TREN also suppressed serum testosterone of intact animals by 80% (2.55 ± 0.46 vs. 0.49 ± 0.07, P ≤ 0.05) and was elevated 18-fold following TE administration (9,774 ± 20.02 ng/ml). Serum DHT was reduced 70% with ORX (644 ± 72 vs. 187 ± 7, P ≤ 0.05) compared with intact vehicle-treated animals.

Experiment 1: Study on Muscle, Bone, Adipose, and Prostate Tissue Responses to Trenbolone Administration in Gonadectomized Male Rats

Serum androgen concentrations. Increasing doses of TREN resulted in a progressive increase in serum trenbolone (P ≤ 0.001; Fig. 2A). Serum testosterone was 1.9 ± 0.1 ng/ml in SHAMs (Fig. 2B) and was elevated 18-fold following TE administration (34.7 ± 2.5, P ≤ 0.01). Testosterone was low to undetectable following ORX (0.04 ng/ml) and in all TREN treatment groups (ORX + low TREN: 0.04 ng/ml; ORX + mod TREN: 0.07 ± 0.03 ng/ml; ORX + high TREN: 0.06 ± 0.02 ng/ml). Serum DHT was 284 ± 33 pg/ml in SHAMs (Fig. 2C) and was elevated 34-fold by TE administration (9,774 ± 522 pg/ml, P ≤ 0.001). DHT was reduced 70% with ORX compared with SHAMs (86 ± 7 pg/ml, P ≤ 0.001) and remained reduced following all TREN treatments (ORX + low TREN: 42 ± 7 pg/ml; ORX + mod TREN: 16 ± 6 pg/ml; ORX + high TREN: 7 pg/ml).

Weight gain, food consumption, and adiposity. During the first 3 days following surgery, ORX animals experienced a 75% reduction in weight gain compared with SHAMs, and weight gain remained lower than SHAMs throughout the duration of the study (P ≤ 0.01; Fig. 3A). Both TE and TREN (at all doses) restored weight gain during the first 3 days following ORX. However, after the initial 3-day post-surgery period, total weight gain was gradually reduced in the TE- and TREN-treated groups such that at euthanization the weight gain in all androgen-treated groups was 30–50% lower than in SHAMs (P ≤ 0.001) and not different from ORX (SHAM: 41.6 ± 2.5 g; ORX: 27.6 ± 1.9 g; ORX + TE: 25.7 ± 2.8 g; ORX + low TREN: 29.7 ± 1.5 g; ORX + mod TREN: 25.1 ± 1.1 g; ORX + high TREN: 20.7 ± 1.8 g). No differences in total weight gain were present between the TE and low and high-dose TREN.

TREN: 93 ± 5 pg/ml; ORX + mod TREN: 150 ± 5 pg/ml; ORX + high TREN: 182 ± 6 pg/ml.

Weight gain, food consumption, and adiposity. During the first 3 days following surgery, ORX animals experienced a 75% reduction in weight gain compared with SHAMs, and weight gain remained lower than SHAMs throughout the duration of the study (P ≤ 0.01; Fig. 3A). Both TE and TREN (at all doses) restored weight gain during the first 3 days following ORX. However, after the initial 3-day post-surgery period, total weight gain was gradually reduced in the TE- and TREN-treated groups such that at euthanization the weight gain in all androgen-treated groups was 30–50% lower than in SHAMs (P ≤ 0.001) and not different from ORX (SHAM: 41.6 ± 2.5 g; ORX: 27.6 ± 1.9 g; ORX + TE: 25.7 ± 2.8 g; ORX + low TREN: 29.7 ± 1.5 g; ORX + mod TREN: 25.1 ± 1.1 g; ORX + high TREN: 20.7 ± 1.8 g). No differences in total weight gain were present between the TE and low and high-dose TREN.
mod TREN treatments, although weight gain was 30% lower following high TREN treatment compared with low TREN ($P \leq 0.05$).

ORX also reduced total food consumption throughout the experiment by 14% when compared with SHAMs ($P \leq 0.01$), and this reduction was not prevented by any androgen treatment (data not shown). Change in body mass was subsequently corrected for total food consumption to determine whether food consumption influenced the observed differences in weight gain. Following this correction, the change in body mass gain corrected for food intake was similarly reduced in all ORX groups compared with SHAM. Values are means ± SE; $n = 9–10$ group. Letters indicate differences from respectively labeled groups at $P < 0.05$ or $*P < 0.01$ (vs. SHAM; a vs. ORX; a vs. ORX + TE; *vs. ORX + low TREN; *vs. ORX + mod TREN).

Prostate. ORX resulted in an 80% reduction in prostate mass compared with SHAMs ($P \leq 0.001$; Fig. 4C). TE and high TREN administrations increased prostate mass by 84% ($P \leq 0.001$) and 68% ($P \leq 0.01$), respectively, compared with SHAMs, whereas, neither low nor mod TREN elevated prostate mass above SHAMs. Ultimately, the low TREN resulted in a prostate that was 34% smaller than TE treatment ($P \leq 0.05$). No differences in prostate mass were present between the high TREN and TE treatments.

Kidney and Hb. ORX reduced kidney mass by 10% compared with SHAMs ($P \leq 0.001$; Table 1). TE administration prevented this reduction and further increased kidney mass by 14% compared with SHAMs ($P \leq 0.001$). Similarly, low TREN completely prevented the ORX-induced loss of kidney mass ($P \leq 0.001$), whereas both mod and high TREN increased kidney mass by ~19% compared with SHAMs ($P \leq 0.001$).

No differences in whole blood Hb concentrations were observed between the SHAM, ORX, ORX + TE, or ORX + low TREN animals (Table 1). However, both mod and high TREN administration elevated Hb by ~8–10% compared with SHAMs and ORX ($P \leq 0.05$).

BMD and bone mechanical strength. ORX resulted in a 12–14% reduction in total bone mineral content (BMC) and total BMD and also reduced trabecular (t)BMC and tBMD by ~50% at the distal femoral metaphysis compared with shams ($P \leq 0.001$; Table 2). Conversely, TE administration prevented the ORX-induced reductions at this skeletal site, ultimately
compared with ORX. No differences were present between TE and any TREN treatments for the bone mineral measurements at this skeletal site, although TREN treatment did not completely restore tBMC and tBMD to SHAM values (P < 0.05). The total and trabecular tissue areas remained unaltered at the distal femoral metaphysis, and no differences were present at the femoral diaphysis for any measurement (data not shown). Additionally, the femoral mass and length remained unaltered by all treatments, and no statistically significant differences were present between any bone mechanical measurements at the femoral midshaft (Table 2).

**Systemic markers of bone remodeling.** One week following surgery, serum Trap5b [an early marker of bone resorption (1)] was 9.4 ± 0.6 (SHAM), 11.7 ± 0.9 (ORX), 10.2 ± 0.4 (ORX + low TREN), 8.2 ± 0.4 (ORX + mod TREN), 7.6 ± 0.4 (ORX + high TREN), and 9.4 ± 0.8 (ORX + TE) (Fig. 5A). ORX resulted in a nonsignificant 25% increase in Trap5b, which was completely prevented by TE. Low TREN also resulted in a nonsignificant 15% reduction in Trap5b compared with ORX, whereas both mod TREN and high TREN reduced Trap5b by 30% compared with ORX (P < 0.01). At euthanization, serum C-telopeptide (a late marker of bone resorption) was 33.3 ± 2.6 (SHAM), 36.3 ± 2.3 (ORX), 30.0 ± 2.2 (ORX + low TREN), 32.0 ± 2.6 (ORX + mod TREN), 26.5 ± 2.7 (ORX + high TREN), and 33.7 ± 2.2 (ORX + TE). High TREN reduced serum C-telopeptide by 27% compared with ORX (P ≤ 0.05), with no other significant differences present between groups (Fig. 5B). At euthanization, serum osteocalcin (a marker of bone formation) was 679 ± 56 (SHAM), 892 ± 95 (ORX), 435 ± 42 (ORX + low TREN), 320 ± 30 (ORX + mod TREN), 282 ± 26 (ORX + high TREN), and 380 ± 48 (ORX + TE) (Fig. 5C). ORX resulted in a nonsignificant 31% increase in serum osteocalcin compared with SHAMs. All androgen treatment groups reduced serum osteocalcin by 36–58% compared with SHAMs (P ≤ 0.01) and by 51–68% compared with ORX (P ≤ 0.001). No differences were present between TREN- and TE-treated groups. In addition, serum osteocalcin was 26% lower in mod TREN (P = 0.063, trend) and 35% lower in high TREN (P ≤ 0.05) groups compared with the low TREN.

**Experiment 2: Study on Prostate and LABC Responses to Trenbolone Administration in Intact Male Rats**

**Hormone responses.** At euthanization, serum trenbolone was 13.7 ± 0.6 ng/ml in intact low TREN-treated animals and 4.8 ± 0.3 ng/ml in half-low TREN-treated animals (P < 0.01; Fig. 6A). Serum testosterone was 5.7 ± 1.5 ng/ml in SHAM animals and was increased ninefold by TE administration (53.0 ± 4.5 ng/ml, P < 0.01; Fig. 6B). Serum testosterone was reduced to nearly undetectable concentrations by both low TREN (0.4 ± 0.1 ng/ml) and half-low TREN (0.3 ± 0.1 ng/ml).

**Table 1. Effects of ORX, TE, or graded doses of TREN on kidney mass and Hb concentrations**

<table>
<thead>
<tr>
<th></th>
<th>SHAM</th>
<th>ORX</th>
<th>ORX + TE</th>
<th>ORX + Low TREN</th>
<th>ORX + Mod TREN</th>
<th>ORX + High TREN</th>
</tr>
</thead>
<tbody>
<tr>
<td>Kidney, g</td>
<td>0.89 ± 0.01</td>
<td>0.79 ± 0.01**</td>
<td>1.00 ± 0.01**</td>
<td>0.95 ± 0.01**</td>
<td>1.05 ± 0.02**</td>
<td>1.06 ± 0.02**</td>
</tr>
<tr>
<td>Hb, mg/dl</td>
<td>15.6 ± 0.2</td>
<td>15.7 ± 0.2</td>
<td>16.6 ± 0.2</td>
<td>167.3 ± 0.3</td>
<td>173.4 ± 0.4**</td>
<td>170.0 ± 0.3**</td>
</tr>
</tbody>
</table>

Values are means ± SE; n = 9–10/group. ORX, orchiectomy; TE, testosterone-enanthate; TREN, trenbolone-enanthate; SHAM, vehicle group; low, mod, and high TREN, low-, moderate- and high-dose TREN groups, respectively. Letters indicate differences from respectively labeled groups at P < 0.05 or *P < 0.01 (vs. SHAM; vs. ORX; vs. ORX + TE; vs. ORX + low TREN; vs. ORX + mod TREN).
treatments (P < 0.01). Serum DHT was 962 ± 198 pg/ml in SHAM animals and was increased 13-fold in TE-treated animals (12,288 ± 1,037 pg/ml, P < 0.01), whereas DHT was reduced below SHAM values by low TREN treatment (126 ± 4 pg/ml, P < 0.01) and below both sham and low TREN treatment values by the half-Low TREN treatment (88 ± 2 pg/ml, P < 0.01) (Fig. 6C).

Weight gain. At baseline, body mass was similar among all groups (SHAM: 270.7 ± 2.6 g; TE: 270.7 ± 3.7 g; low TREN: 271.6 ± 2.2 g; half-Low TREN: 271.4 ± 4.9 g). Body mass

Table 2. Effects of ORX, TE, or graded doses of TREN on femoral anthropometric measurements, metaphyseal BMC and BMD, and midshaft bone mechanical characteristics

<table>
<thead>
<tr>
<th></th>
<th>SHAM</th>
<th>ORX</th>
<th>ORX + TE</th>
<th>ORX + Low TREN</th>
<th>ORX + Mod TREN</th>
<th>ORX + High TREN</th>
</tr>
</thead>
<tbody>
<tr>
<td>Femoral mass, g</td>
<td>0.78 ± 0.01</td>
<td>0.75 ± 0.01</td>
<td>0.78 ± 0.01</td>
<td>0.77 ± 0.01</td>
<td>0.77 ± 0.01</td>
<td>0.77 ± 0.01</td>
</tr>
<tr>
<td>Femoral length, mm</td>
<td>37.1 ± 0.1</td>
<td>36.4 ± 0.2</td>
<td>36.8 ± 0.2</td>
<td>36.9 ± 0.1</td>
<td>36.8 ± 0.1</td>
<td>36.7 ± 0.2</td>
</tr>
<tr>
<td>Total BMC, mg/mm3</td>
<td>12.7 ± 0.3</td>
<td>10.9 ± 0.3a</td>
<td>12.1 ± 0.4a</td>
<td>11.5 ± 0.3a</td>
<td>11.9 ± 0.3</td>
<td>11.9 ± 0.2</td>
</tr>
<tr>
<td>Total BMD, mg/mm3</td>
<td>668 ± 6</td>
<td>587 ± 8**</td>
<td>657 ± 10**</td>
<td>624 ± 7ab</td>
<td>632 ± 8b*a</td>
<td>641 ± 9b**</td>
</tr>
<tr>
<td>Trabecular BMC, mg/mm3</td>
<td>2.0 ± 0.1</td>
<td>1.0 ± 0.1**</td>
<td>1.6 ± 0.1**</td>
<td>1.4 ± 0.1**</td>
<td>1.5 ± 0.1**a</td>
<td>1.5 ± 0.1b**</td>
</tr>
<tr>
<td>Trabecular BMD, mg/cm3</td>
<td>344 ± 16</td>
<td>177 ± 13**</td>
<td>288 ± 17**</td>
<td>243 ± 15ab</td>
<td>257 ± 14ab,a</td>
<td>274 ± 16ab,b</td>
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<tr>
<td>Breaking load, N</td>
<td>136 ± 12</td>
<td>134 ± 11</td>
<td>138 ± 7</td>
<td>139 ± 11</td>
<td>139 ± 10</td>
<td>137 ± 10</td>
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<tr>
<td>Yield load, N</td>
<td>93 ± 17</td>
<td>90 ± 20</td>
<td>93 ± 12</td>
<td>90 ± 17</td>
<td>116 ± 16</td>
<td>98 ± 20</td>
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<tr>
<td>Stiffness, N/mm</td>
<td>245 ± 32</td>
<td>227 ± 30</td>
<td>270 ± 25</td>
<td>257 ± 30</td>
<td>253 ± 26</td>
<td>256 ± 34</td>
</tr>
</tbody>
</table>

Values are means ± SE; n = 9–10/group. BMC, bone mineral content; BMD, bone mineral density. Letters indicate differences from respectively labeled groups at P < 0.05 or *P < 0.01 (avs. SHAM; bvs. ORX; cvs. ORX + TE; dvs. ORX + low TREN).
gain was reduced to a similar degree by all androgen treatments, which became significant on day 21 of the experiment (P < 0.05; Fig. 7A). Combined retroperitoneal and perirenal (visceral) fat mass was also reduced 25% by TE treatment (P < 0.05) and was nonsignificantly reduced 13–16% by both the low and half-low TREN treatments (Fig. 7B).

Prostate and LABC. In intact animals, TE treatment increased prostate mass by 59% compared with SHAMs (P < 0.001; Fig. 8A). Low TREN maintained prostate mass at the level of SHAMs, ultimately resulting in a prostate that was 31% smaller than following TE treatment (P < 0.001). Half-low TREN treatment resulted in a nonsignificant 22% reduction in prostate mass compared with SHAMs, a 29% reduction compared with low TREN treatment (P < 0.01), and a 51% reduction compared with TE treatment (P < 0.001).

Mass of the LABC muscle complex was increased by 21–38% in TE- and TREN-treated animals (P < 0.01; Fig. 8B), with no differences present between any androgen treatment. The LABC/prostate ratio, which illustrates the anabolic/androgenic ratio of androgens, was unaltered by TE treatment (Fig. 8C). Following Low TREN treatment, the LABC/prostate ratio was increased by 12% compared with SHAMs (P < 0.05) and by 25% compared with TE treatment (P < 0.01). Ultimately, half-low TREN treatment resulted in the highest LABC/prostate ratio, a value that was 53% higher than low TREN treatment (P < 0.05), 72% higher than SHAMs (P < 0.01), and 92% higher than TE treatment (P < 0.01).

Kidney and Hb. No differences in kidney mass (sham: 0.93 ± 0.02 g; TE: 1.01 ± 0.01 g; low TREN: 0.98 ± 0.02 g; half-low TREN: 0.94 ± 0.04 g) or Hb concentrations (sham: 14.6 ± 0.2 mg/dl; TE: 15.7 ± 0.2 mg/dl; low TREN: 15.5 ± 0.4 mg/dl; half-low TREN: 15.4 ± 0.4 mg/dl) were observed between groups.
DISCUSSION

Our results demonstrate that TREN prevents the deleterious alterations in body composition associated with ORX to the same extent as supraphysiological TE. Specifically, we observed that 1) regardless of dose, TREN and supraphysiological TE produced equally myotrophic responses in the androgen-sensitive LABC muscle complex in both intact and ORX animals, 2) TREN partially prevented ORX-induced bone loss to roughly the same extent as supraphysiological TE, and 3) at equal doses, TREN was somewhat more lipolytic than TE in visceral fat. In contrast, the lowest doses of TREN maintained prostate mass and Hb concentrations at the level of shams in both intact and orchietomized animals, whereas supraphysiological TE and high-dose TREN produced prostate enlargement and/or elevations in Hb. Thus, at the lowest doses administered, TREN appears to have a higher and more clinically favorable anabolic/androgenic ratio than supraphysiological TE.

Skeletal muscle expresses ARs to varying degrees among specific muscle groups and among species. For example, humans respond robustly to androgenic stimuli due to the high percentages of AR-positive myonuclei (54). Conversely, certain rodent peripheral muscles respond poorly to androgenic stimuli due to the their low concentrations of AR-positive myonuclei (e.g., extensor digitorum longus with 7% AR-positive myonuclei) (34), whereas the rodent LABC muscle complex contains ~74% AR-positive myonuclei and experiences a substantial atrophic response to androgen ablation and a robust myotrophic response to androgen administration (34). In our study, the lower limb muscles were insensitive to androgen status; however, ORX reduced LABC muscle mass, whereas TREN and TE robustly augmented LABC muscle mass. These results demonstrate clearly that TREN is at least as myotrophic as supraphysiological TE within androgen-sensitive skeletal muscle because even the lowest TREN doses produced equal growth of the LABC muscle in intact and ORX animals compared with supraphysiological TE. The mechanism(s) underlying the TREN-induced augmentation of skeletal muscle mass requires further clarification (67), although it is suspected that TREN acts via 1) AR activation, since trenbolone possesses three times the affinity of testosterone for the AR (5), 2) upregulation of endogenous growth factors (e.g., IGF-I or fibroblast growth factor) and/or increased responsiveness of skeletal muscle to such growth factors (29, 60), and/or 3) anticatabolic mechanisms associated with reductions in endogenous glucocorticoid activity (48, 49).

In our study, androgen treatment [either TE or TREN (at all doses)] prevented the initial weight reduction associated with surgical ORX to a similar extent. However, at euthanization, body weight gain was similarly reduced among all ORX groups compared with SHAMs, regardless of androgen treatment. Similarly, all intact androgen-treated animals experienced a reduced body weight gain compared with SHAMs. It remains unclear why TE did not maintain body weight gain throughout these studies, as we have reported previously (66). The reduced weight accrual may be at least partially explained by the lower food consumption within all ORX groups throughout the duration of experiment 1. However, the reduced food consumption did not cause a measurable loss of lower limb skeletal muscle mass. Conversely, we observed that ORX increased retroperitoneal fat mass, albeit nonsignificantly, and that visceral fat mass was reduced by TREN in a dose-dependent manner. Furthermore, at equal doses, TREN induced a greater magnitude of fat loss than TE. These results are somewhat surprising considering that all androgen treatments produced similar myotrophic growth of the androgen-sensitive LABC muscle complex and induced a substantial and roughly equivalent prevention of the ORX-induced BMD loss. Thus, it
appears that TREN is capable of preventing the initial catabolic responses associated with ORX while inducing more potent lipolytic effects within visceral adiposity than endogenous androgens, at least on a dose-to-dose equivalent. Future research examining the mechanisms underlying the lipolytic effects of TREN is appealing considering the preponderance of health decrements associated with central adiposity (15).

Clearly, androgens influence the development of male secondary sexual characteristics, skeletal muscle, bone, and fat, among other tissues (59). Testosterone is the primary circulating endogenous androgen that produces biological responses following nuclear interactions with ARs or through non-genomic signaling pathways (19). However, in tissues that highly express 5α-reductase (e.g., prostate), testosterone is transformed to its more potent metabolite DHT, which increases its androgenic potency within these tissues (63). As such, prostate enlargement is a concern associated with testosterone replacement therapy in hypogonadal men (13). In our studies, the lowest doses of TREN did not induce prostate enlargement in ORX or intact animals, whereas supraphysiological TE and the highest TREN dose increased prostate mass to a similar magnitude. These results are interesting considering that TREN binds to ARs with approximately three times the affinity of TE and an affinity roughly equal to that of DHT, the most potent endogenous androgen. In contrast to testosterone, certain anabolic steroids (e.g., 19-nortestosterone) are transformed to less potent metabolites by 5α-reductase or are not substrates for 5α-reductase (e.g., 7α-methyl-19-nortestosterone) due to slight variations in the structures of these compounds (58). Similarly, the reduced prostate enlargement associated with TREN appears to result from the presence of a 3-oxotriene structure that prevents A-ring reduction of this steroid (46), ultimately limiting its ability to undergo 5α-reduction or aromatization. As a result, trenbolone is primarily metabolized to its less potent 17α-isofrom (epitrenbolone) or trenbolone in humans (55) and in other mammalian species (17, 31, 45, 46), underly its equal potency in tissues that highly express 5α-reductase (e.g., prostate) vs. those that do not (e.g., skeletal muscle) (67).

Another concern with androgen replacement therapy is the potential to initiate or accelerate the growth of undiagnosed prostate cancer, although several recent meta-analyses have failed to detect an increased risk of prostate cancer following testosterone replacement in hypogonadal men (13, 18). In our studies, TREN administration resulted in a nearly complete ablation of circulating testosterone and DHT, the primary endogenous androgens implicated in prostate growth, through mechanisms apparently associated with feedback inhibition; however, we did not directly evaluate the possibility that TREN may initiate or accelerate prostate cancer growth. Several previous reports indicate that trenbolone appears to produce little genotoxic activity and is not an initiator of cancer in various in vitro models (47, 51), although trenbolone has been shown capable of inducing morphological changes in the Syrian hamster embryo cell transformation assay (an in vitro mutagenicity model) (50, 51). As such, future research examining the effects of TREN and other androgens using in vitro and/or in vivo xenograft models of androgen-responsive prostate cancer is warranted and should be conducted prior to this agent being advanced to clinical testing.

Certain androgens also undergo in vivo aromatization (e.g., the conversion of testosterone to estradiol) and subsequently exert systemic or tissue-specific estrogen receptor-mediated effects (28). The aromatization of testosterone appears to be essential for bone development in men (62), as demonstrated by the identification of several men who suffer from congenital aromatase deficiency resulting in osteopenia, which is treatable with estradiol but not testosterone (28). Similarly, male aromatase knockout mice develop severe osteopenia and adverse skeletal development, which is reversible with estradiol administration (37, 41, 43). Conversely, a number of side effects commonly associated with supraphysiological androgen administration are influenced by excessive systemic or tissue-specific aromatization of androgens, including gynececomastia (12), prostate cancer (14), fluid retention (56), and premature epiphyseal closure (42). We were unable to measure circulating estrogens in our study, since the currently available methods lack the sensitivity required for this analysis in orchietomized male rodents. However, in vitro evidence indicates that trenbolone and its primary metabolites are relatively nonestrogenic (30), and in vivo evidence demonstrates that trenbolone induces antiestrogenic effects in oviparous species (3, 16, 23, 32, 38, 53). Additionally, it has been reported that trenbolone does not undergo aromatization due to its 3-oxotriene structure (46), although conflicting reports exist regarding the influence of trenbolone on circulating estrogens in mammalian species (24, 26). It is plausible that the antiestrogenic effects and apparently nonaromatizable aspects of trenbolone partially underlie the slightly diminished bone-protective effects and the apparent adynamic bone state (i.e., reduced markers of bone resorption and bone formation) that we observed following TREN treatment of ORX animals. These results may provide further evidence indicating that the bone-protective effects of androgens are at least partially mediated by the systemic and/or tissue-specific aromatization to estrogens. Clearly, future research examining bone histomorphometry would provide further insight into the effects of TREN on bone turnover. Regardless, our findings indicate that androgens are capable of exerting direct bone protection, as evidenced by the TREN-induced partial prevention of BMD loss and the reversal of ORX-induced increases in serum markers of bone resorption and formation, results that ultimately assisted in the maintenance of bone strength. As such, future research evaluating the mechanism(s) underlying the bone-protective effects of TREN and other nonaromatizable androgens are warranted, especially considering the influence of estrogens on bone health and on the side effects associated with excessive androgen administration.

Recently, SARMs that produce anabolic responses within muscle and bone without inducing prostate enlargement or other androgenic side effects have been developed (33). The mechanism(s) through which steroidal and nonsteroidal SARMs produce selective tissue-specific anabolic responses has not been fully elucidated, although it has been postulated that their reduced activity within the prostate and other androgenic tissues may result from their inability to undergo tissue-specific 5α-reduction to more potent metabolites (22). Our results demonstrate that TREN, a non-5α-reducible testosterone analog, exhibits SARM-like properties since it prevented the ORX-induced reductions in muscle and bone and also reduced fat mass while maintaining prostate mass and Hb at the
level of intact animals. In fact, the half-low TREN treatment reduced prostate mass in intact animals, albeit nonsignificantly, whereas it increased LABC muscle mass, which corroborates previous work indicating that TREN remains myotrophic when administered at doses lower than in our current study (4, 20, 21, 35, 36, 40), and may also be capable of reducing prostate mass in developing animals (25, 64). The likely mechanisms underlying these findings are that 1) TREN ablates circulating testosterone and DHT in intact animals and 2) TREN undergoes tissue-specific biotransformation to less potent androgens in vivo. The long-term SARM-like potential of TREN remains to be determined, as does its anabolic/androgenic effects within older hypogonadal rodents, a model that may more appropriately mimic the effects of aging and/or hypogonadism in men.

In conclusion, administration of TREN, a potent non-5α-reducible and nonestrogenic synthetic testosterone analog, produces robust myotrophic effects, partial inhibition of bone loss, prevention of visceral fat accumulation, and maintenance of Hb and prostate mass at the level of intact animals, at least at the lowest dose administered. These results suggest that lower-dose TREN induces favorable SARM-like effects on musculoskeletal tissue and adiposity and within specific accessory sex organs. Future research examining the safety and efficacy of this androgen in preclinical settings appears to be appropriate because the anabolic/androgenic ratio appears to be higher and because the risk/benefit ratio appears to be less than that of supraphysiological TE, at least in regard to prostate enlargement, although evaluating TREN and other SARMs in models of androgen-responsive prostate cancer is necessary prior to these agents being recommended for clinical testing.

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DISCLOSURES
No conflicts of interest, financial or otherwise, are declared by the authors.

REFERENCES
TISSUE SELECTIVITY OF TRENBOLENE


