Palmitic acid acutely inhibits acetylcholine- but not GLP-1-stimulated insulin secretion in mouse pancreatic islets

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Doliba NM, Qin W, Vinogradov SA, Wilson DF, Matschinsky FM. Palmitic acid acutely inhibits acetylcholine-stimulated insulin secretion in mouse pancreatic islets. Am J Physiol Endocrinol Metab 299: E475–E485, 2010. First published July 6, 2010; doi:10.1152/ajpendo.00072.2010.—Fatty acids, acetylcholine, and GLP-1 enhance insulin secretion in a glucose-dependent manner. However, the interplay between glucose, fatty acids, and the neuroendocrine regulators of insulin secretion is not well understood. Therefore, we studied the acute effects of PA (alone or in combination with glucose, acetylcholine, or GLP-1) on isolated cultured mouse islets. Two different sets of experiments were designed. In one, a fixed concentration of 0.5 mM of PA bound to 0.15 mM BSA was used; in the other, a PA ramp from 0 to 0.5 mM was applied at a fixed albumin concentration of 0.15 mM so that the molar PA/BSA ratio changed within the physiological range. At a fixed concentration of 0.5 mM, PA markedly inhibited acetylcholine-stimulated insulin release, the rise of intracellular Ca2+, and enhancement of cAMP production but did not influence the effects of GLP-1 on these parameters of islet cell function. 2-ADB, an IP3 receptor inhibitor, reduced the effect of PA and did not influence the effects of GLP-1 on these parameters of islet cell function. 2-ADB, an IP3 receptor inhibitor, reduced the effect of PA and did not influence the effects of GLP-1 on these parameters of islet cell function.

In addition to nutrients, insulin secretion is stimulated by many hormones and neurotransmitters. Acetylcholine, the neurotransmitter of the parasympathetic nervous system, plays a significant role in the regulation of insulin secretion in pancreatic β-cells (1, 13, 38). Mutant mice selectively lacking the M3 muscarinic acetylcholine receptor subtype in pancreatic β-cells display impaired glucose tolerance and greatly reduced insulin release (12, 44). In contrast, transgenic mice overexpressing M3 receptors in pancreatic β-cells show a pronounced improvement in glucose tolerance and insulin secretion and are resistant to diet-induced glucose intolerance and hyperglycemia (11). The secretory response of β-cells to fuel stimulation is also markedly enhanced by the gut hormone glucagon-like peptide-1 (GLP-1), a member of the glucagon family, which is released into the portal circulation when a meal is digested (17). Both GLP-1 and acetylcholine greatly increase the sensitivity of β-cells to nutrients, but their effects on secretion depend entirely on availability, levels, and compositions of metabolic substrates. However, to our knowledge, little is known about the interplay between FA metabolism and the neuroendocrine regulation of insulin secretion. Therefore, the goal of the present study was to evaluate the acute action of palmitic acid (PA; present alone or in combination with glucose and/or acetylcholine or GLP-1) on isolated mouse islets. The data demonstrate that PA interferes with the cholinergic regulation of β-cell function, resulting in inhibition of glucose-dependent acetylcholine but not GLP-1 stimulation of insulin secretion.

RESEARCH DESIGN AND METHODS

Animals. Our research was reviewed and approved by the Institutional Animal Care and Use Committee of the University of Pennsylvania (protocol no. 077603). B6D2F1 mice were used throughout. The mice were maintained on a 12:12-h light-dark cycle and were fed a standard rodent chow diet.

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**Islet isolation.** Mouse islets were isolated using collagenase (EC 3.4.24.3 Serva, 17449) digestion in Hanks’ buffer, followed by separation of islets from exocrine tissue in a Ficoll (F-9378; Sigma) gradient. Isolated islets were cultured for 3–4 days in RPMI 1640 medium (Sigma) containing 10% fetal bovine serum, 10 ml/l penicillin-streptomycin-amphotericin B solution (GIBCO-BRL), and 10 mM glucose.

**Preparation of sodium palmitate solution.** A 5 mM stock solution of sodium palmitate (Sigma-Aldrich) was prepared by dissolving the FA salt in 10% bovine serum albumin (BSA; fraction V, FA free; Sigma-Aldrich) in Krebs buffer by stirring continuously for 4 h in a 37°C water bath. The stock solution was then diluted by Krebs buffer to obtain the final concentration of 0.5 mM sodium palmitate and 1% BSA.

**Perfusion of islets for measurement of insulin release.** Cultured islets (130 islets) were placed on a nylon filter in a plastic perfusion chamber (Millipore, Bedford, MA) and were perfused with a flow rate of 2 ml/min. The perfusion apparatus consisted of a computer-controlled fast-performance HPLC system (Waters 625 LC System) that allowed programmable rates of flow and glucose concentration in the perfusate, a water bath (37°C), and fraction collector (Waters Division.

Fig. 1. Effects of palmitic acid (PA) on acetylcholine (Ach)- and glucagon-like peptide-1 (GLP-1)-stimulated insulin release. A: an Ach ramp from 0 to 1 μM (12.5 nM increment/min) was applied after islet preperfusion with 0, 4, and 8 mM glucose (G) for 30, 20, and 20 min, respectively (●). In 1 set of experiments, PA was added after 20 min of preperfusion with no glucose added and then was present during the entire experiment (○). PA significantly affected Ach potentiation of insulin release (from 69 to 144 min of perifusion, P < 0.05) starting at Ach concentration of 62.5 nM. B: a GLP-1 ramp from 0 to 10 nM was applied using the same experimental design as presented in A. C: insulin release was stimulated initially with 4 and 8 mM G, followed by stimulation with Ach (0.5 μM). Afterward, a PA ramp from 0 to 0.5 mM (12.5 μM increment/min; ●) was applied for 40 min. In control experiments, islets were perfused with 0.5 μM of Ach (○) for the same duration as the PA ramp. PA significantly affected Ach potentiation of insulin release (from 97 to 154 min of perifusion, P < 0.05), starting at a PA concentration of 50 μM. D: GLP-1 (10 nM) instead of Ach was used to potentiate insulin secretion with the same experimental design as that presented in C. Note that here and in all other experiments, 10 μM neostigmine, a cholinesterase inhibitor, and 100 μM valine-pyrrolidide, a dipeptidyl peptidase-4 inhibitor, were used to prevent breakdown of Ach and GLP-1, respectively. Each curve represents the mean ± SE of 3–4 perfusions.
of Millipore). The perfusate was Krebs buffer (pH 7.4) containing 114 mmol/l NaCl, 5 mmol/l KCl, 24 mmol/l NaHCO$_3$, 1 mmol/l MgCl$_2$. 6H$_2$O, 2.2 mmol/l Ca$^{2+}$, 1 mM Pi, 10 mmol/l HEPES (pH 7.4), 1% BSA (fraction V, FA free; Sigma-Aldrich) equilibrated with 20% O$_2$, and 5% CO$_2$ balanced with N$_2$.

Perfusion of islets for simultaneous measurement of oxygen consumption and insulin release. The islets (600 islets) were loaded into the 250-μl chamber using a P200 pipette and gel-loading tip and were allowed to settle for 1 min before the flow was resumed at 80 μl/min. The perfusion apparatus consisted of a peristaltic pump, a water bath (37°C), a gas exchanger (artificial lung; medium flowed through the thin-walled silastic tubing loosely coiled in a glass jar that contained 20% O$_2$ and 5% CO$_2$ balanced with N$_2$), and a fraction collector (Waters Division of Millipore) (9). In this set of experiments, the Krebs buffer (pH 7.4) containing 1% BSA (fraction V, fatty acid free) was used.

Oxygen consumption measurements. Oxygen partial pressure was measured by the phosphorescence quenching method, using a new oxygen-sensitive phosphorescent porphyrin-dendrimer Oxyphor G3 (palladium-tetrabenzoxyphorphylin, encapsulated inside gen 2 polyarylglycine dendrimer) (22). The periphery of the dendrimer is modified with oligoethyleneglycol residues, which makes the probe water soluble and biologically inert. Oxyphor G3 does not require serum albumin in the perifusion medium, and thus there are no competitive interactions between dye and FA for binding to albumin.

The measured phosphorescence lifetime may be converted to oxygen pressure using the Stern-Volmer relationship $1/\tau_0 = 1/\tau_0 + k_Q \times [O_2]$, where $\tau_0$ is the phosphorescence lifetime in the absence of oxygen, $\tau$ is the lifetime at oxygen pressure $[O_2]$, and $k_Q$ is a constant describing the efficiency of quenching.

The inflow oxygen tension was measured in the absence of islets in the chamber before and after each experiment. Oxygen consumption by the islets was calculated from the difference in oxygen partial pressure between the influent and the effluent (oxygen extraction) and the rate at which medium flowed through the chamber.

Islet batch incubation studies. Islets were isolated as above, hand-picked, and cultured for 3 days. Batches of 50 islets were loaded into 12 × 75 mm disposable glass culture tubes and preincubated in 1 ml of oxygenated glucose-free Krebs-Ringer bicarbonate buffer (KRBB) at 37°C for 40 min, followed by a 45-min exposure to different stimuli. After incubations, islets and culture medium were transferred to 1.5-ml microcentrifuge tubes and spun at low speed. The supernatant was used for insulin measurements. Pelleted islets were washed two times with cold glucose-free Hanks’ buffer and then homogenized with 100 μl of 1% Triton in Tris-EDTA buffer. Protein concentration was measured in islet lysates using the Bradford method.

cAMP content determination. One-hundred cultured islets were exposed to different treatments for 30 min in the presence of 0.1 mmol/l isobutylmethylxanthine. After incubation, islets were quickly washed two times by cold glucose-free Hanks’ buffer. cAMP was measured in islet lysates by an enzyme-linked immunosorbent assay (GE Healthcare).

Ca$^{2+}$ measurement. Mouse islets were loaded with fura 2-AM during a 40-min pretreatment at 37°C in 2 ml of KRBB supplemented with 5 mmol/l fura 2-AM (Molecular Probes, Eugene, OR). The loaded islets were transferred to the perfusion chamber and placed on the homeothermic platform of an inverted Zeiss microscope. Islets were perfused with KRBB at 37°C at a flow rate of 2 ml/min, and various treatments were applied to the islets. The intracellular Ca$^{2+}$ was determined by the ratio of the excitation of fura at 334 and 380 nm. Emission was measured at 520 nm by an Zeiss charge-coupled device camera and calibrated using the Attofluor Ratio Vision Software.

Insulin measurements. Insulin in the effluent was measured by radioimmunoassay (15).

Statistical analysis. Data are presented as the mean ± SE of three to four experiments. In appropriate cases, significant

![Fig. 2. Effect of PA on glucose and Ach-stimulated oxygen consumption (A) and insulin release (B) in mouse islets. Oxygen consumption and insulin release were stimulated by 4 and then 8 mM G, followed by stimulation with Ach (0.5 μM). Each intervention lasted 40 min. FCCP (5 μM) was added for 30 min to uncouple the respiration and oxidative phosphorylation. NaN$_3$ (1 mM) was used at the end of each experiment to block respiration. In 1 set of experiments, PA (0.5 mM) was added at 0 mM G and was then present during the whole experiment (bold line (A) or (B)). There was a 4-min delay in the islet response to Ach in the presence of PA. Starting from 182 min of perfusion, the changes in insulin release were significant; $P < 0.05$. In these experiments, a new water-soluble generation of oxygen-sensitive dye Oxyphor G3 (Pd-tetrabenzoxyphorphylin, encapsulated inside a 2 polyarylglycine dendrimer) was used (see research design and methods) (22). Each curve represents the mean of 3–4 perfusions. Note that 8 mM G stimulates insulin release 3- to 4-fold but that the effect is dwarfed by the high efficacy of Ach. OCR, oxygen consumption rate.](http://ajpendo.physiology.org/)
differences between groups were determined by ANOVA with post hoc analysis using Dunnett’s multiple comparison test. \( P < 0.05 \) was considered significant. IGORPro data analysis software (Wavemetrics, Lake Oswego, OR) was used to measure the total insulin secretion by calculating the integral under the insulin curve (presented in relative units).

RESULTS

Effects of PA on glucose- and acetylcholine-stimulated insulin release in mouse islets. To examine the effects of PA on islet function, initial experiments utilized a fixed concentration of 0.5 mM of PA bound to 0.15 mM BSA. The PA/BSA molar ratio was 3.3, lower compared with most other studies (29, 33, 43) but moderately higher than that observed in humans (between 0.5 and 2, and rarely exceeding 4 in obese patients) (10, 14).

Fig. 3. The sequence of adding PA determines its effect on glucose-stimulated insulin release and oxygen consumption. A: PA was added 20 min before a glucose ramp from 0 to 25 mM (0.6 mM increment/min; \( n = 3 \)). B: a PA ramp from 0 to 0.5 mM (12.5 \( \mu \)M increment/min) was applied after islet preperfusion with 4 and then 8 mM G (\( n = 4 \)). PA starting from a concentration of 50 \( \mu \)M significantly potentiated insulin release (\( P < 0.05 \)). C: effect of PA on oxygen consumption. PA (0.5 mM) was added after islet preperfusion with 4 and then 8 mM G; D: insulin release for the experiments presented in C (\( n = 3 \)). Note that 120 islets and a 1.8 ml/min flow rate were used in experiments presented in A and B; 700 islets and an 80-ml flow rate were used in perfusions presented in C and D. The difference in number of islets and flow rate may result in a large difference in the absolute rate of insulin release in the 2 sets of experiments for reasons that remain to be determined. Also note that the \( O_2 \) pressure decreases by only 10–13%, such that anoxia cannot be a problem.

Table 1. Insulin secretion of mouse pancreatic islets cultured for 45 min

<table>
<thead>
<tr>
<th>Stimuli</th>
<th>Control</th>
<th>PA</th>
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<tbody>
<tr>
<td>0 mM G</td>
<td>2.26 ± 0.17</td>
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</tr>
<tr>
<td>8 mM G</td>
<td>3.49 ± 0.41</td>
<td>10.01 ± 1.44*</td>
</tr>
<tr>
<td>8 mM G + 10 ( \mu )M Go 6976</td>
<td>4.28 ± 1.44</td>
<td>2.46 ± 0.36*</td>
</tr>
<tr>
<td>8 mM G + 100 ( \mu )M 2-ADB</td>
<td>4.29 ± 2.55</td>
<td>3.06 ± 1.65</td>
</tr>
<tr>
<td>8 mM G + 1 ( \mu )M Ach</td>
<td>27.26 ± 0.80</td>
<td>13.08 ± 2.11*</td>
</tr>
<tr>
<td>8 mM G + 1 ( \mu )M Ach + 10 ( \mu )M Go 6976</td>
<td>14.15 ± 0.83†</td>
<td>13.87 ± 1.12</td>
</tr>
<tr>
<td>8 mM G + 1 ( \mu )M Ach + 100 ( \mu )M 2-ADB</td>
<td>7.81 ± 1.01†</td>
<td>9.72 ± 1.42</td>
</tr>
</tbody>
</table>

Values are means ± SE of 4 experiments. PA, palmitic acid; G, glucose; Go 6976, 12-(2-cyanoethyl)-6,7,12,13-tetrahydro-13-methyl-5-oxo-5H-indolo[2,3-a]pyrrolo[3,4-c]carbazolemodel; 2-ADB, 2-aminoethyl diphenylborinate; Ach, acetylcholine. Batches of 50 islets were preincubated in oxygenated glucose-free Krebs-Ringer bicarbonate buffer at 37°C for 40 min, followed by a 45-min exposure to different stimuli. Data are presented as ng insulin-\( \mu \)g protein\(^{-1}\)-45 min\(^{-1}\). *\( P < 0.05 \) compared with control of the same experimental design; †\( P < 0.05 \) compared with 8 mM glucose + Ach.
Islets were preperfused in the absence of glucose for 30 min, and then 4 and 8 mM glucose were added for 20 min. Afterward, acetylcholine or GLP-1 ramps were applied for 80 and 40 min, respectively, in the presence of 8 mM glucose. It should be noted that ramp protocols are unable to characterize time-dependent changes at a specific concentration of the secretagogue. Finally, 30 mM KCl was added after 30 min of a washout period with glucose-free medium. In one set of experiments, 0.5 mM of PA was included in the perfusate during the entire period. PA did not significantly change glucose-stimulated insulin release but markedly reduced the stimulation of insulin secretion by increasing concentrations of acetylcholine (Fig. 1A). Total insulin release during acetylcholine stimulation was reduced to one-half in the presence of 0.5 mM PA (917 ± 49.75 vs. 458.71 ± 37.35, P = 0.0003). The glucose-dependent GLP-1 stimulation of insulin release was not affected by PA (104.98 ± 2.65 vs. 119.15 ± 4.19, P = 0.06; Fig. 1B). In both sets of experiments, KCl produced a sharp biphasic increase in insulin release. It should be noted that insulin release during GLP-1 stimulation is only one-fifth compared with acetylcholine-induced secretion.

To evaluate the concentration dependence of PA’s effect on acetylcholine-stimulated insulin release, a PA ramp from 0 to 0.5 mM (12.5 µM increment/min with BSA fixed at 0.15 mM) was applied after islet stimulation with 0.5 µM of acetylcholine in the presence of 8 mM glucose (Fig. 1C). The molar PA/BSA ratio changed progressively from 0 to 3.3. In control experiments, islets were perfused with 0.5 µM of acetylcholine for the same duration as the PA ramp. The effect of acetylcholine on insulin release declined linearly with time (−0.053 ng insulin-min⁻¹·100 islets⁻¹), possibly reflecting desensitization of muscarinic receptors. In contrast, a marked exponential decrease in insulin release was seen when the PA ramp was applied. The slopes at the beginning and at the end of the ramp

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**Fig. 4.** Effect of 2-aminoethyl diphenylborinate (2-ADB), an inositol triphosphate (IP₃) receptor inhibitor, on Ach-stimulated insulin release in the presence and absence of PA. A: the sequence of adding PA does not affect its inhibitory action on Ach-stimulated insulin. PA was added after islet preperfusion with 4 mM G and then was present for the duration of the experiment. Potentiation of insulin release by Ach was reduced significantly in the presence of PA (106–165 min of perfusion, P < 0.05). B: a magnified view of 1 section (55–85 min) of the perfusion experiments is presented to clearly show the potentiating effect of PA on glucose-stimulated insulin release. C: effect of 2-ADB, an IP₃ receptor inhibitor, on Ach-stimulated insulin secretion. Each curve represents the mean ± SE of 3 perfusions.
were $-0.18$ and $0.10$ ng insulin·min$^{-1}$·100 islets$^{-1}$. A significant difference between the slopes was already evident at 50 
µM PA ($P = 0.043$). In contrast to the acetylcholine effect, glucose-dependent GLP-1 stimulation of insulin release was potentiated by PA (Fig. 1D).

**Oxygen consumption.** PA caused a small, gradual increase ($\sim 15\%$) in basal oxygen consumption rate (OCR) of perfused mouse islets (Fig. 2A) and did not change basal insulin release (Fig. 2B) in the absence of glucose. Glucose-stimulated insulin release occurred at 8 mM, and this effect was not potentiated by PA. The increment of the OCR due to low glucose (4 mM) was reduced by two-thirds in the presence of PA ($P \leq 0.05$, time interval from 94 to 126 min). Glucose at 8 mM produced the same stimulation of the OCR in both cases; however, the maximal OCR was significantly lower in the presence of PA ($P \leq 0.05$, time interval from 128 to 160 min). In contrast to dramatic stimulation of insulin release, acetylcholine affected oxygen consumption only a little, suggesting that the energy requirement for exocytosis is very small. As seen in the previous sets of experiments, PA inhibited glucose-dependent acetylcholine stimulation of insulin release $>50\%$ (45.67 ± 6.38 vs. 18.43 ± 3.65, $P = 0.009$; Fig. 2B). Uncoupling mitochondrial respiration and oxidative phosphorylation by FCCP (5 µM) blocked insulin release. The effect of FCCP on OCR was biphasic: a small stimulation followed by inhibition. It should be noted that FCCP may change the intracellular pH because it is a protonophore. We have shown previously that stimulation of respiration by FCCP in mouse islets occurs only when the perifusion medium is supplemented by a mixture of amino acids (9) or glutamine (23). It should be noted that, in this set of experiments, 600–700 islets were perfused with a flow rate of 80 µl/min. This resulted in a significantly lower absolute rate of insulin release compared with that in experiments where 130 islets were perfused with a flow rate of 2 
ml/min. Higher number of islets and lower flow rate may lead to higher insulin and glucagon concentration within the islet extracellular space and the closely surrounding medium.

**The sequence of adding PA.** The absence in our experiments of PA potentiation of insulin release by 8 mM glucose is in disagreement with previous studies (6, 31). To clarify these differences, three sets of experiments were performed. In one, PA was added prior to a glucose ramp from 0 to 25 mM (0.6 
mM increment/min; Fig. 3A); in the second, a PA ramp from 0 to 0.5 mM (12.5 µM increment/min) was applied after preperfusion with 8 mM glucose (Fig. 3B); in the third, batch incubation was used to evaluate the effect of PA on insulin secretion (Table 1). In the first scenario, PA did not significantly affect the glucose-stimulated insulin release, although a trend toward decreasing glucose-stimulated insulin release was seen at high concentrations of glucose ($>8$ mM; Fig. 3A). In contrast, a marked stimulation of insulin release by a PA ramp was evident following islet preperfusion with 8 mM glucose (Fig. 3B), with the effect of PA on insulin release already significant at 50 µM PA ($0.97 \pm 0.19$ vs. $2.65 \pm 0.54$, $P = 0.026$).

When, instead of perfusion, batch incubation was used, then PA significantly potentiated insulin release at 8 mM glucose in the course of 45-min incubation (see Table 1).

It was of interest whether such stimulation of insulin secretion by PA in the presence of 8 mM glucose would lead to an increased OCR. Therefore, a similar design was used in OCR experiments. In contrast to markedly elevated insulin release (Fig. 3D), PA did not significantly alter oxygen consumption (Fig. 3C).

We have also tested whether the sequence of adding PA (in the absence or presence of glucose) would change the acetylcholine effect on insulin release. Therefore, in one set of experiments, 0.5 mM of PA was added after islets were preperfused with 4 mM glucose. As in previous experiments, PA did not increase insulin release at a basal glucose concentration (Fig. 4A) but potentiated the second phase of insulin release during stimulation with 8 mM glucose (Fig. 4B). However, the sequence of adding the PA (at 0 or 4 mM glucose) did not affect the inhibitory action of PA on acetylcholine-stimulated insulin release [1,028 ± 64.07 vs. 636.84 ± 74.74, $P = 0.017$; Fig. 4A (also see Fig. 1A)].

**The role of inositol triphosphate receptors, PKC, and cAMP.** It is known that acetylcholine activates PLC, resulting in the generation of diacylglycerol, a potent PKC activator, and
inositol triphosphate (IP$_3$), which causes a rapid mobilization of Ca$^{2+}$ from the endoplasmic reticulum (ER) (13). Both PKC and Ca$^{2+}$ activate adenylyl cyclase (types 2, 3, 5, and 7) and increase cAMP (39). Therefore, the next set of experiments was designed to evaluate the involvement of PKC, cAMP, and IP$_3$-induced Ca$^{2+}$ mobilization in the inhibitory effect of PA on acetylcholine-stimulated insulin secretion.

Gradual increases in concentration of the phorbol 12,13-dibutyrate, an activator of PKC, from 0 to 1 $\mu$M in the presence of 8 mM glucose greatly augmented insulin release (Fig. 5A). The presence of PA in the perfusate did not reduce insulin release; on the contrary, a trend for potentiating insulin secretion at the higher concentrations was evident (641.49 ± 129.16 vs. 775.12 ± 136.38, $P = 0.5$). In the next set of experiments, an inhibitor of PKC, Go 6976 {12-(2-cyanoethyl)-6,7,12,13-tetrahydro-13-methyl-5-oxo-5H-indolo[2,3-a]pyrrolo[3,4-c]carbazolemodel; specific for $\alpha$ (IC$_{50}$ = 2.3 nM) and $\beta$1 (IC$_{50}$ = 6.2 nM) isozymes}, was used, and insulin secretion was evaluated in batch incubation (Table 1). The PKC inhibitor reduced to one-half the acetylcholine effect at 8 mM glucose, supporting the general view that PKC is the major intracellular target for acetylcholine action. At this condition, PA did not significantly affect the acetylcholine effect on insulin secretion. Both sets of data suggest that the inhibitory effect of PA on acetylcholine-stimulated insulin release is upstream of PKC.

In another set of perifusion experiments, forskolin (ramp from 0 to 10 $\mu$M) was used to gradually increase the level of intracellular cAMP. PA did not significantly affect the forskolin-stimulated insulin secretion (432.26 ± 121.19 vs. 725.79 ± 106.1, $P = 0.143$), although the insulin secretion profile was shifted to the left (Fig. 5B).

In the next set of experiments, we tested whether intracellular cAMP levels are modulated by acute exposure to PA. Both high glucose and acetylcholine elevated the intracellular cAMP concentration and enhanced insulin release in batch incubation experiments (Fig. 6, A and B). PA prevented the rise of cAMP during acetylcholine stimulation in the presence of 8 mM glucose and significantly reduced the stimulation of insulin release by acetylcholine. In contrast, the effects of GLP-1 on both cAMP concentrations and insulin release were not influenced by PA administration (Fig. 6, C and D). Thus, the acetylcholine-induced changes in intracellular cAMP level are altered by PA. Since the changes in cAMP are downstream of intracellular Ca$^{2+}$, we tested whether inhibition of the IP$_3$-mediated Ca$^{2+}$ mobilization from the ER by 2-aminoethyl diphenylborinate (2-ADB) would change the PA effect on acetylcholine-induced insulin secretion. 2-ADB greatly reduced acetylcholine’s effect on insulin secretion and, surprisingly, partially reversed the effect of PA on acetylcholine stimulation of hormone release from inhibition to stimulation (Fig. 4C).

Changes in intracellular Ca$^{2+}$ concentration in response to PA. Fura-2 fluorescence techniques were used to evaluate whether intracellular changes in Ca$^{2+}$ contribute to altered insulin release in the presence of PA. Eight millimolar glucose caused biphasic changes in Ca$^{2+}$ (a spike of Ca$^{2+}$ followed by higher basal levels; Fig. 7A). PA, when applied after preperfusion with 8 mM glucose, significantly increased intracellular Ca$^{2+}$ concentrations (Fig. 7A). Acetylcholine also increased...
Ca²⁺ and induced serial Ca²⁺ oscillations in the majority of control islets (Fig. 7B). PA (0.5 mM) did not modify the effect of 8 mM glucose; however, it reduced the acetylcholine-stimulated rise in Ca²⁺ (31,528 vs. 18,847, *P* = 0.042; Fig. 7B).

In the next set of experiments, thapsigargin was used to indirectly measure the calcium storage capacity of the ER when acutely influenced by PA (Fig. 7C). It was found that 35–40 min of islet incubation with PA in the presence of 8 mM glucose significantly reduced the calcium storage capacity of the ER. Blocking IP₃ receptors by 2-ADB decreased thapsigargin’s effect on Ca²⁺ release and abolished the Ca²⁺-depletive effect of PA on ER but decreased the magnitude of the thapsigargin response (Fig. 7D). Based on the mechanism well described in the literature that acetylcholine acts through IP₃-mediated Ca²⁺ release from ER, it is proposed that the emptying of ER calcium by PA interferes with acetylcholine effects on islet function.

**DISCUSSION**

Nutrient-induced insulin secretion is potentiated by many hormones and neurotransmitters. Acetylcholine, the parasympathetic neurotransmitter, is released by intrapancreatic vagal nerve endings during the preabsorptive and absorptive phases of feeding, and it acts mainly by binding to β-cell muscarinic M₃ receptors. It enhances glucose-stimulated insulin secretion (13) by the following mechanism: activation of PLC, an increase in IP₃ that causes mobilization of Ca²⁺ from intracellular stores, and formation of diacylglycerol, which activates PKC, resulting in increased efficiency of Ca²⁺ on exocytosis during glucose-stimulated insulin release (Fig. 8).

Despite a crucial role of acetylcholine in the regulation of insulin release, information on the influence of FFA on this regulation is lacking. Results of the present study demonstrate that PA at physiological levels acutely inhibits acetylcholine-induced stimulation of insulin secretion. This inhibition does not depend on the sequence of adding PA (at 0 or 4 mM glucose) and is also evident when a PA ramp is applied after enhanced insulin secretion is initiated by acetylcholine in the presence of moderately high glucose. In the latter case, the suppression of acetylcholine-stimulated insulin release is evident at very low-concentration PA (50 μM albumin bound). The inhibitory effect of PA is specific for acetylcholine and...
applied at a fixed albumin concentration of 0.15 mM such that two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. In the present study, two different experimental designs were applied. 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oleate and, to a greater extent, palmitate depleted ER Ca\(^{2+}\) by 25 and 40%, respectively, as tested by thapsigargin-induced Ca\(^{2+}\) release (7). The present study indicates that such an effect of PA on ER Ca\(^{2+}\) release is evident even after 35–40 min of incubation with PA. Blocking IP\(_3\) receptors abolished the effect of PA on the ER and therefore also on acetylcholine stimulation of insulin secretion. It has also been shown that the GLP-1 agonists exendin-4 and forskolin protected primary β-cells and INS-1E cells against oleate- and palmitate-induced (14–24 h) ER stress and apoptosis (8). These literature data may explain the difference in the effects of PA on acetylcholine and GLP-1 potentiation of insulin secretion. Thus, the data suggest that acute emptying of ER Ca\(^{2+}\) stores by PA interferes with acetylcholine’s action on islet function.

Effects of PA on oxygen consumption. Current knowledge about effects of fatty acid on oxygen consumption is limited. Conget et al. (5) have shown that PA failed to affect O\(_2\) uptake at 6 mM glucose during 10- to 15-min exposure to this fatty acid. Palmitate and oleate failed to affect the oxidation of H\([U-14C]\)glucose or its conversion to \(^{14}\)C-labeled amino acids (5). Our data suggest that PA increases oxygen consumption only slightly at 0 mM glucose, but it decreased stimulation of oxygen consumption by 4 mM glucose, resulting in a lower total response to glucose (0 → 4 → 8 mM). It is known that cytosolic Ca\(^{2+}\) levels affect intramitochondrial free Ca\(^{2+}\) concentration and activity of several matrix dehydrogenases, resulting in a change of Krebs cycle turnover (24). On the basis of these data, we speculate that the emptying of ER calcium by FFA may contribute to the reduced total oxygen consumption response to glucose.

In conclusion, PA interferes acutely with acetylcholine stimulation of pancreatic β-cell metabolism and secretion, as evidenced by attenuated elevations of insulin release, intracellular cAMP, and Ca\(^{2+}\). In addition, PA reduces the effect of low glucose on oxygen consumption; however, it greatly potentiates insulin secretion in the presence of high glucose dose dependently. In contrast to acetylcholine, the GLP-1 effects on these parameters remain unchanged or potentiated. The results of the present study suggest that acute emptying of ER calcium by PA interferes with cholinergic stimulation of insulin secretion.

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

REFERENCES


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