Deficiency of electron transport chain in human skeletal muscle mitochondria in type 2 diabetes mellitus and obesity


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Ritov VB, Menshikova EV, Azuma K, Wood R, Toledo FG, Goodpaster BH, Ruderman NB, Kelley DE. Deficiency of electron transport chain in human skeletal muscle mitochondria in type 2 diabetes mellitus and obesity. Am J Physiol Endocrinol Metab 298: E49–E58, 2010. First published November 3, 2009; doi:10.1152/ajpendo.00317.2009.— Insulin resistance in skeletal muscle in obesity and T2DM is associated with reduced muscle oxidative capacity, reduced expression in nuclear genes responsible for oxidative metabolism, and reduced activity of mitochondrial electron transport chain. The presented study was undertaken to analyze mitochondrial content and mitochondrial enzyme profile in skeletal muscle of sedentary lean individuals and to compare that with our previous data on obese or obese T2DM group. Frozen skeletal muscle biopsies obtained from lean volunteers were used to estimate cardiolipin content, mtDNA (markers of mitochondrial mass), NADH oxidase activity of mitochondrial electron transport chain (ETC), and activity of citrate synthase and β-hydroxyacyl-CoA dehydrogenase (β-HAD), key enzymes of TCA cycle and β-oxidation pathway, respectively. Frozen biopsies collected from obese or T2DM individuals in our previous studies were used to estimate activity of β-HAD. The obtained data were complemented by data from our previous studies and statistically analyzed to compare mitochondrial content and mitochondrial enzyme profile in lean, obese, or T2DM cohort. The total activity of NADH oxidase was reduced significantly in obese or T2DM subjects. The cardiolipin content for lean or obese group was similar, and although for T2DM group cardiolipin showed a tendency to decline, it was statistically insignificant. The total activity of citrate synthase for lean and T2DM group was similar; however, it was increased significantly in the obese group. Activity of β-HAD and mtDNA content was similar for all three groups. We conclude that the total activity of NADH oxidase in biopsy for lean group is significantly higher than corresponding activity for obese or T2DM cohort. The specific activity of NADH oxidase (per mg cardiolipin) and NADH oxidase/citrate synthase and NADH oxidase/β-HAD ratios are reduced two- to threefold in both T2DM and obesity.

insulin resistance; β-oxidation; cardiolipin; reduced nicotinamide adenine dinucleotide/nicotinamide adenine dinucleotide ratio; trichloroacetic acid cycle

TYPE 2 DIABETES MELLITUS (T2DM) is characterized by insulin resistance in skeletal muscle that can be a result of combining a genetic predisposition with obesity and sedentary lifestyle (12, 34, 44). Insulin resistance in skeletal muscle in obesity or T2DM is associated with reduced muscle oxidative capacity (53, 60) and with reduced expression in a cluster of nuclear genes responsible for oxidative metabolism [peroxisome proliferator-activated receptor-γ coactivator-1α (PGC-1α) and genes activated by PGC-1α] (17, 30, 33, 38). In our previous study, we found a significant reduction in the total activity of mitochondrial electron transport in the skeletal muscle in obese and T2DM individuals compared with lean and healthy controls (24). The content of mitochondria in human skeletal muscle depends strictly on the level of physical activity; it increases with exercise (14, 19–21) and decreases during the detraining period (8). The deficiency in the mitochondrial electron transport in skeletal muscle in T2DM and obesity could be a result of deficiency in mitochondria mass, deficiency in mitochondrial function, or both. The current study was undertaken to address these questions. Vastus lateralis needle biopsies were obtained from sedentary lean individuals before and after completion of a program of moderate-intensity aerobic exercise that could benefit study participants. Frozen tissue biopsies were used to estimate markers of mitochondrial mass [cardiolipin and mitochondrial DNA (mtDNA)], activity of mitochondrial electron transport chain (ETC), and activity of citrate synthase and β-hydroxyacyl-CoA dehydrogenase (β-HAD), which are key enzymes of the TCA cycle and β-oxidation pathway, respectively. Frozen biopsies collected from obese or T2DM individuals in our previous studies (58, 59) were used to estimate activity of β-HAD. The obtained data were complemented by data from our previous studies and statistically analyzed to compare mitochondrial content and mitochondrial enzyme profile in the lean, obese, or T2DM cohort.

Several advanced techniques previously developed in our laboratory for analysis of mitochondrial function and content in skeletal muscle were applied in this study. We developed a unique, nondestructive procedure for analysis of activity of mitochondrial ETC in human skeletal muscle. This procedure is based on the use of the channel-forming peptide alamethicin that provides unrestricted access of NADH inside of mitochondrial ETC in middle-aged sedentary lean subjects. The combined data (from the current and our previous studies) show for the first time that the content of mitochondrial mass markers (cardiolipin, mtDNA) and activity of citrate synthase or β-HAD in middle-aged sedentary lean subjects is
similar to obese or T2DM individuals. However, the average activity of ETC for lean group is significantly higher than corresponding activity for the obese or T2DM cohort. As a result, the NADH oxidase/cardiolipin (specific activity of NADH oxidase), NADH oxidase/citrate synthase, and NADH oxidase/β-HAD ratios are reduced significantly (2- to 3-fold) in both T2DM and obesity compared with sedentary lean subjects. We hypothesize that this specific mitochondrial profile (normal or excessive production of reducing equivalents by TCA cycle and β-oxidation and the deficiency in oxidative capacity of ETC) plus sedentary lifestyle can affect metabolite profile in skeletal muscle in T2DM or obesity by shifting it to increased NADH/NAD⁺ ratio and to accumulation of toxic byproducts of incomplete oxidation of glucose and lipids, and that, in turn, as was recently suggested, can lead to the development of skeletal muscle insulin resistance (37).

MATERIALS AND METHODS

Research volunteers. The protocol was approved by the University of Pittsburgh Institutional Review Board. All participants had a screening medical history, physical examination, and screening laboratory tests. The inclusion criteria for all groups were described elsewhere (58, 59). The obese nondiabetic and obese type 2 diabetic groups were matched for age and body mass index. Clinical characteristics of the research volunteers are presented in Table 1.

Intervention. The intervention lasted 16–20 wk after initial baseline assessments and was monitored by dietitians and exercise physiologists. Participants were asked to participate in four to six exercise sessions weekly, with at least one supervised session weekly. The physical activity intervention was moderate-intensity exercise at 60–70% of maximal heart rate for 30 min for the first 4 wk, increased to 40 min for the next 4 wk, and increasing intensity but not duration for the final 8 wk. Mostly, the exercise included using a stationary cycle, treadmill, or walking (58, 59). In the present study, for analysis of β-HAD, we also used along with frozen biopsies from lean individuals the frozen biopsies obtained from obese or T2DM participants in our previous studies (58, 59). In our previous study, the nutritional intervention was separate or concomitant to exercise and aimed at a target weight loss of ≥5% of baseline weight via a reduction in calorie intake of 500–1,000 kcal/day (58).

Metabolic assessments. Euglycemic clamps were used to measure insulin sensitivity (59). Maximal aerobic capacity (\( \text{V} \text{O}_{2 \text{max}} \)) was measured on a treadmill with a modified Bruce protocol (58).

Analysis of mitochondrial content and function. Muscle biopsy samples of vastus lateralis muscle (15–25 mg wet wt) obtained by Bergstrom needle were analyzed on mtDNA (31) and cardiolipin (47) content, citrate synthase (48), β-HAD, and ETC NADH oxidase activities. Frozen samples (particulate and soluble homogenate fractions) prepared from biopsies collected from obese or T2DM individuals in our previous studies and stored in 25% glycerol at –80°C were used for analyses of activity of β-HAD. Activity of β-HAD was simultaneously estimated in the present study for the lean, obese, or T2DM group. The obtained data were complemented by data from our previous studies and statistically analyzed to compare mitochondrial content and mitochondrial enzyme profile in the lean, obese, or T2DM cohort. Activity of β-HAD was measured at 30°C in the reverse reaction by standard method adapted for higher sensitivity to our HPLC technique (48). Activity of rotenone-sensitive NADH:O₂ oxidoreductase in total particulate fraction prepared from muscle homogenate was measured in the presence of alamethicin, as described previously (48). Creatine kinase activity was used as a marker of muscle fiber content in biopsy (48).

Reagents and equipment. Tetraoleoyl cardiolipin (internal standard) was purchased from Avanti Polar Lipids (Alabaster, AL). HPLC grade chloroform, stabilized by 0.7% ethanol, was obtained from Fisher Scientific (Pittsburgh, PA). Other HPLC grade solvents and reagents were purchased from Sigma Chemical (St. Louis, MO). 1-Pyrenyl-1-diazomethane was obtained from Molecular Probes (Eugene, OR). A Shimadzu high-performance liquid chromatograph (model LC-10AT vp) equipped with an autosampler (model SIL-10AD), a tray cooler, and a Shimadzu fluorescence detector (model RF-10AXl) was used for these studies. The analog signal of the detector was processed and stored in digital form with Shimadzu Class-VP software (Shimadzu Scientific Instruments, Columbia, MD).

Statistical analysis. Data are presented as means ± SE unless otherwise indicated. To address the effects of intervention on insulin sensitivity and skeletal muscle parameters, a paired t-test was applied to all data. ANOVA was used to compare groups. Statistical significance was assumed a priori at \( P < 0.05 \).

RESULTS

Mitochondrial content and mitochondrial enzyme activities in the basal conditions. The biopsies were taken from sedentary nondiabetic lean subjects (current study) or from sedentary type 2 diabetic or nondiabetic obese subjects (58, 59). The clinical characteristics of the research volunteers are presented in Table 1. The content of mitochondria in the biopsy was assessed from analysis of mtDNA (31) and cardiolipin (47). Table 2 presents data from the current study on the lean subjects and data from previous studies on obese and T2DM subjects (58, 59). The statistical analysis of these data shows that there is no significant difference in the cardiolipin or mtDNA content between obese and healthy sedentary lean groups in the basal state before the intervention (Table 2). The T2DM group shows a tendency to reduced level of cardiolipin; however, it is not statistically significant. In conclusion, the

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Table 1. Clinical characteristics of the research volunteers

<table>
<thead>
<tr>
<th></th>
<th>Lean (n = 9)</th>
<th>Obese (n = 9)</th>
<th>T2DM (n = 10)</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>Pre-Int</td>
<td>Post-Int</td>
<td>Pre-Int</td>
</tr>
<tr>
<td>Age, yr</td>
<td>47.1 ± 2.2</td>
<td>42.4 ± 2.7</td>
<td>44 ± 3</td>
</tr>
<tr>
<td>BMI, kg/m²</td>
<td>24.0 ± 0.7</td>
<td>23.6 ± 0.7</td>
<td>31.9 ± 1.2</td>
</tr>
<tr>
<td>Weight, kg</td>
<td>65.4 ± 2.9</td>
<td>64.5 ± 2.8</td>
<td>86.3 ± 6.6*</td>
</tr>
<tr>
<td>Fat mass, kg</td>
<td>20.3 ± 1.3</td>
<td>18.7 ± 0.7</td>
<td>33.1 ± 1.9*</td>
</tr>
<tr>
<td>Fat-free mass, kg</td>
<td>44.3 ± 3.2</td>
<td>44.4 ± 3.3</td>
<td>51.3 ± 3.5</td>
</tr>
<tr>
<td>( \text{V} \text{O}_{2 \text{max}} ), ml·kg⁻¹·FMM⁻¹·min⁻¹</td>
<td>52.7 ± 1.7</td>
<td>53.2 ± 2.1</td>
<td>52.4 ± 2.2* (P &lt; 0.005)</td>
</tr>
<tr>
<td>( \text{G} \text{INSF} ), mg·kg⁻¹·FMM⁻¹·min⁻¹</td>
<td>12.2 ± 2.8</td>
<td>13.8 ± 1.1</td>
<td>8.7 ± 1.1* (P &lt; 0.001)</td>
</tr>
</tbody>
</table>

Values are means ± SE. T2DM, type 2 diabetes mellitus; Pre-Int, before intervention; Post-Int, after intervention; BMI, body mass index; \( \text{V} \text{O}_{2 \text{max}} \), maximal aerobic capacity; FFM, fat-free mass; \( \text{G} \text{INSF} \), glucose infusion rate during euglycemic clamp. *Statistically significant.
The lean participants of this study completed a 16- to 20-wk exercise program, as described in MATERIALS AND METHODS. This exercise intervention significantly increased cardiolipin and mitochondrial DNA content. Nevertheless, after completion of the exercise program, the changes in VO2 max, which is an index of physical fitness, were nonsignificant for lean, previously sedentary volunteers.

The lean subjects that participated in the exercise program showed significant increase in total activity of ETC (NADH oxidase), citrate synthase, and β-HAD calculated per 1,000 U of tissue creatine kinase (Figs. 1 and 2). However, the specific activity of ETC, citrate synthase, or β-HAD calculated relative to cardiolipin as a replacement index of mitochondrial mass remains the same after completion of the exercise program by lean participants (NADH oxidase; Fig. 2A). Accordingly, the completion of the exercise program by lean participants had no effect on the NADH oxidase/citrate synthase or NADH oxidase/β-HAD ratios that were still significantly (2- to 3-fold) higher than corresponding ratios for both T2DM and obese participants (Fig. 2, B and D).

In our previous study, T2DM or obese volunteers completed two programs for lifestyle modification; one was based on a caloric restriction, and another one, in addition to caloric restriction, included the exercise program that was similar to the exercise program that was applied to lean subjects in the current study. Both interventions were aimed to reach a weight loss of ≮5–10% of baseline weight (58, 59). It had been found that only the program that included the exercise induced the increase in cardiolipin content, citrate synthase, and mitochondrial NADH oxidase activities in the biopsies obtained from obese or T2DM individuals (58, 59).

In the current study, we used particulate and soluble fractions prepared from the biopsies obtained from the obese or T2DM individuals who participated in the study that included combined intervention (diet plus exercise) to additionally measure the activity of β-HAD. As can be seen in Fig. 2C, according to the previous data, the intervention program that included exercise significantly increased activity of β-HAD in vastus lateralis muscle of obese or T2DM individuals. We used these data and the previously published data on NADH oxidase, cardiolipin, and citrate synthase to calculate the ratios between NADH oxidase/citrate synthase or NADH oxidase/β-HAD and to compare it with corresponding ratios for lean volunteers. As can be seen in Fig. 2, the caloric restriction plus exercise intervention does not improve the balance between activity of NADH oxidase and citrate synthase or β-HAD.

## Table 2. Markers of mitochondrial mass and total activity electron transport chain in vastus lateralis biopsy

<table>
<thead>
<tr>
<th>Basal State</th>
<th>Lean (n = 9)</th>
<th>Obese (n = 15)</th>
<th>T2DM (n = 10)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total tissue CK, U/g wet wt</td>
<td>5,655±388</td>
<td>5,808±315</td>
<td>6,198±316</td>
</tr>
<tr>
<td>Cardiolipin, µg/mU CK</td>
<td>78.2±9.5</td>
<td>76.0±6.4</td>
<td>58.6±5.5 (P = 0.079)</td>
</tr>
<tr>
<td>CS, U/mU CK</td>
<td>2.70±0.28</td>
<td>3.7±0.5* (P &lt; 0.02)</td>
<td>3.28±0.49</td>
</tr>
<tr>
<td>mtDNA (Rc)</td>
<td>2.417±218</td>
<td>2.271±286</td>
<td>2.114±156</td>
</tr>
<tr>
<td>Rotenone-sensitive NADH oxidase, U/mU CK</td>
<td>0.44±0.12</td>
<td>0.180±0.02* (P &lt; 0.04)</td>
<td>0.160±0.03* (P &lt; 0.03)</td>
</tr>
</tbody>
</table>

Values are means ± SE. T2DM, type 2 diabetes mellitus; CK, creatine kinase; mtDNA, mitochondrial DNA; Rc, relative copy number of mtDNA per diploid nuclear genome; CS, citrate synthase. *P values are for T2DM or obese vs. healthy lean sedentary individuals. *Statistically significant.
DISCUSSION

The primary goal of the presented study was to compare mitochondrial content and activity of selected mitochondrial enzymes in skeletal muscle of biopsies obtained from lean, obese, or healthy lean sedentary individuals. Frozen tissue samples obtained by needle biopsy from research volunteers were homogenized, and the homogenate was separated by high-speed centrifuging to collect the total particulate and soluble fractions. Total particulate fraction that contains 95% of tissue mitochondria was used for analysis of cardiolipin and activity of mitochondrial ETC (rotenone-sensitive NADH:O2 oxidoreductase). Both particulate and soluble fractions were used for analysis of citrate synthase and β-HAD because of partial release of these soluble mitochondrial matrix enzymes from mitochondria due to tissue freezing.

Mitochondrial content in human vastus lateralis muscle in obesity and T2DM. There is little choice for nonenzymatic markers to quantitate mitochondrial content in tissue homogenates or total particulate fractions that are complex mixtures of proteins and subcellular organelles. Mitochondrial DNA or cardiolipin can be considered as suitable markers. Previously, we found decreased mtDNA in obesity and diabetes (46). Mitochondrial DNA is increased significantly in human skeletal muscle during adaptation to high-intensity physical activity (41). Also, in nonobese elderly individuals, a moderate-intensity physical activity intervention induced a significant increase in muscle mtDNA content along with the increase in activity of citrate synthase and ETC (27). As can be seen in Table 2, the content of mtDNA increases significantly in vastus lateralis of lean sedentary study participants after completion

![Fig. 1](http://ajpendo.physiology.org/). The markers of mitochondrial mass and enzyme activity in skeletal muscle biopsy from type 2 diabetes mellitus (T2DM), obese, or healthy lean sedentary individuals. Data represent means ± SE. The total content of cardiolipin (A), citrate synthase enzymatic activity (B), and rotenone-sensitive NADH oxidase enzymatic activity (C) in biopsy were normalized to the creatine kinase activity. D: mitochondrial DNA (mtDNA) presented as a relative copy number of mtDNA/diploid nuclear genome. Biopsies were analyzed before (Pre-Int) and after intervention (Post-Int). P values are for mitochondrial parameters Pre-Int vs. after Post-Int. CK, creatine kinase.
of an exercise program. However, it should be taken into consideration that mitochondrial nucleoid contains a variable number of DNA copies (2–10 copies) that makes mtDNA content only a semiquantitative indicator for mitochondrial mass. In fact, Menshikova et al. (29) observed that the adaptation to moderate-intensity exercise in obesity occurs without an increase in muscle mtDNA content. Additionally, the mtDNA content in skeletal muscle does not always reflect the state of mitochondrial biogenesis. Combined weight loss plus exercise intervention in obese or T2DM cohort significantly increases content of cardiolipin, activity of NADH oxidase, citrate synthase (58, 59), or β-HAD (Fig. 1 and 2). Taken together, it is a clear indication of the activation of mitochondrial biogenesis induced by intervention. However, the same intervention is unable to induce the increase in mtDNA in these obese or T2DM study participants (Fig. 2) (58, 59). A more reliable marker for mitochondrial mass is mitochondrial phospholipid cardiolipin. Cardiolipin is a phospholipid that is specific to the inner mitochondrial membrane (18). This phospholipid is a basic structural component of mitochondria that accounts for ~15% of total lipids in inner mitochondrial membrane (10). Cardiolipin is a sensitive marker of mitochondrial content and mitochondrial biogenesis. Cardiolipin content in vastus lateralis obtained from the highly trained lean volunteers is ~1.0 mg/g wet wt, which is more than twofold higher than the cardiolipin content in biopsies obtained from sedentary volunteers in the present study (28). Cardiolipin content in vastus lateralis biopsy is increased after any intervention that includes exercise (28, 29, 58). Conversely, cardiolipin decreases along enzymes after muscle inactivity induced by denervation in animals (62). It is increased independently on concomitant caloric restriction, and this increase is proportional to the increase in mitochondrial activity of ubiquinol oxidase (respiratory complexes III plus IV) (28). Inner membrane in skeletal muscle mitochondria, which forms densely packed mitochondrial cristae, contributes >80% of the total mitochondrial mass.
The predominant molecular form is tetralinoleoyl cardiolipin, which accounted for ~80% of cardiolipin in human skeletal muscle (47, 52). In the basal conditions, there is no significant deficiency in the mitochondrial mass in insulin-resistant obese subjects assessed by analysis of cardiolipin (Table 1 and Fig. 1). This is in accordance with previous findings that showed the preservation of a high level of citrate synthase in skeletal muscle in obesity but not in T2DM (24). The T2DM group showed a tendency to reduce the level of cardiolipin; however, this trend does not reach the statistical significance for the present number of study participants, although in our first study on mitochondria in obesity and T2DM, the trend in the slight decrease in mitochondria mass for participants from the T2DM group (estimated from citrate synthase activity) had reached statistical significance (24). The absence of effects of obesity/insulin resistance on mitochondrial content in biopsy suggests that the reduction in mitochondrial mass in skeletal muscle and changes in mitochondrial morphology that linked to T2DM (32, 42) can be secondary to this condition.

Mitochondrial enzyme profile in obesity and T2DM. Measurements of activity of mitochondrial enzymes require eliminating a permeability barrier for mitochondrial substrates created by inner mitochondrial membrane. This especially concerns activity of NADH oxidase because of the lack of NADH transporter in mitochondria to cross inner membrane. The standard approach to eliminating the permeability barrier in mitochondria is based on exposing mitochondria to detergents. It effectively allows measurements of activity of separate respiratory complexes and matrix enzymes (5). However, it prevents analysis of activity of whole ETC because detergents disturb the interaction between respiratory complexes in inner mitochondrial membrane. Previously, we developed a detergent-free assay for estimation of activity of whole mitochondrial ETC directly from analysis of oxidation of NADH (24). The assay includes a channel-forming peptide alamethicin that provides unrestricted access of NADH inside of mitochondria without any effects on the integrity of mitochondrial ETC. This assay provides information on the real oxidative capacity of mitochondrial ETC contrary to respiration technique (6) that measures maximal oxygen consumption in the state of mitochondrial ATP synthesis (state 3) or in the presence of uncouplers and TCA cycle substrates. The respiration or ATP synthesis in mitochondria is controlled mostly by specific transporters that deliver ADP, phosphate, and other substrates into mitochondria (57, 63, 64). The flux control of substrates into mitochondria is based on exposing mitochondria to detergents. It effectively allows measurements of activity of separate respiratory complexes and matrix enzymes (5). However, it prevents analysis of activity of whole ETC because detergents disturb the interaction between respiratory complexes in inner mitochondrial membrane. Previously, we developed a detergent-free assay for estimation of activity of whole mitochondrial ETC directly from analysis of oxidation of NADH (24). The assay includes a channel-forming peptide alamethicin that provides unrestricted access of NADH inside of mitochondria without any effects on the integrity of mitochondrial ETC. This assay provides information on the real oxidative capacity of mitochondrial ETC contrary to respiration technique (6) that measures maximal oxygen consumption in the state of mitochondrial ATP synthesis (state 3) or in the presence of uncouplers and TCA cycle substrates. The respiration or ATP synthesis in mitochondria is controlled mostly by specific transporters that deliver ADP, phosphate, and other substrates into mitochondria (57, 63, 64). The flux control of substrates into mitochondria is controlled mostly by specific transporters that deliver ADP, phosphate, and other substrates into mitochondria.

The comparison of the data from the present study on the lean sedentary volunteers and data from our two previous studies on obese and T2DM participants (58, 59) shows that the distribution of molecular forms of cardiolipin for lean sedentary, obese, or T2DM was the same (data not shown). The predominant molecular form is tetralinoleoyl cardiolipin, which accounted for ~80% of cardiolipin in human skeletal muscle (47, 52). In the basal conditions, there is no significant
The mitochondrial enzyme profile specific for obesity or T2DM can be reproduced in an animal model. Our recent data show that high-fat diet induces similar changes in rat liver mitochondria (increase in citrate synthase activity and β-HAD and decrease in activity of ETC) (26). We can hypothesize that obesity that is associated with increased mass of body fat and increased fat production in the liver has long-term consequences for the mitochondrial function and can produce a specific imprinting in skeletal muscle metabolism. However, it is not clear what is an immediate cause of dysfunction of mitochondrial ETC in obesity. It can be the result of excessive production of muscle lactate due to accelerated reduction of pyruvate and slow down the pyruvate oxidation in pyruvate dehydrogenase reaction (Fig. 4). Also, it can increase the production of methylglyoxal, which has an ability to suppress insulin signaling (22, 45). Inhibition of pyruvate dehydrogenase reaction by high NADH can also divert more pyruvate into oxaloacetate through pyruvate carboxylase. Availability of oxaloacetate and high activity of citrate synthase can provide an accelerated production of citrate from fatty acid-derived acetyl-CoA. From the other side, the consumption of citrate in TCA cycle can be decreased due to inhibition of NAD-dependent isocitrate dehydrogenase by high NADH (Fig. 4). With these considerations, we can expect an increased production of citrate by skeletal muscle mitochondria in obesity and T2DM. Also, we can expect increased citrate transport from mitochondria into the cytoplasm, where it serves in biosynthetic pathways as a precursor of acetyl-CoA, malonyl-CoA, and diacyl- and triacylglycerols. The increased content of malonyl-CoA in the muscle tissue can be an indirect indicator of increased citrate leak into the cytoplasm. Indeed, the recent study from Bandyopadhyay et al. (3) indicates the increased malonyl-CoA levels in skeletal muscle from obese and type 2 diabetic subjects. The steady-state concentration of malonyl-CoA can also depend on the rate of acetyl-CoA carboxylase reaction and on the rate of malonyl-CoA decarboxylation (49). Synthesis of malonyl-CoA by acetyl-CoA carboxylase is activated by citrate (50), and we can suggest that the rate of citrate delivery is an important factor that controls the concentration of malonyl-CoA in cytoplasm.
Excess of NADH also can interfere with β-oxidation at the level of β-HAD. This can lead to accumulation of the products of incomplete oxidation of fatty acids. Finally, the sustained high level of NADH can increase the production of intracellular hydrogen peroxide in NAD(P)H-oxidase reaction (55). Thus, the selective defect of mitochondrial electron transport chain can explain many metabolic abnormalities linked to insulin resistance: increased excretion of muscle lactate (9), accumulation of methylglyoxal (4), diacylglycerols (23), malonyl-CoA (50), and products of incomplete oxidation of fatty acids (56), and also excessive reactive oxygen species markers (61).

Combined weight loss plus exercise intervention significantly increases mitochondrial biogenesis in vastus lateralis muscle in obesity or T2DM as it follows from increase in cardiolipin, citrate synthase β-HAD, or NADH oxidase (Figs. 1 and 2). However, the increase in NADH oxidase is not sufficient to restore the normal balance between ETC and enzymes of TCA cycle or β-oxidation pathway (Fig. 2). We cannot exclude a possibility of a counterbalancing effect of caloric restriction on mitochondria. It should be taken to account that mitochondrial ETC includes proteins encoded in mitochondrial genome contrary to enzymes of TCA cycle, β-oxidation, or cardiolipin biosynthesis that solely need expression of nuclear-encoded mitochondrial genes. Additionally, skeletal muscle mitochondria in T2DM or obesity are unable to efficiently replicate mtDNA in response to exercise plus caloric restriction (Fig. 2). It can be an effect of caloric restriction; however, it is tempting to connect the “resistance” in the replication of mtDNA with reduced activity of ETC in T2DM and obesity. The simultaneous decrease in the activity of ETC with hampered replication of mtDNA can be explained by disturbed balance in the interaction of mtDNA with mitochondrial transcription factor A (TFAM). TFAM is an abundant structural protein that is essential for stability of mitochondrial genome and for compaction of mtDNA copies into mitochondrial chromosome (nucleoid). TFAM is also critical for both replication and transcription of mtDNA (39, 51). We hypothesize that an excessive generation of active byproducts of oxidative metabolism, including oxoaldehydes, can affect interaction of mtDNA with TFAM in mitochondrial chromosome and attenuate the transcription of peptides essential for respiratory complexes. The previously shown reduction in the expression of nuclear-encoded mitochondrial genes in T2DM and obesity can be a secondary event as a response to insufficient syntheses of ETC subunits encoded in mitochondrial genome (11, 30, 33, 38).

In conclusion, the presented data indicate that insulin resistance is a condition that is marked with a deficiency of ETC and disbalance between ETC, β-oxidation, and TCA cycle. Sustained exercise program or calorie restriction improves insulin resistance in human skeletal muscle (58, 59). We hypothesize that this improvement can be a result of sustained reduction in muscle NADH/NAD ratio due to increased mitochondrial respiration or reduced fatty acid delivery. Finding an appropriate intervention(s) to restore the normal enzyme profile and conceivably normalize NADH/NAD ratio in skeletal muscle mitochondria could be an important accomplishment in the treatment of T2DM and obesity by making intensive blood glucose control more safe and beneficial.

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DYSFUNCTION OF MITOCHONDRIA IN OBESITY AND DIABETES

GRANTS

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DISCLOSURES

No conflicts of interest are declared by the author(s).

REFERENCES

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