Role of PP2C in cardiac lipid accumulation in obese rodents and its prevention by troglitazone

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Wang, May-yun, and Roger H. Unger. Role of PP2C in cardiac lipid accumulation in obese rodents and its prevention by troglitazone. Am J Physiol Endocrinol Metab 288: E216–E221, 2005. First published September 14, 2004; doi:10.1152/ajpendo.00004.2004.—In obese rodents, excess myocardial lipid accumulation (lipotoxicity) of myocardium may cause cardiomyopathy that in the obese Zucker diabetic fatty (ZDF) fa/fa rat can be prevented by treatment with troglitazone (TGZ). To determine the underlying mechanisms, we measured total 5′-AMP-activated kinase (AMPK) protein and its activated, phosphorylated form, P-AMPK. P-AMPK was significantly reduced in both ZDF fa/fa rat and ob/ob mouse hearts compared with lean, wild-type controls. TGZ treatment of obese ZDF rats, which lowered cardiac lipid content, increased P-AMPK. Expression of protein phosphatase 2C (PP2C), which inactivates AMPK activity by dephosphorylation, was increased in untreated ZDF fa/fa rat hearts, but fell with TGZ treatment, suggesting that PP2C can influence AMPK activity. In cultured myocardocytes, fatty acids reduced P-AMPK, suggesting a feed-forward effect of lipid overload. Our findings highlight a role of PP2C and AMPK in the derangements of cardiac lipid metabolism and lipotoxicity of leptin-unresponsive obese rats and its prevention by TGZ. They also suggest that the phosphorylation state of the enzyme may be influenced by the expression level of PP2C.

MATERIALS AND METHODS

Animals. Male obese homozygous fa/fa Zucker diabetic fatty (ZDF) rats and their lean wild-type (+/+) littermates were bred in our laboratory from ZDF/Drt-fa (F10) rats, originally purchased from R. Peterson (University of Indiana School of Medicine, Indianapolis, IN). Male obese ob/ob mice and their wild-type control (+/+) mice (10–15 wk old) were obtained from the Jackson Laboratory (Bar Harbor, ME). All rats and mice received a standard laboratory chow (Teklad F6 8664; Teklad, Madison, WI) and tap water ad libitum. Obese fa/fa ZDF rats (6–7 wk old) were given powdered standard chow either with or without TGZ (Sankyo, Tokyo, Japan) or rosiglitazone in a dose of 130–200 mg or 15 mg/500 g chow diet, respectively, for 2 mo. Food intake was measured daily and body weight weekly. Rats were killed at 14 wk of age. While the rats were under pentobarbital sodium anesthesia, nonfasting blood samples were obtained from inferior vena cava. The heart was rapidly excised, and a portion of the left ventricle was frozen in liquid nitrogen and stored at −70°C. Institutional guidelines for animal care and use were followed. The animal protocol was approved by the Institutional Animal Care and Research Advisory Committee of University of Texas Southwestern Medical Center at Dallas.

Cell culture. Rat myocardium H9C2 cell line was purchased from ATCC (Manassas, VA) and was cultured in petri dishes in DMEM with 10% FBS at 37°C under normoxic conditions (5% CO2–20% O2), as recommended by the supplier. Cultured cells of ~90–90% confluency were starved in DMEM (5 mM glucose)-0.2% BSA without serum at 37°C for 4–6 h, followed by incubation with either glucose (5 or 20 mM) or fatty acid palmitate-BSA complexes (0.1, 0.5 or 1.5 mM palmitate at a 20:1 FFA-to-BSA molar ratio) and glucose (5

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mM) for 20 h. After treatment, the cells were washed with cold PBS two times and then lysed with 1× cell lysis buffer (Cell Signaling Technology, Beverly, MA). After being spun for 10 min at 4°C, supernatants were saved for protein concentration assays and immunoblotting analysis (see below).

Real-time quantitative PCR. Total RNA from the hearts of lean +/+ or obese fa/fa rats, fed with or without TGZ, was extracted using TRIZol reagent (Life Technologies, Torrance, CA) following the manufacturer’s protocol. For cDNA synthesis, 1 µg of rat heart total RNA was reverse transcribed with oligo(dT)18 and random hexamer primers by SuperScript II RT. Reactions were performed for 60 min at 42°C and terminated by incubating for 15 min at 70°C. Parallel reactions for each RNA sample were run in the absence of SuperScript II to assess the degree of any contaminating genomic DNA. TaqMan reactions for each RNA sample were run in the absence of SuperScript II to assess the degree of any contaminating genomic DNA. TaqMan fluorogenic probes and oligonucleotide primers were designed using Primer Express software (Perkin-Elmer Applied Biosystems, Foster City, CA). Primers and probes were ordered from Integrated DNA Technologies (Coralville, IA). The probe and primer sequences used in this study were listed in Table 1. TaqMan PCR assays for each gene target were carried out in triplicate using cDNA samples described above in 384-well optical plates with an ABI Prism 7900HT Sequence Detection system (Perkin-Elmer Applied Biosystems). For each 10-µl TaqMan reaction, 2 µl cDNA were mixed with 1 µl sense primer (200 nM), 1 µl antisense primer (200 nM), and 1 µl TaqMan fluorogenic probe (100 nM). The PCR scheme used was 50°C for 2 min, 95°C for 10 min, and then 40 cycles of 95°C for 15 s and 60°C for 1 min. The housekeeping gene used for the subsequent normalization of data in each TaqMan assay was ribosomal RNA (18S; Perkin-Elmer Applied Biosystems) or 36B4 (12). For each PCR sample, an amplification plot was generated from the collected data and a threshold cycle (CT) value was calculated with the software suite. With the use of the standard curve, CT values for each gene of interest were used to calculate the initial quantity of this cDNA present in each input. To correct for RNA quality and quantity, data were then normalized by dividing copies of the target gene by the copies of the chosen housekeeping gene, as indicated above. Relative quantitation of PP2C mRNA against 36B4 was determined using the SYBR Green PCR Master Mix and total cDNA described above.

Immunoblotting analysis. Total protein extracts prepared from heart tissues of ZDF obese fa/fa and lean +/+ rats or ob/ob and lean +/+ mice were resolved by SDS-PAGE and transferred to a polyvinylidene difluoride membrane (Amersham Biosciences, Piscataway, NJ). The blotted membrane was blocked in 1× TBS containing 0.1% Tween and 5% nonfat dry milk (TBST-MLK) for 1 h at room temperature with gentle, constant agitation. After incubation with primary antibodies anti-phospho-AMPKα (Thr172), anti-AMPKα (Cell Signaling Technology), anti-PP2C (Upstate, Lake Placid, NY), or anti-γ-tubulin (Sigma, St. Louis, MO) in freshly prepared TBST-MLK at 4°C overnight with agitation, the membrane was washed two times with TBST buffer. This was followed by incubating with secondary anti-rabbit, -mouse, or -sheep horseradish peroxidase-conjugated Ig antibodies in TBST-MLK for 1 h at room temperature with agitation. The membrane was then washed three times with TBST buffer, and the proteins of interest on immunoblots were detected by an enhanced chemiluminescence detection system (Amersham Biosciences). The corresponding bands were quantified using NIH Image software (version 1.6; available at http://rsb.info.nih.gov/nih-image/). Plasma measurements. Plasma glucose was measured by the glucose oxidase method with a colorimetric Kit (Sigma). The levels of plasma triacylglycerol (TG) were determined by a GPO-Trinder triglyceride Kit (Sigma). Plasma free fatty acid (FFA) concentration was assayed with a Wako FFA C test kit (Wako Chemicals, Richmond, VA).

Echocardiographic evaluation. Echocardiographic analysis of fa/fa ZDF rats was carried out as described (30).

Statistical analysis. Results obtained in this study are presented as means ± SE for n = 3–6 samples per group and were evaluated with Student’s t-test for statistical significance.

RESULTS

Total and P-AMPK in the hearts of ZDF (fa/fa) rats and wild-type (+/+ ) controls. To compare the activity of AMPK in normal and lipid-laden hearts of untreated obese ZDF (fa/fa) rats, we measured the levels of total AMPK protein α-subunit (AMPKα) and active, phospho-AMPKα (P-AMPKα; phosphorylated at Thr172 of the AMPK α-subunit). As shown in Fig. 1A, top and middle, the mean P-AMPKα level in these 14-wk-old, leptin-unresponsive obese fa/fa hearts was 67% lower than that of lean wild-type (+/+ ) ZDF controls (P < 0.005). Total AMPKα protein was slightly reduced in fa/fa compared with lean hearts. There were no significant changes in the mRNA of AMPK α1-, α2 (Fig. 1A, bottom), β-, or γ-subunits (data not shown).

Total and P-AMPK in the hearts of ob/ob mice and wild-type (+/+ ) controls. To determine if a similar reduction in P-AMPKα was present in the hearts of another unleptinized rodent model, the leptin-deficient ob/ob mouse, we compared them with their wild-type (+/+ ) controls. P-AMPKα was 81% lower in ob/ob mice (P < 0.0005; Fig. 1B, top and middle). There were no changes in the mRNA of AMPK α1-, α2 (Fig. 1B, bottom), β-, or γ-subunits (data not shown). Thus our

Table 1. Sequences of TaqMan probes and primers and SYBR Green PCR primers used in this study

<table>
<thead>
<tr>
<th>Name</th>
<th>GenBank No.</th>
<th>Forward/Reverse Primer</th>
<th>TaqMan Probe (5′-Fam 3′-TAMRA)</th>
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<tr>
<td>AMPKα1</td>
<td>U40819</td>
<td>TGAAGATCGGCGCCACTCATCTCT</td>
<td>ACGCTGCGGCTGCGACC</td>
</tr>
<tr>
<td>AMPKα2</td>
<td>U12149</td>
<td>GAGGCGGCGGACATG</td>
<td>GCCGCTGGCTGTCTGCTCTAG</td>
</tr>
<tr>
<td>AMPKβ</td>
<td>U42411</td>
<td>ATGAGCTGCGGATCTCT</td>
<td>CTTGACACTGTGCTGAG</td>
</tr>
<tr>
<td>AMPKγ</td>
<td>X95578</td>
<td>GACAGGCGGCGGACATCTCG</td>
<td>GACAGGCGGCGGACATCTCG</td>
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<tr>
<td>PPARγ</td>
<td>AF156666</td>
<td>TGCAAGGGCGGCGTGCTG</td>
<td>TGCAAGGGCGGCGTGCTG</td>
</tr>
<tr>
<td>36B4</td>
<td>NM_007475</td>
<td>ACCCTTCCGAGGGCTGA</td>
<td>ACCCTTCCGAGGGCTGA</td>
</tr>
<tr>
<td>PP2C*</td>
<td>J04503</td>
<td>TGCTCAGCCGCTCTCCTCCT</td>
<td>TGCTCAGCCGCTCTCCTCCTCCT</td>
</tr>
</tbody>
</table>

AMPK, 5′-AMP-activated kinase; PPAR, peroxisome proliferator-activated receptor; PP2C, protein phosphatase 2C. *SYBR Green PCR primers.
Fig. 1. Downregulation of active, phospho-(P)-5′-AMP-activated kinase (AMPKα) in obese hearts. A: top and middle: the expression of P-AMPKα and AMPKα protein in Zucker diabetic fatty (ZDF) lean (+/+ ) (open bars) and obese fafa (filled bars) rat hearts, as determined by immunoblotting analysis with anti-P-AMPKα, AMPKα, or γ-tubulin antibody. Bottom: AMPKα1 and α2 mRNA levels in +/+ (open bars) and fafa (filled bars) hearts were measured by real-time quantitative PCR analysis. B: top and middle: the expression of P-AMPKα and AMPKα protein in wild-type (+/+ ) (open bars) and ob/ob (filled bars) mouse hearts. Bottom: AMPKα1 and α2 mRNA levels in +/+ (open bars) and ob/ob (filled bars) hearts. Bars in A and B, middle, represent means ± SE (n = 4 and 6 for lean and obese groups each, respectively) of P-AMPKα protein normalized to γ-tubulin in respective groups of animals. Bars in A and B, bottom, represent means ± SE (n = 4 and 6 for lean and obese groups each, respectively) of AMPKα mRNAs normalized to 18S ribosomal RNA. *P < 0.005.

Results suggest that the reduction of cardiac AMPK activity could be a consequence of congenital lack of leptin action.

Effects of glucose and FFAs on AMPK activity of myocardial cells. The lower cardiac AMPK activation could have been secondary to the metabolic abnormalities that constitute the ZDF (fafa) phenotype. Obese ZDF fafa and ob/ob rodents have higher plasma levels of glucose and FFA than their lean counterparts (13). To determine if the decreased AMPK activity in fafa and ob/ob hearts was secondary to the high concentrations of plasma glucose and/or FFA (in complexes with BSA), we examined the expression of AMPKα in the rat myocardial H9C2 cells cultured for 20 h with varying concentrations of these nutrients. There was no difference between P-AMPKα in cells cultured in 5 or 20 mM glucose (Fig. 2). However, in medium containing 5 mM glucose in the presence of 0.5 mM FFA-BSA or above, AMPKα protein and P-AMPKα were both greatly reduced (Fig. 2). This suggests that the elevation in intracellular fatty acids or their metabolites in the tissues of unleptinized rodents lowers cardiac AMPK activity and thus amplifies the consequences of a lipid surplus by interfering with their oxidation.

The treatment of ZDF fafa rats restores cardiac AMPK activity. Because TGZ is effective in preventing lipid accumulation in the heart of ZDF (fafa) rats (30), we examined the effects of TGZ treatment on cardiac AMPK activity. Previously, we had found that in obese fafa rats TGZ lowered cardiac TG and ceramide content, preventing apoptosis and loss of contractile function (30). The rises in plasma FFA, TG, and glucose that occurred in untreated control rats were prevented by TGZ (30). The clinical and laboratory data obtained in the present study of untreated and TGZ-treated obese fafa rats are summarized in Table 2. Plasma glucose, FFA, and TG were significantly lower (P ≤ 0.005) in TGZ-treated fafa rats than those of untreated animals, consistent with our earlier results. In addition, we found that, compared with untreated fafa ZDF rats, cardiac P-AMPKα of TGZ-treated fafa ZDF rats is increased four- to fivefold (P < 0.016) with only modest change in total AMPKα protein (Fig. 3A). There was no change in the mRNA levels of AMPK α1-, α2-, β-, and γ-subunits (data not shown). These results are consistent with the hypothesis that an increased AMPK activity contributes to the prevention of excess lipid accumulation in the fafa heart by TGZ.

Relationship of PP2C to changes in AMPK activity. PP2C is known to directly dephosphorylate and thus inactivate P-AMPKα in mammals (5, 17). Therefore, it seemed possible that the reduction in active P-AMPKα in untreated ZDF fafa rats.
rats and its restoration toward near normal levels by TGZ treatment could be because of changes in this phosphatase. We therefore monitored the changes in cardiac PP2C mRNA and protein (Fig. 3, B and C) before and after TGZ treatment. PP2C expression was increased in the hearts of untreated ZDF rats compared with lean +/+ controls (Fig. 4A). After TGZ therapy, its mRNA and protein levels were decreased \( (P < 0.037 \) and 0.002, respectively; Fig. 3, B and C) concordantly with the rise in P-AMPK\(\alpha\) levels (Fig. 3A). Similarly, administration to \( fa/fa \) rats for 2 mo of another PPAR\(\gamma\) agonist, rosiglitazone, which was shown to reduce TG content in \( ob/ob \) hearts (20), led to a decrease in cardiac PP2C, an increase in P-AMPK\(\alpha\), and improved contractile performance in vivo (Table 3). These results are consistent with a role of PP2C in the change in cardiac AMPK activity in ZDF \( fa/fa \) rats. Because AMPK phosphorylation was also reduced in the hearts of obese \( ob/ob \) mice, we compared their cardiac PP2C with that of lean wild-type controls. As shown in Fig. 4B, PP2C was also increased in that obese unleptinized group. We further examined mRNA expression of cardiac PPAR\(\gamma\) and found its levels were similar in \( fa/fa \) rats and lean littermates \( (1.34 \pm 0.12 \text{ vs. } 1.04 \pm 0.28; P = 0.20) \) and were not altered in \( fa/fa \) rats by TGZ \( (1.06 \pm 0.24 \text{ for TGZ-treated vs. } 0.98 \pm 0.4 \text{ for untreated animals}) \).

**DISCUSSION**

Lipotoxic cardiomyopathy is a prominent component of the metabolic syndrome of leptin-unresponsive obese ZDF \( (fa/fa) \) rats \( (30) \). It is completely prevented by treatment with TGZ beginning at the age of 7 wk. By the age of 20 wk, untreated \( fa/fa \) rats exhibited increased myocardial TG and ceramide, with increased DNA laddering, an index of apoptosis, and decreased contractility by echocardiogram. TGZ-treated rats, by contrast, remain virtually normal in all of the foregoing respects \( (30) \). In this report, we attempt to understand the mechanisms of the increase in lipids in unleptinized myocardium and the dramatic improvement provided by treatment with the PPAR\(\gamma\) agonist.

Our results revealed that the defects of cardiac AMPK and PP2C are associated with lipotoxic cardiomyopathy in obese rodents. Although the mechanisms remain to be elucidated, it is possible that changes in AMPK and PP2C activity are attributable to lipid overload in the heart rather than to, for example, reduced contractility. Several lines of evidence support this idea. First, incubation of myocardium \( \text{H}_2\text{C}_2\text{O}_4 \) cells with fatty acid palmitate but not with glucose leads to a decrease in P-AMPK\(\alpha\), suggesting palmitate or its metabolites mediates the inhibition. Previously, the same fatty acid was also shown to impair AMPK and increase ceramide synthesis and apoptosis in rat neonatal cardiac myocytes \( (9) \). In addition, metformin, an activator of AMPK \( (14) \), can override this reduction in P-AMPK\(\alpha\) in cultured cells by fatty acids (data not shown). These results thus support cardiac lipid accumulation as a contributor to the alteration in AMPK activity. Second, cardiac AMPK activity is depressed before overt contractile dysfunction appears. In the \( fa/fa \) rat model, the earliest time point at which there was uniform evidence of heart failure in all animals was 20 wk of age \( (30) \), at least 6 wk older than the ones with reduced cardiac P-AMPK\(\alpha\) reported here. Similarly, we found that, at \( <18 \) wk of age, \( ob/ob \) hearts show excess lipid accumulation and decreased P-AMPK\(\alpha\) but no contractile

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**Table 2. Phenotypic profiles of untreated and TGZ-treated ZDF \( fa/fa \) rats**

<table>
<thead>
<tr>
<th>Parameters (ages)</th>
<th>Untreated</th>
<th>TGZ Treated</th>
<th>( P ) Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight, g (6 wk old)</td>
<td>189.3±12.2</td>
<td>191.3±7.4</td>
<td>0.7646</td>
</tr>
<tr>
<td>Body weight, g (14 wk old)</td>
<td>419.7±44.7</td>
<td>592.0±34.9</td>
<td>0.0002</td>
</tr>
<tr>
<td>Food intake, g/day (average)</td>
<td>45 ± 6.0</td>
<td>46.8 ± 3.0</td>
<td>0.7646</td>
</tr>
<tr>
<td>Plasma glucose, mg/dl</td>
<td>140.0 ± 15.0</td>
<td>142.0 ± 13.0</td>
<td>0.7646</td>
</tr>
<tr>
<td>Plasma triacylglycerol, mg/dl</td>
<td>160.0 ± 30.0</td>
<td>170.0 ± 35.0</td>
<td>0.7646</td>
</tr>
<tr>
<td>Plasma FFA, mM (14 wk old)</td>
<td>0.752 ± 0.223</td>
<td>0.201 ± 0.109</td>
<td>0.0028</td>
</tr>
<tr>
<td>Myocardial triacylglycerol†</td>
<td>100%</td>
<td>59%</td>
<td>&lt;0.01</td>
</tr>
</tbody>
</table>

Data are means ± SE; \( n = 6 \) rats in each group. TGZ, troglitazone; ZDF, Zucker diabetic fatty; FFA, free fatty acid. Data were analyzed by Student’s \( t \)-test. *Animals were untreated or treated with TGZ for 2 mo. †From Ref. 30. **
dysfunction (data not shown; see Ref. 2). Heart failure became evident only in 24-wk-old ob/ob mice (2). In this regard, it is conceivable that the accumulation of lipids, the loss of function, and the apoptosis are likely not happening simultaneously in all cells. The lipids accumulate in a few cells initially, and those are the cells that will deteriorate first. Only when a substantial dropout of cardiomyocytes has occurred will echocardiographic evidence of failure be evident.

Our findings in fa/fa and ob/ob hearts are different from that reported in insulin-resistant JCR:LA-cp rats, which showed slightly decreased cardiac AMPK activity in spite of elevated TG in the heart (1). The reasons for the discrepancy are not clear. Notable differences are that cp rats seem to have less dyslipidemia compared with fa/fa and ob/ob animals and have no cardiac dysfunction or diabetes (1). TGZ or metformin treatment did not affect plasma lipid profiles in these rats (24). Some technical issues also make the comparison less straightforward. For instance, in their study, AMPK activity was measured with cytosolic fractions recovered from polyethylene glycol precipitation (1).

The report by Fryer et al. (7) that rosiglitazone activates AMPK in skeletal muscle suggested that a similar mechanism might prevail cardiac muscle as well. Because the obese human heart, like the heart of obese ZDF rats, has increased intramyocardial lipids (25, 26), the present findings may have clinical therapeutic implications for obese humans (25).

In fact, a reduction in active P-AMPK was observed in untreated ZDF rats and ob/ob mice in association with an increase in PP2C, a protein phosphatase that is known to inactivate it (5, 17). Treatment with TGZ or rosiglitazone increased P-AMPKα to near-normal levels, and PP2C was reduced. Thus PP2C was increased in both models, and, in the thiazolidinedione-treated ZDF rat, it declined toward normal as P-AMPKα increased. Because the fatty acid-induced increase in PP2C has been linked to apoptosis in cultured chick neurons (11), whereas AMPK activation inhibits ceramide synthesis and apoptosis in astrocytes (3) and apoptosis in INS-1 cells (6), suppression of PP2C activity in obesity might provide a useful therapeutic target.

It is not clear from these results if the TGZ effects on the heart represent a direct action in the cardiomyocytes or are secondary to the lipopenic action of the drug. TGZ may exert its effects on PP2C and AMPK directly at the level of the heart. Treatment with TGZ in rat myocardium H9C2 cells for 24 h showed a decrease in PP2C, concurrent with an increase in P-AMPKα (data not shown), the same as that seen in the hearts of TGZ or rosiglitazone-treated obese rodents. PPARγ agonist Wy-14643, however, has no effects on PP2C and P-AMPKα levels in H9C2 cells (data not shown). Furthermore, in skeletal muscle, AMPK can also be stimulated by rosiglitazone (7). These findings are concordant with the concept that thiazolidinediones can act directly on the cardiac myocytes via PPARγ-dependent pathways. It is noteworthy that a putative PPARγ RXR response sequence (AGGTCGAAGGGCA) can be identified on the rat PP2C promoter (located at −728 to −716 of transcription start site). The cardiac PPARγ pathway in obese rodents seems to be normal, since PPARγ mRNA is similar in fa/fa rats or ob/ob mice and lean controls. Alternatively, TGZ may modulate the fluxes of lipids through the heart of treated animals (10, 22) by which cardiac PP2C and AMPK activity somehow can be normalized. Stimulation of AMPK in turn would increase fatty acid oxidation (15, 16) and reduce lipid content in cardiac muscle. Further studies are needed to assess these possibilities.

In summary, our results support a role of AMPK and PP2C defects in the dysfunction of cardiac lipid metabolism in obesity and provide new insights as to the mechanisms of the liporegulatory disorder leading to lipotoxic cardiomyopathy.

### ACKNOWLEDGMENTS

We thank Christie Fisher for excellent secretarial work and Daniel J. Garry, R. Haris Naseem, and Maggie Robledo for echocardiographic studies.

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**Table 3. Phenotypic profiles of untreated and RSG-treated ZDF fa/fa rats**

<table>
<thead>
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<th>Parameters</th>
<th>Untreated</th>
<th>RSG Treated</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma glucose, mg/dl</td>
<td>679.6±70.9</td>
<td>269.2±37.4</td>
<td>0.0003</td>
</tr>
<tr>
<td>Plasma triacylglycerol, mg/dl</td>
<td>485.3±203.3</td>
<td>72.7±47.3</td>
<td>0.0238</td>
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<tr>
<td>Fractional shortening*, %</td>
<td>36.5±3.1</td>
<td>56.3±6.7</td>
<td>0.0033</td>
</tr>
<tr>
<td>Cardiac PP2C protein (arbitrary unit)</td>
<td>1.00±0.49</td>
<td>0.13±0.06</td>
<td>0.026</td>
</tr>
<tr>
<td>Cardiac P-AMPKα (arbitrary units)</td>
<td>1.01±0.35</td>
<td>1.95±0.17</td>
<td>0.057</td>
</tr>
</tbody>
</table>

Data are means ± SE; n = 7 rats in each group. RSG, rosiglitazone; PP2C, protein phosphatase 2C; P-AMPKα, phosphorylated AMPKα. Data were analyzed by Student’s t-test. * Animals were untreated or treated with RSG for 2 mo. †An index of contractile function of the heart, measured by echocardiography. ‡Determined by immunoblotting analysis.
REFERENCES


