Low-dose dexamethasone in the rat: a model to study insulin resistance

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Severino, C., P. Brizzi, A. Solinas, G. Secchi, M. Maioli, and G. Tonolo. Low-dose dexamethasone in the rat: a model to study insulin resistance. Am J Physiol Endocrinol Metab 283: E367–E373, 2002. First published February 5, 2002; 10.1152/ajpendo.00185.2001.—The main aim of this study was to set up a new animal model to study insulin resistance. Wistar rats (6 or 7 per group) received the following for 4 wk in experiment 1: (1) vehicle, (2) 2 μg/day subcutaneous dexamethasone, (3) metformin (400 mg·kg−1·day−1 os), and (4) dexamethasone plus metformin. In experiment 2 the rats received the following: (1) vehicle, (2) dexamethasone, (3) dexamethasone plus arginine (2%; as substrate of the nitric oxide synthase for nitric oxide production) in tap water, and (4) dexamethasone plus isosorbide dinitrate (70 mg/kg; as direct nitric oxide donor) in tap water. Insulin sensitivity was significantly reduced by dexamethasone already at week 1, before the increase in blood pressure (day 15) and without significant changes in body weight compared with vehicle. Dexamethasone-treated rats had significantly higher triglycerides, hematocrit, and insulin, whereas serum total nitrates/nitrites were lower compared with vehicle. The concomitant treatment with metformin minimized all the described effects of dexamethasone. In experiment 2, only isosorbide dinitrate was able to prevent the observed dexamethasone-induced metabolic, hemodynamic, and insulin sensitivity changes. Chronic low-dose subcutaneous dexamethasone (2 μg/day) is a useful model to study the relationships between insulin resistance and blood pressure in the rat, and dexamethasone might decrease insulin sensitivity and increase blood pressure through an endothelium-mediated mechanism.

THERE IS CONSIDERABLE evidence that abnormalities of glucose, insulin, and lipoprotein metabolism occur more frequently in untreated hypertensive patients than in normotensive subjects. It could be argued that the relationship between high blood pressure and metabolic disorder is incidental, but, on the other hand, there is evidence that changes in glucose, insulin, and lipoprotein metabolism play a role in the etiopathology and/or clinical course of hypertension. In human hypertension, subtle changes in adrenal steroid metabolism have been suggested (28, 29, 36), indicating that glucocorticoids, besides the well-known Cushing syndrome, might have an important role in the development of high blood pressure. Moreover, glucocorticoids have been shown to reduce cellular glucose uptake affecting the glucose transport system per se (7), with no direct effects on the insulin receptor (26). Thus glucocorticoids alter glucose metabolism, and in turn they have a role in the development of peripheral insulin resistance. Insulin resistance may be an impetus for the development of hypertension, impaired carbohydrate tolerance, and lipid alteration, but the underlying mechanisms are still unclear. Similar metabolic abnormalities occur in rodent models of hypertension. For example, endothelial dysfunction precedes hypertension in an experimental model of fructose-induced insulin resistance (9, 24), but fructose-induced insulin resistance is not easily comparable to humans. At the best of our knowledge, there is no established model of insulin resistance induced by dexamethasone without the dramatic catabolic side effects commonly seen with glucocorticoids. We previously showed that it is possible to increase blood pressure for long term in rats with subcutaneous doses of dexamethasone on the order of micrograms per day (31) without appreciable catabolic effects. This animal model is not associated with sodium retention but with sodium shift from the intracellular to the extracellular space (12), and the effects on blood pressure are opposite to those obtained by intracerebroventricular dexamethasone administration (32). The main aim of this study was to establish if our previously described “old model” of glucocorticoid-induced hypertension in the rat is indeed a useful “new” model to study insulin resistance in the metabolic syndrome. To do so we used our previously described animal model (31) to evaluate the effects of metformin, a well-established drug able to ameliorate insulin sensibility in rats (25).

RESEARCH DESIGN AND METHODS

Experiment 1

Animals. Male Wistar rats, with a weight of 400 g, were used through the experiments in groups of 6 or 7 animals
each. All rats were housed in an automatically light-controlled animal facility (12 h on, 12 h off) with constant temperature (22°C) and humidity, with free access to food (Mil mice and rats GLP Diets, Mucedola Srl, Italy) and tap water ad libitum. Two weeks before start of the experiment, animals were accustomed to handling as well as blood pressure and blood glucose measurements (by a vein puncture in the tail). Four groups were thereafter treated for 4 wk: 1) vehicle: tap water and daily subcutaneous injection of 0.9% NaCl (75 μl) at 8:00 AM and 8:00 PM; 2) dexamethasone (Dex): tap water and daily subcutaneous injections of Dex (1 μg in 75 μl 0.9% NaCl) at 8:00 AM and 8:00 PM; 3) metformin (Met): Met dissolved in tap water (3.5 mg/ml) and daily subcutaneous injection of 0.9% NaCl (75 μl) at 8:00 AM and 8:00 PM; and 4) Dex + Met: Met dissolved in tap water (3.5 mg/ml) and daily subcutaneous injection of Dex (1 μg in 75 μl 0.9% NaCl) at 8:00 AM and 8:00 PM.

Methods. Three times a week systolic blood pressure was measured in the morning (tail cuff method, Letica, Le 5001 pressure meter) in the conscious lightly restrained animal after the animals were prewarmed at 38°C for 10 min as previously described (12, 31, 32); body weight and tap water consumption were recorded at the same time. At days +8, +14, and +26 after blood pressure measurement, in the conscious rats blood from a tail vein was obtained for glucose measurement with a reflectometer (Lifescan One Touch Profile, Johnson-Johnson, Minneapolis, MN), immediately before and 30 min after rats received intraperitoneal fast-acting insulin (1.6 U/kg Actrapid, Novo Nordisk). In this “insulin-tolerance test,” the 1.6-U/kg dose and the 30-min values were chosen to calculate the maximum decrease in blood glucose. This method was, in our laboratory, repetitive as shown in Fig. 1, where the effects of three different doses of intraperitoneal insulin (0, 0.8, and 1.6 U/kg) on blood glucose, over 80 min after insulin injection, are given. The reflectometer was validated in our laboratory against an automatic analyzer (Hitachi 912, automatic analyzer, Boehringer Mannheim, using standard reagents) for values from 30 to 200 mg/dl (n = 68, r² = 0.921 P < 0.001).

At day +28 after the last blood pressure measurement, while the rats were fasting from midnight, under pentobarbital sodium anesthesia a large blood sample from the abdominal aorta was obtained. Serum cholesterol, triglycerides, β-hydroxybutyrate and uric acid (Hitachi 912), Na⁺, and K⁺ (flame photometer, Beckman) were measured. One hundred fifty microliters of blood were collected in a microhematocrit capillary tube, spun at 5,000 rpm for 10 min, and used for the determination of hematocrit. Insulin was measured in duplicate by use of a rat insulin radioimmunoassay kit (Diasorin, Stillwater, Minnesota, MN). Total nitrates/nitrates were measured by a validated colorimetric assay kit (Cayman) with high sensibility (1 μM) for both NO₂⁻ and NO₃⁻ in serum to achieve an estimate of in vivo nitric oxide (NO) production, with nitrates (NO₂⁻) and nitrates (NO₃⁻) the end product of NO (13). Intraerythrocyte Na⁺ was measured following a published method (5) with the only modification being the use of 0.5 ml full blood. Blood for free fatty acids was obtained at the same time and collected in iced (+4°C) EDTA tubes; it was immediately centrifuged at 4°C, and the plasma was frozen at −80°C until assayed. Free fatty acids were measured by a colorimetric method using a Boehringer-Mannheim kit.

Heart, liver, and the left kidney were excised and weighed. The measurement was corrected for body weight.

Histology. Liver and left kidney tissues after fixation in 10% buffered formalin were dehydrated with ethyl alcohol and then included in paraffin. Sections of 5 μm were obtained by a microtome (Top Rotary S-130, pabish). Periodic acid-Schiff coloration for polysaccharides has been used to evaluate glycogen content in the hepatocytes. Two independent observers not aware of the different treatments gave independent comment on the histological material.

Steady-state glucose concentration. Steady-state glucose concentration (24) was measured to confirm the findings of the insulin-tolerance test in the period preceding the rise in blood pressure observed in Dex-treated rats. Steady-state glucose concentration was measured in an additional four rats after 8 days of Dex treatment alone (before any significant rise in systolic blood pressure) or with Met following the described protocol of Reaven et al. (24). In brief, under anesthesia with pentothal sodium (Tiopentale Sodico 0.5 g, Farmaceutici Gellini), the right femoral vein was cannulated for insulin and glucose infusion at the fixed doses of 2.5 mU·kg⁻¹·min⁻¹ and 8 mg·kg⁻¹·min⁻¹, respectively, for 3 h. Tail blood samples for glycemia were taken at 15-min intervals during the last hour of infusion, and the four obtained values were used to find the mean.

Experiment 2

Animals and methods. To better characterize the possible endothelial involvement in the development of insulin resistance in this animal model, four additional groups of six animals each were compared in a 28-day experiment. The main reason for this second study was to evaluate if NO production (measured, as said above, as total nitrates and nitrates) was restored giving exogenously the endogenous substrate for NO production, arginine. The arginine effects were compared with those obtained by exogenous direct NO gift through a NO donor, isosorbide dinitrate.

The experimental groups were as follows: 1) vehicle: tap water and daily subcutaneous injection of 0.9% NaCl (75 μl) at 8:00 AM and 8:00 PM; 2) Dex: tap water ad libitum and daily subcutaneous injections of Dex (1 μg in 75 μl of 0.9% NaCl) (Met): Met dissolved in tap water (3.5 mg/ml) and daily subcutaneous injection of 0.9% NaCl (75 μl) at 8:00 AM and 8:00 PM; and 3) metformin (Met): Met dissolved in tap water (3.5 mg/ml) and daily subcutaneous injection of Met (1 μg in 75 μl 0.9% NaCl) at 8:00 AM and 8:00 PM.

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The experimental groups were as follows: 1) vehicle: tap water and daily subcutaneous injection of 0.9% NaCl (75 μl) at 8:00 AM and 8:00 PM; 2) Dex: tap water ad libitum and daily subcutaneous injections of Dex (1 μg in 75 μl of 0.9% NaCl)
NaCl) at 8:00 AM and 8:00 PM; 3) Dex + arginine (Arg): daily subcutaneous injections of Dex (1 μg in 75 μl of 0.9% NaCl) at 8:00 AM and 8:00 PM and 2% Arg in tap water ad libitum; 4) Dex + isosorbide dinitrate (Isn): daily subcutaneous injections of Dex (1 μg in 75 μl of 0.9% NaCl) at 8:00 AM and 8:00 PM and 70 mg/kg Isn in tap water ad libitum.

Thus, in this second experiment, Arg was given as substrate of NO synthase for the production of NO, while Isn was given as a direct NO donor to establish the role of endothelium in this animal model. Animals were studied as in experiment 1 including insulin-tolerance test evaluation. After 1 and 4 wk of treatment, immediately after blood pressure measurements, 2 ml of blood were obtained from the tail in the conscious rat for biochemistry determination and hematocrit.

**Statistical Analysis**

Data are presented as means ± SE. After ANOVA measurements, with pairwise Newman-Keuls test for multiple comparisons, parameters found significantly different were subsequently analyzed with the paired (within groups, different times) or unpaired (between groups, same time) two-tailed Student’s t-test when appropriate. *P* < 0.05 was taken as significant. Data were analyzed using the Sigma Stat 3.0 program.

Principles of laboratory animal care [Department of Health and Human Services Publication No. (NIH) 85–23, Revised 1985] were followed in these experiments.

**RESULTS**

**Experiment 1**

All rats completed the 28 days of experiments. Water consumption was equal in the groups (32–38 ml·day⁻¹·rat⁻¹), apart from a slight decrease limited only between days +1 and +2 for rats receiving Met. Both Dex + Met and Met rats drank 21–26 ml·day⁻¹·rat⁻¹ during these days, probably due to the rats becoming accustomed to the drug taste; thereafter, the daily water intake was similar to that of the other groups. Thus the average Met intake in the Dex + Met and Met rats was 183–228 mg·kg⁻¹·day⁻¹ for days 1 and 2 and 280–333 mg/kg for the remaining days. Systolic blood pressure (SBP) increased significantly only in Dex rats from day +15, reaching the zenith at day +18 and remaining significantly elevated until day +28, with a net increase of >20 mmHg from basal (Fig. 2A). The remaining rats did not show significant changes in SBP through the study, although final SBP in Dex + Met rats was somehow 5 mmHg more elevated, but not significantly, than in the other two remaining groups (vehicle and Met). Heart rate did not show significant difference during the study in the four groups of rats (Table 1). Body weight did not change significantly through the study in any group (Table 1). At the end of the 4 wk, serum insulin was significantly increased in Dex rats and decreased in Met rats compared with vehicle, whereas blood glucose was equal in the four groups (Table 1). Dex-treated rats showed also significantly lower (*P = 0.032*) levels of serum total nitrates/nitrites, whereas hematocrit, cholesterol, triglycerides, free fatty acids, and insulin were significantly higher compared with vehicle (Table 1). These effects were completely reversed by the concomitant use of Met (Dex + Met), but serum cholesterol remained higher than in vehicle. No significant changes for the other considered parameters were evident among the different groups of animals, including intraerythrocyte Na⁺ (12.1 ± 1.3, 10.4 ± 0.5, 9.8 ± 0.8, 12.5 ± 1.4, meq/10⁶ red blood cells for vehicle, Met, Dex, and Dex + Met rats, respectively).

Insulin sensitivity as estimated by the 30-min drop of blood glucose after intraperitoneal fast-acting insulin was significantly reduced in Dex-treated rats compared with the other groups at days +8, +12, and +26 (Fig. 2B). Similar results were obtained when insulin sensitivity was estimated by the steady-state plasma glucose during insulin/glucose infusion. Mean steady-state glycemia during the third hour of insulin/glucose infusion was significantly higher in Dex rats compared with Dex + Met rats: 128 ± 6 vs. 84 ± 5 mg/dl (*P* <
0.02), while serum insulin at the end of the 3-h infusion was comparable in the two groups.

At the end of the study, no significant difference in heart, liver, and kidney weight (corrected for body weight) was evident in the four groups of rats. Glycogen content, determined histologically in the hepatocytes, was reduced in Dex-treated rats compared with vehicle and Met-treated rats. Moreover, in Dex-treated rats, an increased deposit of lipids at the liver level was evident (Fig. 3). The addition of Met in Dex rats partially restored the content of glycogen at the liver level, dramatically reducing the liver lipids content. No significant differences in the different groups of rats were evident regarding kidney structure, although the glomerular apparatus in Dex rats appeared bigger than in the other groups (data not shown).

**Experiment 2**

All animals completed the 28 days of experiment, and water consumption was equal in the groups (30–37 ml·day⁻¹·rat⁻¹) without significant changes in rats given both Arg or Isn compared with vehicle or Dex. Rats treated with Dex + Isn did not show any significant change in SBP, heart rate, and body weight through the 28 days of experiments compared with vehicle (data for final measurement at day 28 are shown in Table 2). Isn treatment in addition to Dex was also able to completely reverse the effects of Dex treatment in terms of triglycerides, hematocrit, and delta glycemia changes after intraperitoneal insulin (Table 2). On the other hand, Dex + Arg rats did not significantly differ from Dex-only rats (Table 2). Thus only the direct NO donor Isn given in association with Dex was able to restore the conditions seen in the vehicle group.

**DISCUSSION**

In this paper we describe for the first time, to the best of our knowledge, a new animal model of glucocorticoid-induced insulin resistance possibly due to endo-

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**Table 1. Body weight, heart rate, biochemical serum parameters, and hematocrit at the end of the 4-wk experiment in the different groups of rats (experiment 1)**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Vehicle (g)</th>
<th>Met (g)</th>
<th>Dex (g)</th>
<th>Dex + Met (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight</td>
<td>415 ± 18</td>
<td>400 ± 21</td>
<td>390 ± 17</td>
<td>395 ± 22</td>
</tr>
<tr>
<td>Heart rate, beats/min</td>
<td>330 ± 25</td>
<td>340 ± 30</td>
<td>345 ± 28</td>
<td>365 ± 35</td>
</tr>
<tr>
<td>Hematocrit, %</td>
<td>40.2 ± 0.5</td>
<td>40.4 ± 0.2</td>
<td>42.0 ± 0.3*</td>
<td>41.4 ± 0.7*</td>
</tr>
<tr>
<td>Total cholesterol, mg/dl</td>
<td>70 ± 6</td>
<td>72 ± 3</td>
<td>99.6 ± 9†</td>
<td>115 ± 10†</td>
</tr>
<tr>
<td>Triglycerides, mg/dl</td>
<td>142 ± 18</td>
<td>121 ± 17</td>
<td>249 ± 20†</td>
<td>136 ± 18</td>
</tr>
<tr>
<td>Free fatty acids, mM</td>
<td>0.29 ± 0.14</td>
<td>0.25 ± 0.14</td>
<td>0.81 ± 0.18†</td>
<td>0.18 ± 0.08†</td>
</tr>
<tr>
<td>β-hydroxybutyrate, mM</td>
<td>0.76 ± 0.3</td>
<td>0.72 ± 0.4</td>
<td>0.86 ± 0.4</td>
<td>0.79 ± 0.5</td>
</tr>
<tr>
<td>Insulin, μU/ml</td>
<td>22 ± 2.1</td>
<td>14 ± 0.9‡</td>
<td>32 ± 2.3*</td>
<td>21 ± 1.9</td>
</tr>
<tr>
<td>Glucose, mg/dl</td>
<td>77 ± 4.5</td>
<td>78 ± 2.4</td>
<td>75 ± 4.8</td>
<td>73 ± 3.2</td>
</tr>
<tr>
<td>Uric acid, mg/dl</td>
<td>1.1 ± 0.1</td>
<td>0.7 ± 0.1</td>
<td>0.9 ± 0.1</td>
<td>1.1 ± 0.1</td>
</tr>
<tr>
<td>Nitrates/nitrites, μmol/ml</td>
<td>78 ± 9</td>
<td>62 ± 9</td>
<td>42 ± 10*</td>
<td>63 ± 5</td>
</tr>
</tbody>
</table>

Values are means ± SE. Met, metformin; Dex, dexamethasone. *P < 0.05, †P < 0.01 vs. vehicle. ‡P < 0.05 vs. Dex.
An Insulin-Resistance Model

Table 2. Principal biochemical serum parameters, systolic blood pressure, and body weight in the four groups of animals at day 7 and at the end of the 28 days of experiment 2

<table>
<thead>
<tr>
<th></th>
<th>Vehicle</th>
<th>Dex</th>
<th>Dex + Arg</th>
<th>Dex + Ins</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>+7</td>
<td>+28</td>
<td>+7</td>
<td>+28</td>
</tr>
<tr>
<td>SBP, mmHg</td>
<td>118 ± 2</td>
<td>119 ± 3</td>
<td>120 ± 5</td>
<td>151 ± 6†</td>
</tr>
<tr>
<td>Body weight, g</td>
<td>405 ± 25</td>
<td>400 ± 21</td>
<td>386 ± 23</td>
<td>388 ± 30</td>
</tr>
<tr>
<td>Glycemia, mg/dl</td>
<td>75 ± 3</td>
<td>78 ± 4</td>
<td>72 ± 4</td>
<td>73 ± 4</td>
</tr>
<tr>
<td>Insulin, µU/ml</td>
<td>23 ± 20</td>
<td>21 ± 19</td>
<td>38 ± 2.3*</td>
<td>33 ± 2.5*</td>
</tr>
<tr>
<td>Δ% G</td>
<td>-44 ± 4</td>
<td>-41 ± 5</td>
<td>-27 ± 5*</td>
<td>24 ± 4*</td>
</tr>
<tr>
<td>TG, mg/dl</td>
<td>138 ± 19</td>
<td>232 ± 21‡</td>
<td>215 ± 20‡</td>
<td>135 ± 20</td>
</tr>
<tr>
<td>FFA, mM</td>
<td>0.24 ± 0.13</td>
<td>0.79 ± 0.15†</td>
<td>0.68 ± 0.16</td>
<td>0.23 ± 0.10</td>
</tr>
<tr>
<td>Nitrates/nitrites, µmol/ml</td>
<td>64 ± 7</td>
<td>80 ± 8</td>
<td>35 ± 10*</td>
<td>41 ± 10*</td>
</tr>
<tr>
<td>Hematocrit, %</td>
<td>40.3 ± 0.7</td>
<td>40.0 ± 0.8</td>
<td>41.8 ± 0.5†</td>
<td>43.2 ± 0.4†</td>
</tr>
</tbody>
</table>

Values are means ± SE. +7 and +28 refer to treatment days. SBP, systolic blood pressure; TG, triglycerides; FFA, free fatty acids; Δ% G, blood glucose variation (% from basal) after 1.6 U insulin Actrapid ip. *P < 0.05, †P < 0.01, ‡P < 0.001 vs. vehicle (same day).

Insulin resistance, before the development of type 2 diabetes and/or hypertension, may cause endothelial dysfunction with a key role in the pathogenesis of vascular complications (8, 33). Metformin is able to increase peripheral insulin sensitivity and insulin-mediated glucose uptake in the cells, increasing insulin-induced translation of GLUT4 from an intracellular pool to the plasma membrane and increasing the functional activity of the glucose carrier without altering the de novo synthesis of the glucose carrier both in vivo (15) and in vitro (15). In this way, metformin might reverse the concomitant endothelial dysfunction. Indeed, recently, the United Kingdom Prospective Diabetes Study (UKPDS) has shown that metformin treatment in overweight type 2 patients reduces significantly the occurrence of myocardial infarction (35), compared with sulfonylureas or insulin in normal-weight type 2 patients (34), suggesting additional effects of metformin (2) in addition to lowering glycemia. It is well known that glucocorticoids alter insulin sensitivity, and they have a role in altering glucose metabolism and blood pressure regulation. Most of the already described glucocorticoid animal models do not...
take into account catabolism, with muscle atrophy (6, 17, 37) and increased blood pressure that can affect the measurement of insulin sensitivity by altering peripheral blood flow. Moreover, no data on the long-term glucocorticoid treatment are available as well regarding a possible endothelial involvement. We chose metformin as a well-established (25), both in vivo and in vitro, drug therapy to ameliorate peripheral insulin resistance in rats as well in humans. Its wide use in diabetes is mainly due to its effects on peripheral insulin action to increase glucose uptake and utilization (30), although recently a direct effect of metformin on restoring β-cell insulin secretion response in vitro has been described. The reduction in blood pressure induced by metformin in insulin-resistant rats is apparently through a direct mechanism with a NO-dependent relaxation (10). Indeed, metformin has been shown able to attenuate the development of hypertension in the spontaneously hypertensive rats (19) usually reported to be insulin resistant (4, 17). These effects of metformin on blood pressure are not present in insulin resistance (38), indicating that metformin is not able per se to decrease blood pressure. An additional alternative “nonesterified fatty acid hypothesis” may be added to explain glucocorticoid-induced insulin resistance (17). Our data show that dexamethasone treatment increases free fatty acids in plasma, and this might have contributed to insulin resistance, while metformin treatment restores free fatty acid metabolism (21), as we found in this experiment. Metformin increases the effects of insulin on lowering blood pressure, decreasing platelet aggregation and blood viscosity in non-insulin-dependent diabetes mellitus patients. We did not measure blood viscosity in our experiment, but metformin plus dexamethasone rats showed, compared with dexamethasone-only rats, lower hematocrit levels, lower triglycerides, and lower free fatty acids, all features that can explain a decrease in blood viscosity. Feeding rats with erides, and lower free fatty acids, all features that can be observed with dexamethasone treatment and reversed by the concomitant use of metformin. Thus we postulate that dexamethasone, by altering NO synthase expression, might alter lipid metabolism with an increase in free fatty acid and consequently insulin resistance. Because in human hypertension subtle changes in adrenal steroid metabolism have been suggested (28, 29, 36), indicating that glucocorticoids in addition to the well-known Cushing syndrome might have an important role in the development of high blood pressure, this animal model is useful to study this aspect of human hypertension.

In conclusion, the two most important findings of this study are as follows: 1) long-term low-dose subcutaneous dexamethasone induces insulin resistance that precedes hypertension, and both conditions are reversed by the concomitant treatment with metformin; and 2) dexamethasone-induced insulin resistance might be due to endothelial dysfunction. Our data indicate that low-dose dexamethasone-induced hypertension in rats, an old model (12, 31, 32) of hypertension, can be revisited as an endothelial dysfunction causing insulin resistance and consequently some of the commonly observed features of the metabolic syndrome (hypertension, lipid-metabolism alteration, fatty liver, etc.). In particular, the presence also of the hepatic consequences of the insulin resistance in humans (fatty liver) appears to complete the picture of metabolic syndrome due to reduced insulin sensitivity. This model would be useful for studying insulin resistance-blood pressure relationships.

We acknowledge the excellent technical work of Maristella Spissu and Gianluigi Fenu.

C. Severino is at present working in the Servizio Diabetologia, Dipartimento Struttura Clinica Medica e Patologia Speciale Medica, University of Sassari as a postdoctorate fellow.

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