Impairment of albumin and whole body postprandial protein synthesis in compensated liver cirrhosis

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Tessari, P., R. Barazzoni, E. Kiwanuka, G. Davanzo, G. De Pergola, R. Orlando, M. Vettore, and M. Zanetti. Impairment of albumin and whole body postprandial protein synthesis in compensated liver cirrhosis. Am J Physiol Endocrinol Metab 282:E304–E311, 2002; 10.1152/ajpendo.00333.2001.—To investigate the anabolic effects of feeding in cirrhosis, we measured albumin fractional synthesis rate (FSR) and whole body protein synthesis in six nondiabetic patients with stable liver cirrhosis (three in the Child-Pugh classification Class A, three in Class B) and in seven normal control subjects, before and after administration of a 4-h mixed meal. Leucine tracer precursor-product relationships and whole body kinetics were employed at steady state. Basal levels of postabsorptive albumin concentration and FSR, whole body leucine rate of appearance, oxidation, and nonoxidative leucine disposal (NOLD, =protein synthesis) were similar in the two groups. However, after the meal, in the patients neither albumin FSR (from 8.5 ± 1.5 to 8.8 ± 1.8 %/day) nor NOLD (from 1.69 ± 0.22 to 1.55 ± 0.26 μmol·kg⁻¹·min⁻¹) changed (P = nonsignificant vs. basal), whereas they increased in control subjects (albumin FSR: from 10.9 ± 1.5 to 15.9 ± 1.9 %/day, P < 0.002; NOLD: from 1.80 ± 0.14 to 2.10 ± 0.19 μmol·kg⁻¹·min⁻¹, P = 0.032). Thus mixed meal ingestion did not stimulate either albumin FSR or whole body protein synthesis in compensated liver cirrhosis. The mechanism(s) maintaining normoalbuminemia at this disease stage need to be further investigated.

LIVER CIRRHOSIS is a protein-catabolic disease, which at advanced stages is usually associated with loss of lean body mass (represented mainly by proteins and water), decreased albumin concentration, and an increased ratio of aromatic (AAA) to branched-chain amino acid (BCAA) concentrations (18, 23, 24, 39). These metabolic alterations reflect the impaired liver functional capacity and can heavily complicate the clinical course of the disease (21).

The maintenance of protein homeostasis is achieved physiologically through meal ingestion and assimilation, which counteract the interprandial catabolic phases (8, 20, 37). One key component of the anabolic response to a meal is the stimulation of protein synthesis. Indeed, in healthy humans, intake of a mixed meal of adequate amino acid and caloric content results in the acute increase of whole body, albumin, and muscle protein synthesis (8, 20, 37). However, whether these effects are preserved in cirrhosis has not been thoroughly investigated. In a previous study (3), albumin synthesis was found to be grossly unaltered in postabsorptive cirrhotic patients, but it correlated inversely with the patients’ clinical status. However, the response to a mixed meal was not tested. In another study (22), no stimulation by an enteral diet of whole body protein synthesis was found in cirrhosis, although the caloric and amino acid content might have been insufficient to elicit a response (37). As a matter of fact, no study has yet investigated whether a balanced mixed meal of adequate caloric intake is capable of stimulating both whole body and albumin protein synthesis in liver cirrhosis.

The changes in body protein content as well as in plasma protein and amino acid concentrations that occur in advanced cirrhosis likely represent the endpoint of long-lasting alterations. Thus it is of interest to investigate whether the mechanism(s) controlling protein synthesis postprandially in cirrhosis are already altered at an early disease stage. To this aim, we have measured the response of both whole body protein and plasma albumin synthesis to mixed meal ingestion in nondiabetic, stable cirrhotic patients and in normal subjects by use of leucine tracer precursor-product relationships and whole body kinetics.

METHODS

Subjects. Six male patients with established liver cirrhosis (CP) and seven male normal control subjects (N) were recruited. All subjects were informed about the aims of the study and signed their consent to it. The study was approved by the Ethics Committee of the Medical Faculty at the University of Padova, Italy, and it was performed according to

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the Helsinki Declaration (as revised in 1983) and following the recommendations of the local Radiation Safety Officer. Whole body radiation exposure from the [14C]leucine isotope was estimated to be 1 mGy (7). All cirrhotic subjects were ambulatory and in good health except for their liver disease. All subjects were adapted to a standard weight-maintaining diet (25–30 kcal/body wt) containing ~50% of calories as carbohydrates, ~20% as proteins, and ~30% as lipids for ≥1 yr before the study. The diagnosis of cirrhosis was established through medical history, clinical examination, ultrasound criteria, biochemical data, and laparoscopy in all patients. In addition, in five patients a liver biopsy was performed, and a chronic hepatitis C virus (HCV) infection was diagnosed. In the sixth, nonbiopsied patient, cirrhosis was defined as cryptogenic. Only one patient had (minimal) HCV infection was diagnosed. In the sixth, nonbiopsied patient, cirrhosis was confirmed also by ultrasound. In the patients, a liver biopsy was performed, and a chronic hepatitis C virus (HCV) infection was diagnosed. In the sixth, nonbiopsied patient, cirrhosis was defined as cryptogenic. Only one patient had (minimal) HCV infection diagnosed. In the sixth, nonbiopsied patient, cirrhosis was confirmed also by ultrasound. In all patients, ascites was excluded through both clinical and ultrasonographic criteria. No patient had any sign of hepatic encephalopathy or was taking any pharmacological treatment (30).

In these patients, DEXA-determined LBM (56 ± 0.72 kg) was directly measured also by Hume’s formula. In the patients, body mass index (BMI, expressed as weight in kg divided by squared height in meters) and LBM were slightly greater (P < 0.05) than those measures in control subjects (Table 1), whereas the fraction of body weight accounted for by LBM was lower (−7%, P < 0.05) (Table 1).

Because glucose intolerance and/or diabetes is frequent in cirrhosis, and it may determine alterations in both whole body and albumin protein turnover, an oral glucose tolerance test was performed in all patients to exclude any abnormality in glucose homeostasis, according to established criteria (31). In the normal subjects, there was no clinical or familial history of either diabetes or impaired glucose tolerance, and their glucose tolerance was normal.

Table 1. Clinical characteristics of the cirrhotic and normal subjects studied

<table>
<thead>
<tr>
<th>Age, yr</th>
<th>BMI, kg/m²</th>
<th>LBM, kg</th>
<th>% body wt</th>
<th>Histology</th>
<th>Class</th>
<th>Albumin, g/l</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>49</td>
<td>27.8</td>
<td>67</td>
<td>HCV-cirrhosis</td>
<td>B</td>
<td>36</td>
</tr>
<tr>
<td>2</td>
<td>56</td>
<td>28.9</td>
<td>56</td>
<td>HCV-cirrhosis</td>
<td>B</td>
<td>44.2</td>
</tr>
<tr>
<td>3</td>
<td>53</td>
<td>26.3</td>
<td>58</td>
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<td>A</td>
<td>43.5</td>
</tr>
<tr>
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<td>45</td>
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<td>67</td>
<td>HCV-cirrhosis</td>
<td>A</td>
<td>42</td>
</tr>
<tr>
<td>5</td>
<td>38</td>
<td>27.1</td>
<td>63</td>
<td>Cryptogenic cirrhosis</td>
<td>B</td>
<td>48</td>
</tr>
<tr>
<td>6</td>
<td>54</td>
<td>30.1</td>
<td>65</td>
<td>HCV-cirrhosis</td>
<td>A</td>
<td>42.2</td>
</tr>
<tr>
<td>Mean ± SE</td>
<td>49 ± 3</td>
<td>26.9 ± 0.7</td>
<td>61.1 ± 2.4</td>
<td>76.4 ± 1.1</td>
<td>42.7 ± 1.6</td>
<td></td>
</tr>
<tr>
<td>Normal subjects</td>
<td>35 ± 7</td>
<td>23.9 ± 1.3</td>
<td>56.2 ± 0.9</td>
<td>82.0 ± 1.6</td>
<td>42.3 ± 0.9</td>
<td></td>
</tr>
</tbody>
</table>

BMI, body mass index; LBM, lean body mass; Class, discrimination on basis of Child-Pugh classification (5); HCV, hepatitis C virus. *P ≤ 0.05 vs. control subjects.
those attained after a mixed meal ingested as a bolus, at least at its peak value (26). In addition, a continuous oral feeding was shown to stimulate whole body, albumin, and muscle protein synthesis (8, 37).

Analytic measurements. Plasma leucine and KIC concentrations and SA were determined by HPLC (13, 33). Plasma total amino acid concentrations were determined by ion exchange chromatography with a Beckman amino acid analyzer. The $^{14}$CO$_2$ in the expired air was determined as described previously (33, 37). Insulin and glucagon concentrations were measured by radioimmunoassay (37). Plasma glucose was determined using a Beckman Glucose Analyzer 2.

Albumin was isolated from plasma as described previously (13), with an additional acid precipitation to avoid contamination from free amino acids. Hydrolysis was accomplished by adding 4 ml of 4 N HCl to the isolated protein and by incubating the samples in air-tight vials at 110°C for 72 h. The albumin-derived free amino acids were
then further purified through cation-exchange Ag 50 × 8 columns. Leucine SA in the hydrolysate was determined by HPLC (13).

Calculations. The steady-state values of plasma leucine and KIC SA in the last 80 min of the basal, postabsorptive state and in the last 80 min of meal administration (i.e., between 170 and 250 min) were averaged. All calculations were performed using these mean values.

Whole body leucine rate of appearance (R,) was calculated using plasma [14C]KIC SA as precursor pool (32). Leucine oxidation was calculated by dividing the rate of 14CO₂ expiration (in dpm/kg⁻¹·min⁻¹), corrected for 20% fixation in body bicarbonate pool, over plasma [14C]KIC SA. The rate of nonoxidative leucine disposal (NOLD), indicating leucine disposal into whole body protein synthesis, was calculated by subtracting oxidation from total leucine R, (equal to the rate of disappearance at steady state).

Albumin FSR was calculated from the samples taken at −80, −60, −40, −20, and 0 min in the basal state, and at 170, 190, 210, 230, and 250 min after the start of meal administration, by use of a standard precursor-product linear relationship (8, 11, 13)

$$\text{FSR} = \frac{\text{SA}_{t(2)} - \text{SA}_{t(1)}}{t(2) - t(1)} \times \frac{1440 \times 100}{[14C]KIC \text{ SA}}$$

where SAₜ(2) and SAₜ(1) are albumin-bound leucine specific activities (in dpm/mmol) at time points t(2) and t(1), respectively; [14C]KIC SA is steady-state plasma [14C]KIC specific activity (in dpm/mmol); the factor “1,440” is used to convert the data from minutes to 1 day; and “100” is used to convert FSR to a percentage. The term at the numerator of Eq. 1 actually corresponds to the slope of the change in leucine-bound SA vs. time, calculated by use of five time points. The coefficients of linear regression of the slopes were >0.9 in both the basal and the meal periods in each subject of both groups. Thus, as precursor pool for both whole body protein and albumin synthesis, we used plasma KIC SA, which represents a standard approach recently validated also as a surrogate of leucyl-tRNA in liver (1, 32).

Statistical analysis. All data are expressed as means ± SE. The changes vs. basal within each group were analyzed with the two-tailed Student t-test for paired data. Comparison of single sets of data between the two groups (such as the relative differences vs. baseline, expressed as either percentages or deltas) was analyzed using the two-tailed Student t-test for unpaired data. The two-way ANOVA for repeated measures was also employed to compare the meal effects in the two groups simultaneously. A P value <0.05 was considered to be statistically significant.

RESULTS

Albumin FSR and leucine kinetics. In the fasting state, albumin FSR in the cirrhotic patients (CP; 8.5 ± 1.5 %/day) was slightly but insignificantly lower than that of the normal subjects [N; 10.9 ± 1.5 %/day, P = nonsignificant (NS)]. After the meal, albumin FSR did not change in the CP (to 8.8 ± 1.7 %/day, P = NS vs. baseline), whereas it increased in the N by an average of ≈45% (to 15.9 ± 1.9 %/day, P < 0.002 vs. baseline; P < 0.025 by ANOVA between groups). The albumin FSR value in the CP during meal absorption was significantly lower (P = 0.022) than in the N (Fig. 2).

Fasting leucine Rₐ, expressed over kilogram of body weight, was not different between the two groups (2.20 ± 0.24 μmol·kg⁻¹·min⁻¹ in CP; 2.36 ± 0.16 μmol·kg⁻¹·min⁻¹ in N). Similarly, leucine oxidation (0.51 ± 0.08 vs. 0.56 ± 0.07) and NOLD (1.68 ± 0.22 vs. 1.80 ± 0.14, respectively) were not different between CP and N (Fig. 3). Also, no differences were observed when the data were expressed over the estimated LBM (CP: leucine Rₐ, 2.89 ± 0.32 μmol·kg⁻¹·min⁻¹; oxidation, 0.68 ± 0.11; NOLD, 2.21 ± 0.3; N: leucine Rₐ, 2.88 ± 0.18; oxidation, 0.68 ± 0.08; NOLD, 2.20 ± 0.17). After the meal, total leucine Rₐ increased (P < 0.01 or less vs. basal) in both groups (to 2.78 ± 0.27 μmol·kg⁻¹·min⁻¹ in CP; and to 3.29 ± 0.26 μmol·kg⁻¹·min⁻¹ in N; Fig. 3). Although postprandial leucine Rₐ tended to be lower in the patients, this difference was not significant. Leucine oxidation increased (P < 0.0001) to similar values (P = NS between groups) in both patients (to 1.24 ± 0.14 μmol·kg⁻¹·min⁻¹) and normal subjects (to 1.18 ± 0.11). In contrast, NOLD increased only in N (to 2.10 ± 0.19 μmol·kg⁻¹·min⁻¹, P = 0.032 vs. basal), whereas it did not change in CP (to 1.54 ± 0.26 μmol·kg⁻¹·min⁻¹, P > 0.1 vs. basal; P < 0.01 by ANOVA for interaction between groups). The relative change vs. basal (expressed as μmol·kg⁻¹·min⁻¹) in NOLD was significantly greater (P < 0.01) in N (+0.31 ± 0.11 μmol·kg⁻¹·min⁻¹) than in CP (−0.15 ± 0.07 μmol·kg⁻¹·min⁻¹). The same pattern was observed when the data were expressed over LBM (CP: leucine Rₐ, 3.66 ± 0.38 μmol·kg⁻¹·min⁻¹; oxidation, 1.64 ± 0.19; NOLD, 2.02 ± 0.34; N: leucine Rₐ, 3.99 ± 0.27; oxidation, 1.43 ± 0.11; NOLD, 2.56 ± 0.22).

Plasma and albumin-bound SAs. Albumin concentration was similar in CP and N groups (Table 1). Postabsorptive plasma [14C]KIC SA was 2.38 ± 0.16

![Image](http://ajpendo.physiology.org/DownloadedFrom/10.2203/03.2/OnSeptember8,2017)
and 2.45 ± 0.19 dpm/nmol in CP and in N, respectively (Fig. 1). Leucine \([^{14}C]\)SA was 3.09 ± 0.23 and 2.96 ± 0.25 dpm/nmol in CP and N, respectively. No significant differences between groups were found in the KIC-to-leucine SA ratios (CP: 0.83 ± 0.03; N: 0.78 ± 0.04). After the meal, the \([^{14}C]\)KIC SA decreased (P < 0.01 or less vs. baseline) to 1.91 ± 0.12 and to 1.75 ± 0.17 dpm/nmol in CP and N, respectively, and that of \([^{14}C]\)leucine to 2.18 ± 0.18 and to 2.24 ± 0.26 dpm/nmol, respectively. The ratio of \([^{14}C]\)KIC to \([^{14}C]\)leucine SA tended to increase (although insignificantly) in the CP (to 0.88 ± 0.03, P > 0.1), whereas it did not change at all in the N (to 0.79 ± 0.05). The slope of changes of albumin-bound leucine \([^{14}C]\)SA vs. time tended to decrease (although insignificantly) in the CP (from 0.138 ± 0.016 to 0.119 ± 0.025 dpm \(\mu\)mol\(^{-1}\)min\(^{-1}\); P = NS), whereas it did not change in the N (from 0.182 ± 0.029 to 0.191 ± 0.029 dpm \(\mu\)mol\(^{-1}\)min\(^{-1}\)).

Substrate and hormone concentrations. Plasma concentrations of the AA phenylalanine and tyrosine were ~40% greater (P < 0.01) in CP than in N in both the basal and the postprandial states (Table 2). Methionine concentration in the CP was also increased (P < 0.001; fasting state: 25 ± 2 vs. 15 ± 1; postprandial state: 42 ± 4 vs. 31 ± 1). Leucine and KIC concentrations were not different between the two groups (Fig. 1, Table 2), although the sum of the BCAA concentrations was slightly greater (by ~15%, P < 0.05) in CP. The BCAA-NOAA (BCAA/NOAA) ratio in the CP was ~20% lower (P < 0.05) than in N both in the fasted (CP: 3.25 ± 0.26; CS: 3.92 ± 0.2) and the fed states (3.23 ± 0.25 and 3.79 ± 0.18, respectively). The sum of total amino acids was not different between the groups (Table 2).

Plasma glucose and insulin concentrations in the CP were greater than in N (P < 0.01 by ANOVA) both in the fasted and the fed states, whereas glucagon concentration was not different (Table 2). In the CP, plasma insulin concentration increased to an ~100% greater value (P < 0.01) than that in N postprandially (Table 2). Glucagon concentration did not significantly change from basal in either group (Table 2).

**DISCUSSION**

This study shows that ingestion of a mixed meal over 4 h did not acutely stimulate either albumin or whole body protein synthesis in compensated cirrhotic patients, in contrast to normal control subjects.

Several factors may account for these defects. Albumin and body protein synthesis are stimulated physiologically by meal ingestion and/or by hyperaminoacidemia (8, 9, 35, 37), the former also by insulin (8, 9). Both of these stimuli appeared to be ineffective in the cirrhotic patients postprandially. Hyperinsulinemia is
a common finding in cirrhosis, and it may be due to hypersecretion and/or decreased hepatic extraction (27). The defective postprandial albumin synthesis despite hyperinsulinemia in cirrhosis is compatible with an insulin-resistant state involving also the hormone proteosynthetic hepatic effect. Such a resistance, in turn, may be secondary to alterations of liver architecture, circulation, and function, which are characteristic of this disease (6), including a shunt of insulin from the portal vein to the posthepatic circulation. A decreased first-pass splanchnic (liver) extraction of some dietary amino acids, such as phenylalanine (34), and the altered plasma profiles [i.e., the increased AAA concentrations and/or the decreased BCAA/AAA ratio (25)] might also have limited albumin synthesis. Among other possible alterations that are observed in cirrhosis (although not measured in this study), toxic effects by cytokines (2, 38), as well as a decreased insulin-like growth factor I (IGF-I) concentration (12), could also be considered. An interesting hypothesis is that albumin synthesis was already maximally stimulated in the postabsorptive state in the patients, thus preventing a further increase after the meal.

Because albumin concentration was normal in the patients despite the defective postprandial stimulation, a decreased albumin catabolism, helping to maintain a nearly normal balance between synthesis and degradation, cannot be excluded. Albumin fractional catabolism could be determined in vivo with the intravenous injection of iodine-labeled albumin (10). In this study, however, we did not perform such a test, mainly because of limitations in subjects’ exposure to radioactive isotopes. Also, iodine kinetics may not correspond accurately to that of native albumin, rather reflecting a different pattern of albumin distribution between intravascular and extravascular sites, i.e., its transcapillary escape (10), which may itself be altered in cirrhosis (19). On the other hand, by definition, synthesis must equal catabolism at steady state. Thus the decreased postprandial albumin FSR in the cirrhotic patients may also indicate a decreased fractional catabolic rate. As a matter of fact, the complex relationships between albumin synthesis and catabolism need to be further investigated.

Interestingly, a defective stimulation of whole body protein synthesis in cirrhosis was previously reported by our group, which followed the intravenous infusions of the branched-chain-enriched amino acids, glucose, and insulin (36), and was in agreement with the present findings. The defective albumin synthesis here observed could account at least in part for the impaired postprandial whole body protein synthesis. Indeed, in humans, albumin synthesis accounts for ≈30% of the increase of body protein synthesis after a meal (8), and it may act as a carrier of protein-bound amino acids from visceral to peripheral organs, like muscle, postprandially (8, 10). Thus the impaired postprandial albumin synthesis might contribute to the reduced muscle mass often found in cirrhotic patients.

The defects in protein synthesis observed in the patients cannot be explained by their slightly greater body weight and LBM, because in obesity whole body protein synthesis was found to be normally stimulated by hyperaminoacidemia during insulin infusion (16). Furthermore, we found no correlation between either body weight or estimated LBM and albumin FSR in a large series of observations (unpublished observations). In regard to body composition, we acknowledge that neither Hume’s method (14) nor DEXA (17) may be accurate enough to measure LBM in cirrhosis because of a possible shift of body water from intracellular to extracellular space(s) (10, 19), even in the absence of clinically and instrumentally demonstrable edema and ascites. Nevertheless, this uncertainty may affect only the absolute values of leucine kinetics but not albumin FSR or the relative changes vs. basal of both albumin and whole body protein kinetics.

A key issue is that of the validity of the use of KIC SAs as the precursor pool of albumin synthesis also in cirrhosis. Theoretically, a decreased liver protein degradation in cirrhosis could increase the intrahepatic leucine SA locally and lead to an underestimation of albumin FSR, which could be interpreted as decreased synthesis. Only the measurement of intrahepatic leucyl tRNA could address this issue. However, such a measurement is clearly unfeasible in humans, for both ethical and technical reasons. As a matter of fact, limitations of current methods to measure albumin synthesis in vivo both in healthy and diseased states should be kept in mind.

Leucine Rₐ increased postprandially in both groups as a result of meal ingestion. Whether this increase was proportional to the leucine ingested with the meal cannot be established from our data, because no oral amino acid tracer was administered. For the same reason, neither first-pass leucine splanchnic uptake nor leucine Rₐ from endogenous protein breakdown could be accurately determined. However, these issues had been previously investigated by use of combined oral and intravenous isotope administrations (34).

Although either early or late anabolic effects might have been missed in the patients during the 4-h meal administration, these possibilities seem unlikely. Indeed, both whole body protein and albumin kinetics exhibit quite slow patterns (8, 9, 20, 37), i.e., it would be surprising if a brisk stimulation of protein synthesis occurred in cirrhosis early after the start of feeding and before the chosen sampling times. On the other hand, an impaired and/or delayed meal absorption cannot entirely be ruled out in the patients. Nevertheless, the near-elementary composition of the liquid meal, as well as the adequate increase of both amino acid concentrations and leucine oxidation in the patients postprandially, makes this possibility rather unlikely.

The patients’ nearly normal fasting plasma leucine concentration is in agreement with previous (15, 18), although not all, reports (23, 24, 39). These variances might depend on the patients’ weight, body composition, and clinical/nutritional status, as well as on insu-
lin resistance and/or concentration, because BCAA levels are very sensitive to insulin action (9, 28, 35). Thus the patients’ nearly normal BCAA levels may be accounted for by their mild fasting hyperinsulinemia.

In conclusion, a mixed meal ingestion did not acutely stimulate either albumin or whole body protein synthesis in stable cirrhotic patients as opposed to healthy control subjects. The mechanism(s) underlying these abnormalities need to be further investigated.

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REFERENCES


