Calcitonin gene-related peptide receptor antagonist human CGRP-(8–37)

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Calcitonin gene-related peptide receptor antagonist human CGRP-(8–37). Am. J. Physiol. 256 (Endocrinol. Metab. 19): E331-E335, 1989.—From this study, we predicted that the human calcitonin gene-related peptide (hCGRP) fragment hCGRP (8–37) would be a selective antagonist for CGRP receptors but an agonist for calcitonin (CT) receptors. In rat liver plasma membrane, where CGRP receptors predominate and CT appears to act through these receptors, hCGRP-(8–37) dose dependently displaced 125I-[Tyr']rat CGRP binding. However, hCGRP-(8–37) had no effect on adenylate cyclase activity in liver plasma membrane. Furthermore, hCGRP-(8–37) inhibited adenylate cyclase activation induced not only by hCGRP but also by hCT. On the other hand, in LLC-PK1 cells, where calcitonin receptors are abundant and CGRP appears to act through these receptors, the bindings of 125I-[Tyr']rat CGRP and 125I-hCT were both inhibited by hCGRP-(8–37). In contrast to liver membranes, interaction of hCGRP-(8–37) with these receptors led to stimulation of adenosine 3', 5'-cyclic monophosphate (cAMP) production in LLC PK1 cells, and moreover, this fragment did not inhibit the increased production of cAMP induced not only by hCT but also by hCGRP. Thus hCGRP-(8–37) appears to be a useful tool for determining whether the action of CGRP as well as that of CT is mediated via specific CGRP receptors or CT receptors.

 Moreover, recent receptor cross-linking studies followed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) analysis have revealed the existence of specific CGRP receptors, which are distinct from calcitonin receptors (2, 13), with differing molecular sizes in various tissues (3, 4, 7). Because of the structural similarities mentioned above, however, these two peptides have also been shown to cross-react with each other at the levels of their receptors (6, 12, 19, 20). Furthermore, it has also been reported that specific receptors for both calcitonin and CGRP appear to coexist within the same tissue, such as brain (6). As a result, it is extremely difficult to elucidate the direct actions of these peptides on their respective receptors. Previously, however, we have demonstrated that rat liver plasma membrane exclusively possesses CGRP receptors and that calcitonin appears to interact with these receptors (20). On the other hand, Wohlwend et al. (19) have reported that, in LLC-PK1, pig kidney cells, calcitonin receptors predominate and that the stimulation of adenylate cyclase activity by CGRP is probably mediated by these receptors. Therefore, by using both LLC-PK1 cells and rat liver plasma membrane, it should be possible to characterize specific CGRP receptors and calcitonin receptors separately.

 Recently, we have shown that a human CGRP fragment, human CGRP-(8–37), binds to CGRP receptors in rat liver plasma membrane (20). In this study, by examining the effects on not only CGRP receptors in liver plasma membrane but also calcitonin receptors on LLC-PK1 cells, we found that this fragment, human CGRP-(8–37), is a specific antagonist for CGRP receptors but not for calcitonin receptors, raising the possibility of being able to clarify whether the CGRP or the calcitonin receptor is responsible for the action of either of these two peptides under various conditions using this CGRP antagonist.

MATERIALS AND METHODS

Peptides and radioligands. Synthetic [Tyr']rat CGRP and human calcitonin were purchased from Peninsula
Human CGRP and human CGRP-(8–37) were synthesized using an automatic solid-phase synthesizer (430-A Peptide Synthesizer, Applied Biosystems. 125I-[Tyr]rat CGRP (2,000 Ci/mmol) was prepared as previously described (14), and 125I-human calcitonin (2,000 Ci/mmol) was supplied by Amersham (Buckinghamshire, UK). Human calcitonin was radioiodinated by the chloramine-T method, and the methionine-oxidized form of 125I-human calcitonin was removed by reverse-phase high-performance liquid chromatography (HPLC). An adenosine 3',5'-cyclic monophosphate (cAMP) assay kit was obtained from Yamasa Shoyu (Choshi, Japan) (17).

Preparation of rat liver plasma membranes, membrane receptor binding study, and membrane adenylate cyclase assay. Rat liver plasma membranes were prepared from adult male Sprague-Dawley rats by the method of Pilkis et al. (15) with slight modification, as described previously (20). The fraction collected at the 42.5–45% sucrose interface was removed and washed twice with 10 mM N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid (HEPES) buffer (pH 7.4) containing 1 mg/ml bacitracin. The final pellet was resuspended in 10 mM HEPES buffer (pH 7.4) containing 120 mM NaCl, 4.7 mM KCl, 5 mM MgCl2, 1 mM ethylene glycol-bis(β-aminoethyl ether)-N,N,N',N'-tetraacetic acid (EGTA), 1 mg/ml bacitracin, and 5 mg/ml bovine serum albumin (BSA). One-half milliliter of this membrane suspension (200 µg of tissue protein) containing 125I-[Tyr]rat CGRP (25 pM) was incubated at 24°C for 60 min with or without various unlabeled peptides. Bound and free hormone were separated by centrifugation and radioactivity bound to the pellets was counted by an autogamma counter. Nonspecific binding was assessed as the fraction of label that remained bound in the presence of 10⁻⁶ M [Tyr]rat CGRP.

The adenylate cyclase activity in the membranes was determined by following the synthesis of cAMP from nonradioactive ATP, as described by Katada and Ui (8). Membranes (10 µg of protein) were incubated for 10 min at 37°C in 100 µl of tris(hydroxymethyl)aminomethane (Tris) buffer (25 mM, pH 7.6) containing 1 mM EGTA, 1 mg/ml of BSA, 100 mM NaCl, 100 µM GTP, 1 mM ATP, 5 mM MgCl2, 1 mM 3-isobutyl-1-methylxanthine (IBMX), 5 mM phosphocreatine, 50 U/ml of creatine phosphokinase, and 1 mM aprotinin with or without various stimulants. The reactions were terminated by adding 0.33 M ZnSO4 (40 µl) and 0.5 M Na2CO3 (40 µl), and after being left to stand for 1 h at 4°C, samples were centrifuged and cAMP levels in the supernatant were measured by a sensitive radioimmunoassay (RIA) (17).

LLC-PK1 cell culture and study of receptor binding and cAMP production in these cells. LLC-PK1 pig kidney cells (ATCC, CRL 1392) (19) were grown at 37°C in a 1:1 mixture of Ham's F12-Dulbeco's modified Eagle's medium with 10% fetal calf serum (FCS) under humidified 95% O₂-5% CO₂ atmosphere. For the experiments, the cells were washed with phosphate-buffered saline, dis-

FIG. 1. Dose-response inhibition of 125I-[Tyr]rat calcitonin gene-related peptide (CGRP) binding to rat liver plasma membranes by human CGRP (○), human CGRP-(8–37) (■), and human calcitonin (○). Rat liver plasma membranes were incubated for 60 min at 24°C with 25 pM 125I-[Tyr]rat CGRP plus peptides as indicated. Total and specific bindings were 11.4 ± 1.8 and 10.3 ± 1.4% of total radioactivity added, respectively. Data are expressed as percentages of maximal specific binding in absence of unlabeled peptides. Values are means of 6 separate experiments differing by <5%. Each experiment was done in duplicate.

FIG. 2. Effect of human calcitonin gene-related peptide (CGRP) (8–37) on human CGRP-stimulated adenylate cyclase activation in rat liver plasma membrane. Rat liver plasma membranes were incubated either with increasing concentrations of human CGRP (■), human CGRP-(8–37) (□), human calcitonin (○), or human calcitonin plus human CGRP-(8–37) (○–○) for 10 min at 37°C. Each value represents mean ± SE of 5 separate experiments. Each experiment was done in duplicate. Statistical analysis was done by analysis of variance in conjunction with Duncan's new multiple range test. *P < 0.01 vs. control value.
mixture, and washing the supernatant three times with ice-cold water-saturated diethyl ether. cAMP levels were determined by a specific RIA (17).

**RESULTS**

The specific binding of $^{125}$I-[Tyr$^9$]-rat CGRP to rat liver plasma membrane was displaced by human CGRP, human CGRP-(8–37) and by human calcitonin in a dose-dependent manner with relative affinities as follows: human CGRP > human CGRP-(8–37) > human calcitonin (Fig. 1). Of these three peptides, both human CGRP and human calcitonin significantly stimulated adenylate cyclase activation in a dose-dependent fashion in rat liver membrane, but human CGRP was clearly more potent than human calcitonin (Fig. 2). Furthermore, the doses of these peptides necessary for producing a half-maximal effect on adenylate cyclase activation (human CGRP, 21.5 pM; human calcitonin, 0.46 μM) were comparable to those producing half-maximal inhibition of label binding to the membrane (32.1 pM and 1.32 μM, respectively). In contrast, human CGRP-(8–37) did not have any effect on adenylate cyclase activity up to $10^{-5}$ M, and moreover, in the presence of $10^{-5}$ M human CGRP-(8–37), the dose-response curve for adenylate cyclase activation induced not only by human CGRP but also by human calcitonin was shifted to the right (Fig. 2). Indeed, the inhibitory effect of human CGRP-(8–37) on human CGRP-induced as well as human calcitonin-induced adenylate cyclase activation was dose-dependent (Fig. 3). However, human CGRP-(8–37) ($10^{-5}$ M) had no effect on either epinephrine ($10^{-6}$ M)- or glucagon ($10^{-7}$ M)-induced activation of adenylate cyclase (Table 1).

On the other hand, all of human CGRP, human CGRP-(8–37) and human calcitonin displaced the specific binding of $^{125}$I-[Tyr$^9$]-rat CGRP as well as that of $^{125}$I-human calcitonin to LLC-PK$_1$ cells in a dose-dependent fashion (Fig. 4). The relative potencies of these peptides for the displacement of $^{125}$I-[Tyr$^9$]rat CGRP and $^{125}$I-human calcitonin were similar, and in contrast to the result obtained with rat liver plasma membrane, human calcitonin was most potent followed by human CGRP and then by human CGRP-(8–37) (4.5 M) on adenylate cyclase activity induced by epinephrine ($10^{-6}$ M)- or glucagon ($10^{-7}$ M)-induced activation of adenylate cyclase (Table 1).

In striking contrast to the data obtained with rat liver plasma membrane, it was found that in addition to human CGRP and human calcitonin, human CGRP-(8–37) also evoked a significant and dose-dependent increase in cAMP production in LLC-PK$_1$ cells (Fig. 5). Furthermore, human CGRP-(8–37) ($10^{-5}$ M) had no effect on either human CGRP- or human calcitonin-induced adenylate cyclase activation (Fig. 5). The relative potencies of these peptides in stimulating cAMP production were similar to those in displacing the label bindings from LLC-PK$_1$ cells.

**DISCUSSION**

In the previous report, we identified specific binding sites for CGRP in rat liver plasma membrane and showed that various kinds of calcitonin interacted with these CGRP receptors (20). The present study demonstrated that human CGRP as well as human calcitonin induced...
A SPECIFIC CGRP ANTAGONIST

A dose dependent activation of adenylate cyclase in rat liver plasma membrane. Thus, it appears that receptors for CGRP in rat liver plasma membrane are linked to the adenylate cyclase-cAMP system. However, in contrast to the effects of human CGRP and human calcitonin and despite its ability to bind to CGRP receptors, the present data clearly demonstrated that a human CGRP fragment, human CGRP-(8-37), had no effect on adenylate cyclase activity in rat liver plasma membrane. On the other hand, LLC-PK1 pig kidney cells have been reported to possess specific receptors for calcitonin, and CGRP appears to exert its action on these cells through the calcitonin receptors (19). In the present experiment, human calcitonin was more potent than human CGRP not only in displacing the specific binding of 125I-human calcitonin or [Tyr']rat CGRP but also in activating adenylate cyclase in LLC-PK1 cells, thus confirming previous reports (19). We, therefore, examined whether this CGRP receptor antagonist, human CGRP-(8-37), also antagonizes the actions of human CGRP and human calcitonin on calcitonin receptors in LLC-PK1 cells, and it was found that this fragment not only stimulated adenylate cyclase activation but also had no inhibitory effect on adenylate cyclase activation induced by either human CGRP or human calcitonin. Thus it is evident that this CGRP receptor antagonist, human CGRP-(8-37), acts as an agonist for calcitonin receptors in LLC-PK1 cells.

In a previous report of a study using rat liver plasma membranes, we suggested that the COOH-terminal portion of CGRP is essential for binding to its receptors, since an NH2-terminal fragment of human CGRP could not bind to CGRP receptors, whereas COOH-terminal fragments were able to displace label binding (20). The present data add new evidence to support the fact that in spite of their ability to bind to CGRP receptors, COOH-terminal fragments of CGRP lacking a ring structure with a disulfide bond may not be sufficient to induce a subsequent intracellular signal transduction event, i.e., adenylate cyclase activation. Of particular interest in this study, however, was that this COOH-terminal fragment of human CGRP was able to promote adenylate cyclase activation, probably through specific calcitonin receptors in LLC-PK1 cells. This is rather surprising, since this COOH-terminal portion of human CGRP has only slight amino acid sequence homology with human calcitonin. Goltzman (5), on the other hand, has reported that various modifications of the NH2-terminal portion...
of the calcitonin molecule including the disulfide bridge did not result in substantial reduction of its capacity to inhibit $^{125}\text{I}$-salmon calcitonin binding and to stimulate adenylate cyclase in rabbit renal membranes. In agreement with his data, we found that the potency of human CGRP-(8-37), a COOH-terminal fragment of human CGRP, both for the binding and the stimulation of adenylate cyclase in LLC-PK$_2$ cells was not substantially different from that of human CGRP. Taken together, the COOH terminal portion of the calcitonin molecule appears to be more important than the NH$_2$-terminal portion both in binding to its receptor and in stimulating adenylate cyclase, although the calcitonin receptor may appear to exist for both CGRP and calcitonin (6) and these peptides are known to cross-react with each other on their receptors (12, 19, 20). Thus, although it has been reported that intracerebroventricular administration of CGRP and calcitonin often exerts similar effects such as inhibition of gastric acid secretion (10) and reduction of food intake (9), it is still uncertain whether the actions of these peptides are mediated through calcitonin receptors or CGRP receptors. It is, therefore, tempting to predict that this newly discovered CGRP antagonist, human CGRP-(8-37), could be a useful tool for precise characterization of the actions of CGRP and calcitonin at the levels of their respective receptors.

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REFERENCES