Calcitonin gene-related peptide receptor antagonist human CGRP-(8–37)

TSUTOMU CHIBA, AKINORI YAMAGUCHI, TOSHIYUKI YAMATANI, AKIRA NAKAMURA, TOMOYUKI MORISHITA, TETSUYA INUI, MASAAKI FUKASE, TOSHIHARU NODA, AND TAKUO FUJITA
Third Department of Internal Medicine, Kobe University School of Medicine, Kobe 650; and Toyo Jozo Research Laboratory, Shizuoka 632-1, Japan

Calcitonin gene-related peptide receptor antagonist human CGRP-(8–37). Am. J. Physiol. 256 (Endocrinol. Metab. 19): E331-E335, 1989.-From this study, we predicted that the human calcitonin gene-related peptide (hCGRP) fragment hCGRP-(8–37) would be a selective antagonist for CGRP receptors but an agonist for calcitonin (CT) receptors. In rat liver plasma membrane, where CGRP receptors predominate and CT appears to act through these receptors, hCGRP-(8–37) dose dependently displaced 125I-[Tyr5]rat CGRP binding. However, hCGRP-(8–37) had no effect on adenylate cyclase activity in liver plasma membrane. Furthermore, hCGRP-(8–37) inhibited adenylate cyclase activation induced not only by hCGRP but also by hCT. On the other hand, in LLC-PK1 cells, where calcitonin receptors are abundant and CGRP appears to act via these receptors, the bindings of 125I-[Tyr5]rat CGRP and 125I-hCT were both inhibited by hCGRP-(8–37). In contrast to liver membranes, interaction of hCGRP-(8–37) with these receptors led to stimulation of adenosine 3’, 5’-cyclic monophosphate (cAMP) production in LLC PK1 cells, and moreover, this fragment did not inhibit the increased production of cAMP induced not only by hCT but also by hCGRP. Thus hCGRP-(8–37) appears to be a useful tool for determining whether the action of CGRP as well as that of CT is mediated via specific CGRP receptors or CT receptors.

LLC-PK1 cells; calcitonin gene

ALTERNATIVE TISSUE specific processing of initial mRNA transcripts from the calcitonin gene generates two distinct peptides: calcitonin and calcitonin gene-related peptide (CGRP) (1, 16). Although these two peptides have little sequence homology, calcitonin and CGRP possess similar structural characteristics, i.e., similar sizes, a disulfide bridge at the NH2-terminal portion, and an amidated carboxy-terminal end. On the other hand, previous pharmacological studies have demonstrated the presence of distinct receptors for calcitonin and for CGRP in various tissues and cells (5, 7, 11, 12, 19, 20). Moreover, recent receptor cross-linking studies followed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) analysis have revealed the existence of specific CGRP receptors, which are distinct from calcitonin receptors (2, 13), with differing molecular sizes in various tissues (3, 4, 7). Because of the structural similarities mentioned above, however, these two peptides have also been shown to cross-react with each other at the levels of their receptors (6, 12, 19, 20). Furthermore, it has also been reported that specific receptors for both calcitonin and CGRP appear to coexist within the same tissue, such as brain (6). As a result, it is extremely difficult to elucidate the direct actions of these peptides on their respective receptors. Previously, however, we have demonstrated that rat liver plasma membrane exclusively possesses CGRP receptors and that calcitonin appears to interact with these receptors (20). On the other hand, Wohlwend et al. (19) have reported that, in LLC-PK1 pig kidney cells, calcitonin receptors predominate and that the stimulation of adenylate cyclase activity by CGRP is probably mediated by these receptors. Therefore, by using both LLC-PK1 cells and rat liver plasma membrane, it should be possible to characterize specific CGRP receptors and calcitonin receptors separately.

Recently, we have shown that a human CGRP fragment, human CGRP-(8–37), binds to CGRP receptors in rat liver plasma membrane (20). In this study, by examining the effects on not only CGRP receptors in liver plasma membrane but also calcitonin receptors on LLC-PK1 cells, we found that this fragment, human CGRP-(8–37), is a specific antagonist for CGRP receptors but not for calcitonin receptors, raising the possibility of being able to clarify whether the CGRP or the calcitonin receptor is responsible for the action of either of these two peptides under various conditions using this CGRP antagonist.

MATERIALS AND METHODS

Peptides and radioligands. Synthetic [Tyr5]rat CGRP and human calcitonin were purchased from Peninsula

0193-1849/89 $1.50 Copyright © 1989 the American Physiological Society
(Helmont, CA). Human CGRP and human CGRP-(8—37) were synthesized using an automatic solid-phase synthesizer (430-A Peptide Synthesizer, Applied Biosystems. $^{125}$I-[Tyr]$^{	ext{a}}$rat CGRP (2,000 Ci/mmol) was prepared as previously described (14), and $^{125}$I-human calcitonin (2,000 Ci/mmol) was supplied by Amersham (Buckinghamshire, UK). Human calcitonin was radioidinated by the chloramine-T method, and the methionine-oxidized form of $^{125}$I-human calcitonin was removed by reverse-phase high-performance liquid chromatography (HPLC). An adenosine 3',5'-cyclic monophosphate (cAMP) assay kit was obtained from Yamasa Shoyu (Choshi, Japan) (17).

Preparation of rat liver plasma membranes, membrane receptor binding study, and membrane adenylate cyclase assay. Rat liver plasma membranes were prepared from adult male Sprague-Dawley rats by the method of Pilikis et al. (15) with slight modification, as described previously (20). The fraction collected at the 42.5—48.2% sucrose interface was removed and washed twice with 10 mM N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid (HEPES) buffer (pH 7.4) containing 1 mg/ml bactracin. The final pellet was resuspended in 10 mM HEPES buffer (pH 7.4) containing 120 mM NaCl, 4.7 mM KCl, 5 mM MgCl$_2$, 1 mM ethylene glycol-bis(β-aminoethyl ether)-N,N,N',N'-tetraacetic acid (EGTA), 1 mg/ml bactracin, and 5 mg/ml bovine serum albumin (BSA). One-half milliliter of this membrane suspension (200 μg of tissue protein) containing $^{125}$I-[Tyr]$^{	ext{a}}$rat CGRP (25 pM) was incubated at 24°C for 60 min with or without various unlabeled peptides. Bound and free hormone were separated by centrifugation and radioactivity bound to the pellets was counted by an autogamma counter. Nonspecific binding was assessed as the fraction of label that remained bound in the presence of 10$^{-6}$ M [Tyr]$^{	ext{a}}$rat CGRP.

The adenylate cyclase activity in the membranes was determined by following the synthesis of cAMP from nonradioactive ATP, as described by Katada and Ui (8). Membranes (10 μg of protein) were incubated for 10 min at 37°C in 100 μl of tris(hydroxymethyl)aminomethane (Tris) buffer (25 mM, pH 7.6) containing 1 mM EGTA, 1 mg/ml of BSA, 100 mM NaCl, 100 μM GTP, 1 mM ATP, 5 mM MgCl$_2$, 1 mM 3-isobutyl-1-methylxanthine (IBMX), 5 mM phosphocreatine, 50 μM of creatine phosphokinase, and 1 mM aprotinin with or without various stimulants. The reactions were terminated by adding 0.33 M ZnSO$_4$ (40 μl) and 0.5 M NaCO$_3$ (40 μl), and after being left to stand for 1 h at 4°C, samples were centrifuged and CAMP levels in the supernatant were measured by a sensitive radioimmunoassay (RIA) (17). The adenylate cyclase activity was expressed as picomoles of cAMP synthesized by 1 mg of membrane protein during 1 min of incubation.

LLC-PK$_1$ cell culture and study of receptor binding and cAMP production in these cells. LLC-PK$_1$ pig kidney cells (ATCC, CRL 1392) (19) were grown at 37°C in a 1:1 mixture of Ham’s F12-Dulbeco’s modified Eagle’s medium with 10% fetal calf serum (FCS) under humidified 95% O$_2$-5% CO$_2$ atmosphere. For the experiments, the cells were washed with phosphate-buffered saline, disperse with 0.5% trypsin and 1 mM EDTA, and preincubated in HEPES-buffered Krebs-Ringer bicarbonate buffer with 1 mg/ml BSA and 2.5 mM glucose (HKRBG), pH 7.4, for 1 h. The binding study was carried out by...
incubating the cells (2 × 10^6/0.5 ml) with ^125^I-[^Tyr^9]rat CGRP (50 pM) or ^125^I-human calcitonin (25 pM) in the presence or absence of various unlabeled peptides at 37°C for 60 min. Cells were separated by centrifugation, and the radioactivity bound to the cells was counted. Specific binding was defined as total binding minus nonspecific binding in the presence of 10^-6 M unlabeled human calcitonin.

In the cAMP production study, the cells (10^6) were incubated in 1 ml HKRBG with or without various agents in the presence of IBMX (10^-4 M) at 37°C. After 15 min, the incubations were terminated by adding 1 ml of ice-cold 12% trichloroacetic acid, centrifuging the mixture, and washing the supernatant three times with water-saturated diethylether. cAMP levels were determined by a specific RIA (17).

**RESULTS**

The specific binding of ^125^I-[^Tyr^9]-rat CGRP to rat liver plasma membrane was displaced by human CGRP, human CGRP-(8-37) and by human calcitonin in a dose-dependent manner with relative affinities as follows: human CGRP > human CGRP-(8-37) > human calcitonin (Fig. 1). Of these three peptides, both human CGRP and human calcitonin significantly stimulated adenylate cyclase activation in a dose-dependent fashion in rat liver membrane, but human CGRP was clearly more potent than human calcitonin (Fig. 2). Furthermore, the doses of these peptides necessary for producing a half-maximal effect on adenylate cyclase activation (human CGRP, 21.5 pM; human calcitonin, 0.46 pM) were comparable to those producing half-maximal inhibition of label binding to the membrane (32.1 pM and 1.32 pM, respectively). In contrast, human CGRP (8-37) did not have any effect on adenylate cyclase activity up to 10^-5 M, and moreover, in the presence of 10^-5 M human CGRP-(8-37), the dose-response curve for adenylate cyclase activation induced not only by human CGRP but also by human calcitonin was shifted to the right (Fig. 2). Indeed, the inhibitory effect of human CGRP-(8-37) on human CGRP-induced as well as human calcitonin-induced adenylate cyclase activation was dose-dependent (Fig. 3). However, human CGRP-(8-37) (10^-5 M) had no effect on either epinephrine (10^-6 M)- or glucagon (10^-7 M)-induced activation of adenylate cyclase (Table 1).

On the other hand, all of human CGRP, human CGRP-(8-37) and human calcitonin displaced the specific binding of ^125^I-[^Tyr^9]-rat CGRP as well as that of ^125^I-human calcitonin to LLC-PK_1_ cells in a dose-dependent fashion (Fig. 4). The relative potencies of these peptides for the displacement of ^125^I-[^Tyr^9]rat CGRP and ^125^I-human calcitonin were similar, and in contrast to the result obtained with rat liver plasma membrane, human calcitonin was most potent followed by human CGRP and then by human CGRP-(8-37) (10^-5 M) had no effect on either epinephrine (10^-6 M)- or glucagon (10^-7 M)-induced activation of adenylate cyclase (Table 1). In striking contrast to the data obtained with rat liver plasma membrane, it was found that in addition to human CGRP and human calcitonin, human CGRP-(8-37) also evoked a significant and dose-dependent increase in cAMP production in LLC-PK_1_ cells (Fig. 5). Furthermore, human CGRP-(8-37) (10^-5 M) had no effect on either human CGRP- or human calcitonin-induced adenylate cyclase activation (Fig. 5). The relative potencies of these peptides in stimulating cAMP production were similar to those in displacing the label bindings from LLC-PK_1_ cells.

**DISCUSSION**

In the previous report, we identified specific binding sites for CGRP in rat liver plasma membrane and showed that various kinds of calcitonin interacted with these CGRP receptors (20). The present study demonstrated that human CGRP as well as human calcitonin induced...
A SPECIFIC CGRP ANTAGONIST

A dose dependent activation of adenylate cyclase in rat liver plasma membrane. Thus, it appears that receptors for CGRP in rat liver plasma membrane are linked to the adenylate cyclase-cAMP system. However, in contrast to the effects of human CGRP and human calcitonin and despite its ability to bind to CGRP receptors, the present data clearly demonstrated that a human CGRP fragment, human CGRP-(8-37), had no effect on adenylate cyclase activity in rat liver plasma membrane and that, moreover, it significantly inhibited the activation of adenylate cyclase induced not only by human CGRP but also by human calcitonin. Thus it is strongly suggested that human CGRP-(8-37) acts as an antagonist for the peptides that bind to CGRP receptors in rat liver plasma membrane.

On the other hand, LLC-PK₁ pig kidney cells have been reported to possess specific receptors for calcitonin, and CGRP appears to exert its action on these cells through the calcitonin receptors (19). In the present experiment, human calcitonin was more potent than human CGRP not only in displacing the specific bindings of ¹²⁵I-[Tyr']rat CGRP and ¹²⁵I-human calcitonin but also in activating adenylate cyclase in LLC-PK₁ cells, thus confirming previous reports (19). We, therefore, examined whether this CGRP receptor antagonist, human CGRP-(8-37), also antagonizes the actions of human CGRP and human calcitonin on calcitonin receptors in LLC-PK₁ cells, and it was found that this fragment not only stimulated adenylate cyclase activation but also had no inhibitory effect on adenylate cyclase activation induced by either human CGRP or human calcitonin. Thus it is evident that this CGRP receptor antagonist, human CGRP-(8-37), acts as an agonist for calcitonin receptors in LLC-PK₁ cells.

In a previous report of a study using rat liver plasma membranes, we suggested that the COOH-terminal portion of CGRP is essential for binding to its receptors, since an NH₂-terminal fragment of human CGRP could not bind to CGRP receptors, whereas COOH-terminal fragments were able to displace label binding (20). The present data add new evidence to support the fact that in spite of their ability to bind to CGRP receptors, COOH-terminal fragments of CGRP lacking a ring structure with a disulfide bond may not be sufficient to induce a subsequent intracellular signal transduction event, i.e., adenylate cyclase activation. Of particular interest in this study, however, was that this COOH-terminal fragment of human CGRP was able to promote adenylate cyclase activation, probably through specific calcitonin receptors in LLC-PK₁ cells. This is rather surprising, since this COOH-terminal portion of human CGRP has only slight amino acid sequence homology with human calcitonin. Goltzman (5), on the other hand, has reported that various modifications of the NH₂-terminal portion
REFERENCES


